

Norfolk Island Airport

27 February 2026

PFAS Ongoing Monitoring Plan - Year 4 Monitoring Report





Document Information

PFAS Ongoing Monitoring Plan - Year 4 Monitoring Report, Norfolk Island Airport

Prepared by:

Senversa Pty Ltd

ABN: 89 132 231 380

Level 6, 15 William Street, Melbourne, VIC 3000

tel:+61 3 9606 0070

www.senversa.com.au

Prepared for:

Department of Infrastructure, Transport, Regional Development, Communication and the Arts

111 Alinga Street

Canberra, ACT, 2601

| Revision | Date | Author | Reviewed | Approved | Detail |
|----------|------------|----------------|------------------|------------------|-------------------------|
| 0 | 22/01/2026 | Michelle Agnew | Katie Richardson | Katie Richardson | Draft for client review |
| 1 | 27/02/2026 | Michelle Agnew | Katie Richardson | Katie Richardson | Final |

Project Manager: Michelle Agnew

Project Director: Katie Richardson

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Executive Summary

Senversa Pty Ltd (Senversa) has been engaged by the Department of Infrastructure, Transport, Regional Development, Communications, Sport and the Arts (DITRDCSA), to implement the fourth year of the Norfolk Island Airport perfluoroalkyl and polyfluoroalkyl substances (PFAS) Ongoing Monitoring Plan (OMP). The PFAS OMP (Senversa, 2021d) establishes the ongoing monitoring actions required to assess the nature and extent of PFAS at the Norfolk Island Airport (the site) and surrounding catchments. The site location and surrounding catchments are indicated on **Figure 1**.

Scope and Objectives

The objectives of the PFAS OMP Year 4 event were to assess:

- Trends in PFAS concentrations in the environment.
- The effectiveness of the selected management options in managing current risks.
- Whether changing conditions exist which may result in changes in the risk profile (and therefore changes to the required management actions).

The monitoring event was undertaken in November 2025 and comprised PFAS analysis of:

- Tanks and taps on-site (Airport terminal, maintenance shed, fire station, and Airport Bore).
- Tanks and taps on council sites (works depot).
- Surface water along Mission Creek, Watermill Creek and the end of Cascade Creek and Headstone Creek.
- Private bores along Mission Creek.
- Additional locations, not in the OMP, to address data gaps. Broadly:
 - Drinking water points to confirm concentrations remain below the revised guideline levels.
 - Soil, produce and dam biota sampling following revised residential soil guideline levels.
 - Soil samples to further delineate PFAS source areas.

Summary of Results

The following key results have been reported:

- Airport Bore and the upper Mission Creek catchment surface water PFAS concentrations generally decreased between 2020 and November 2025, representing a statistically significant downwards trend in the Airport Bore, WWII Dam and MC_OMP01.
- PFAS concentrations have been variable over time in both the remaining Mission Creek surface water locations and Watermill Creek. Approximately half of the sampled locations reported PFAS concentrations marginally higher in 2025 than 2024. Most remained within the historical range.
- Concentrations in surface water generally decreased with increased distance from source areas.
- PFAS were detected in home grown produce grown in PFAS-impacted soil and Duck Dam sediments, requiring further assessment.
- Concentrations in water carting water were below the adopted assessment criteria. However, detections of PFAS for which guidelines are unavailable require further assessment.

Assessment of Management Actions

PFAS were below the Upper Trigger Values (UTVs) at all sample locations. This indicates that the selected management options are considered appropriate for the purpose of managing current risks identified in the Human Health and Ecological Risk Assessment (HHERA) (Senversa, 2021c). A number of point of use sample locations reported PFAS concentrations below the Lower Trigger Values (LTVs). This indicates that a reduction in monitoring and/or management may be warranted. This will be considered in the revised PMP.



The following risk pathways identified in the HHERA continue to require management, detailed in the draft PFAS Management Plan (PMP) (Senversa, 2021e):

1. Home consumption or public consumption of cattle, chicken eggs or other animal products where the animal drinks water sourced from Mission Creek.
2. Use of surface water from the Mission Creek Catchment for irrigation, drinking water or domestic uses. Use of groundwater for drinking water or domestic use from the Lower Mission Creek Catchment.
3. Use of groundwater from or nearby the airport or nearby surrounding area for any extractive use.
4. Drinking or washing water at public facilities formerly supplied by the Airport Bore.
5. Use of surface water or groundwater for drinking water or domestic use from the Upper Watermill Creek Catchment.
6. Exposures to freshwater aquatic ecosystems.

In addition, the following changes in management options and assessment should be considered:

- Stock watering: PFAS concentrations during this monitoring event remained above those measured in 2020-2021 sampling, following which it was assessed that further assessment/management was warranted. Further assessment and/or management of cattle access to PFAS impacted water sources requires consideration. Alternatively, measures to manage human exposure (e.g. livestock product consumption advice) could be considered.
- Airport Bore water use: The POET filtration system has been installed on the Airport Bore. While there is low exposure potential for the known uses of this treated water, continued testing of the treated water will be required to assess the ongoing effectiveness of the POET filtration system and whether further management is warranted. Suitable concentration threshold limits will require assessment prior to utilisation of this treated water for other purposes.
- Bore water carting: Further assessment of the quality of water carting water sources used for potable water supply is recommended to confirm detected PFAS concentrations and facilitate a more detailed assessment including consideration of PFAS compounds for which no DWG are available. It is noted that no unacceptable risk has been identified to-date.
- Home produce growing: Where PFAS was detected in home grown produce, further assessment is warranted which considers the nature of the produce in which exceedances were identified and likely consumption rates.
- Biota and sediment re-use: Based on a preliminary assessment, the re-use of biota from Duck Dam, e.g. for compost, is considered to pose a low and acceptable risk. The re-use of sediment should be avoided or appropriately managed to avoid increasing the level of risk on less-impacted sites.

The figure below summarises water uses for which risks are assessed to be low and acceptable, and water uses for which further assessment and/or management is required. The assessment is based on currently measured PFAS concentrations, known current uses of water in each catchment, and the current PFAS regulatory framework.



| Water source | Risk Profile via Current Identified Exposure Pathways | | | | |
|--|---|--------------------|------------------------|----------------------|-----------------------------|
| | Drinking Water | Stock watering | Produce irrigation | Chicken Watering | Freshwater Ecosystem Health |
| Mission Creek | | | | | |
| Lower Mission Creek Catchment Groundwater ¹ | | ✓ | ✓ | ✓ | NA |
| Airport Bore (untreated) & Upper Mission Creek Catchment Groundwater | | | | | NA |
| Watermill Creek | | ✓ | ✓ | ✓ | |
| Cascade Creek | | ✓ | ✓ | ✓ | |
| Headstone Creek | NM | ✓ | ✓ | ✓ | NM |
| Bore and tank water (outside Mission Creek Catchment) | ✓ | ✓ | ✓ | ✓ | NA |

Key:

| | |
|----|---|
| | HHERA identified risks; further assessment and/or management required |
| | HHERA identified risks; management required |
| | HHERA identified risks; no current pathway identified |
| ✓ | Risks low and acceptable |
| NM | Not measured. PFAS not analysed at laboratory limit of reporting protective of this water use |
| NA | Not applicable, no complete exposure pathway |

Notes: 1

¹ At assessed groundwater bores beyond Douglas Drive.



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List of Acronyms

| Acronym | Definition |
|-----------------|---|
| ADWG | Australian Drinking Water Guidelines |
| AFFF | Aqueous film forming foam |
| ALS | Australian Laboratory Services |
| BoM | Bureau of Meteorology |
| CSIRO | Commonwealth Scientific and Industrial Research Organisation |
| DITRDCSA | Department of Infrastructure, Transport, Regional Development, Communications, Sport and the Arts |
| DO | Dissolved oxygen |
| DQI | Data quality indicator |
| DQO | Data quality objective |
| DSI | Detailed Site Investigation |
| DWG | Drinking Water Guidelines |
| EC | Electrical conductivity |
| EMNI | Emergency Management Norfolk Island |
| FSANZ | Food Safety Australia and New Zealand |
| FTS | Fluorotelomer sulfonic acid |
| ha | Hectare |
| HBGV | Health-based guideline value |
| HEPA | Heads of Environment Protection Authority |
| HHERA | Human health and ecological risk assessment |
| HIL | Health investigation level |
| km | Kilometre |
| LOR | Limit of reporting |
| LTV | Lower trigger value |
| mg/kg | Milligrams per kilogram |

| Acronym | Definition |
|--------------|--|
| mV | Millivolts |
| n | Number of samples |
| NA | Not applicable |
| NATA | National Association of Testing Authorities |
| NEMP | National Environmental Management Plan |
| NHMRC | National Health and Medical Research Council |
| NIRC | Norfolk Island Regional Council |
| NM | Not measured |
| OMP | Ongoing monitoring plan |
| PFAS | Per- and poly- fluoroalkyl substances |
| PFBS | Perfluorobutanesulfonic acid |
| PFHxS | Perfluorohexane sulfonate |
| PFOA | Perfluorooctanoic acid |
| PFOS | Perfluorooctane sulfonate |
| PMP | PFAS management plan |
| POET | Point of entry treatment |
| PS | PFAS source |
| PSI | Preliminary site investigation |
| QAQC | Quality assurance and quality control |
| SAQP | Sampling and analysis quality plan |
| µg/L | Micrograms per litre |
| µS/cm | Micro siemens per centimetre |
| UTV | Upper trigger value |



1.0 Introduction and Objectives

Senversa Pty Ltd (Senversa) has been engaged by the Department of Infrastructure, Transport, Regional Development, Communications, Sport and the Arts (DITRDCA) to implement the Norfolk Island Airport perfluoroalkyl and polyfluoroalkyl substances (PFAS) Ongoing Monitoring Plan (OMP). The OMP (Senversa, 2021d) established the ongoing monitoring actions required to assess the nature and extent of PFAS at the Norfolk Island Airport (the site) and surrounding catchments to ensure suitable management actions can be employed. The site location and surrounding catchments are indicated on **Figure 1**.

This report details the results of the PFAS OMP Year 4 monitoring event, undertaken in November 2025.

1.1 Background

Norfolk Island Airport PFAS investigations were initiated after a CSIRO-led¹ assessment of water resources identified elevated levels of PFAS in the Mission Creek water catchment in December 2019². In January 2020, Senversa commenced a Preliminary Site Investigation (PSI) which found that legacy aqueous film-forming foam (AFFF) containing PFAS was used on Norfolk Island from the early 1980s until 2015 to suppress liquid fuel fires and for fire training activities, and confirmed the presence of PFAS in the Mission Creek catchment, together with some other areas of the island (at lower concentrations). These findings were confirmed in a Detailed Site Investigation (DSI) and potentially unacceptable risks were quantitatively assessed in a Human Health and Ecological Risk Assessment (HHERA).

Based on the results of the DSI and the HHERA, risks were assessed as low and acceptable for many of the ways in which people might be exposed to PFAS in the environment, including drinking water. Due to the presence of PFAS in the environment, a draft PFAS Management Plan (PMP) was prepared. The strategy includes actions to manage PFAS sources on the Airport (aimed at reducing the migration of PFAS from the Airport over the longer term) and specific management actions to manage people's exposure to PFAS, including:

- Managing the use of water from Mission Creek for watering cattle or chickens.
- Continued management of water use for drinking water / domestic use more broadly.

The risks to people who might be exposed to PFAS through other uses of water was assessed to be low and these pathways were not included in the PMP.

Potentially elevated risks to ecosystems were identified. However, management options for ecosystems risks are limited. Specifically, remediation at the exposure point (e.g. within creek beds) is unlikely to be feasible or practicable, and unlike for human health exposure pathways, advisory notices to reduce exposure are not an option. As such, management options for ecosystems pathways are restricted to source management actions (which are covered in the PMP), and ecosystems risks are not otherwise the focus of the PMP.

The PFAS OMP was developed to support the implementation of the PMP by assessing changing conditions on-island to determine if the current management actions remain appropriate to manage risks. A Sampling and Analysis Quality Plan (SAQP) was prepared to guide the field works proposed to be undertaken during completion of the PFAS OMP.

¹ Commonwealth Scientific and Industrial Research Organisation

² Commonwealth Scientific and Industrial Research Organisation (2020). *Norfolk Island Water Resource Assessment Hydrology Report_A summary report from the CSIRO Norfolk Island Water Resource Assessment, CSIRO, Australia.*



Key reports are listed below:

- Senversa, 2021a. *Preliminary Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*, revision 2, dated 3 February 2021.
- Senversa, 2021b. *Detailed Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*, revision 5, dated 12 November 2021.
- Senversa, 2021c. *Human Health and Ecological Risk Assessment (PFAS), Norfolk Island Airport*, revision 3, dated 12 November 2021.
- Senversa 2021d. *Ongoing Monitoring Plan, Norfolk Island Airport*, revision 0, dated 24 November 2021.
- Senversa 2021e. *PFAS Management Plan, Norfolk Island Airport*, revision 1, dated 10 December 2021.
- Senversa, 2022a. *PFAS Sampling and Analysis Quality Plan – Year 1 Ongoing Monitoring*, revision 0, dated 3 May 2022.
- Senversa 2022b. *PFAS Ongoing Monitoring Plan - Year 1 Monitoring Report, Norfolk Island Airport*, revision 1, dated 14 October 2022.
- Senversa 2023. *PFAS Ongoing Monitoring Plan – Year 2 Monitoring Report, Norfolk Island Airport*, revision 1, dated 11 October 2023.
- Senversa 2024. *PFAS Ongoing Monitoring Plan – Year 3 Monitoring Report, Norfolk Island Airport*, revision 1, dated 16 October 2024.

1.2 Objectives

The overall objective of the PFAS OMP is to establish the ongoing monitoring required to assess:

- Trends in PFAS concentrations in the environment.
- The effectiveness of the selected management options in managing current risks.
- Whether changing conditions exist which may result in changes in the risk profile (and therefore changes to the required management actions).

The objective of the Year 4 monitoring event was to meet the requirements of the PFAS OMP for the fourth annual monitoring event and to assess temporal variations in PFAS concentrations.

Information from the monitoring program will be used on an ongoing basis to identify whether the currently selected management actions should change. Future changes to the management actions could be:

- **Additional** required actions (for instance where additional water uses are identified, or if PFAS concentrations in the environment increase).
- **Reduced** required actions (for instance where lower PFAS concentrations in the environment mean that previously established management actions are no longer necessary to manage risks).



1.3 Scope of Works

To achieve the above objectives, Senversa completed the following scope of work detailed in the OMP:

- Water sampling of tanks and taps on-site (Airport terminal, maintenance shed, fire station, and Airport Bore).
- Sampling of tanks and taps on council sites (works depot).
- Sampling of surface water along Mission Creek, Watermill Creek and the end of Cascade Creek and Headstone Creek.
- Sampling of private bores along Mission Creek.
- Additional sampling, not in the OMP, to address data gaps:
 - Drinking water sample locations to confirm concentrations remain below the revised guideline levels including the hospital, electricity depot, waste management centre, Bureau of Meteorology (BoM) station, additional fire station taps, additional airport terminal taps, council office and permitted water carting sources.
 - Residential soil and produce sampling following revised residential soil guideline levels.
 - Dam biota and sediment sampling to indicate risk of application to land following dam clearing.
 - Soil samples to further delineate potential source areas at Ball Bay fuel storage area and down pipe supplied by the Airport Bore.
- Preparation of this PFAS OMP Year 4 Monitoring Report.



2.0 Background

2.1 Airport Details

Site identifying details are summarised below.

Table 2-1: Airport Details

| Item | Relevant Site Information |
|---------------|--|
| Site Location | The Airport location is indicated on Figure 1 . |
| Site Area | ~120 hectares (ha). |
| Site Use | <p>The Airport layout is indicated on Figure 2A. Key site features include:</p> <ul style="list-style-type: none"> Operational Airport with two runways. Aircraft and Airport operational infrastructure in the northeast portion of the site, including terminals, storage and cargo facilities. Maintenance facilities in the mid-eastern portion of the site. Fire station in the mid-eastern portion of the site, south of the maintenance facilities. Waste management centre, BoM station and electricity depot in the northern portion of the site. Wastewater treatment plant in the central northern portion of the site. The fire training facility. Airport groundwater bore, pumped to a concrete holding tank prior to treatment through a point of entry treatment (POET) filter. |
| Zoning | <ul style="list-style-type: none"> Site: light industry land use. Surrounding land: rural and rural residential land use. |

2.2 Environmental Setting

The environmental setting is summarised below in **Table 2-2**. Further environmental setting details are outlined in the DSI (Senversa, 2021b).

Table 2-2: Environmental Setting

| Item | Relevant Site Information |
|-----------------------|--|
| Surrounding Land Uses | <ul style="list-style-type: none"> North: Mission Creek is located to the immediate north-west of the site followed by St Barnabas Chapel, rural properties and Headstone Reserve. The Norfolk Island National Park is located approximately 2 kilometres (km) to the north of the site. East: Northeast of the site is the township of Burnt Pine, consisting of mixed land use. The land to the immediate east consists of rural and rural residential land. South: Rural residential properties, Point Ross and Bumbora Reserve followed by the South Pacific Ocean approximately 400 metres (m) from the most southern point of the site. West: Rural residential properties, Rocky Point and 100 Acres Reserve followed by the South Pacific Ocean approximately 400 m from the most western point of the site. |
| Climate | <p>Norfolk Island is classified as a sub-tropical climate which is primarily affected by high-pressure systems which fluctuate over the island annually. The mean maximum temperatures on the island range from 18°C in winter to 25°C in summer with a high average relative humidity of 73% to 81% (BoM, 2022). Norfolk Island's median annual rainfall is 1,280 mm with the highest rainfall between April to August and monthly means of approximately 120 to 143 mm. The driest month is typically November with an average rainfall of 73 mm (BoM, 2024). Rainfall on the island between 2016 and early 2020 was below average and little to no rain fell on the island between October 2019 and February 2020 (BoM, 2022). Above average rainfall was recorded from April to November 2025 preceding the Year 4 monitoring event.</p> |



| Item | Relevant Site Information |
|----------------------------------|---|
| Topography | The Airport site is generally flat, between 95 and 115 m above sea level (Geoscience Australia, 2020). The surrounding island undulates rapidly with several water catchment zones creating steep valleys and low-lying creeks. |
| On-site Drainage Networks | On-site stormwater in the north-east of site drains into a low-lying area on the boundary of the site into a stormwater drainage pipe which runs perpendicular to and under the road leading to the waste treatment centre. This stormwater drainage is understood to discharge into Mission Creek. Across the Airport in general, stormwater is expected to run towards the site boundary, away from the runways. |
| Hydrology | Creeks are largely ephemeral, flowing only during rainfall events. Water catchment zones are shown on Figure 1 . Mission Creek, Headstone Creek, Watermill / Town Creek and Rocky Point Creek are considered down-gradient of the Airport, with the Mission Creek Catchment considered the most vulnerable to PFAS impacts migrating from the Airport due to historical fire training activities undertaken on that side of the Airport which used AFFF containing PFAS. All creeks discharge to the South Pacific Ocean. |
| Geology and Soils | Norfolk Island is the erosional remnant of Pliocene aged volcanic centres located on a north trending continental ridge between New Zealand and North Caledonia (Abell, R S & Falkland A C, 1991). The island consists of the former shield volcano (Mt Pitt) and horizontal basalt flows. The prominent soil type found at and surrounding the site is the Rooty Hill Clay. |
| Acid Sulphate Soils | Peaty acid sulfate soils are present in the lower landscape portion of the island, with the largest known area located in the lower portion of the Watermill Creek Catchment. |
| Hydrogeology | The following hydrostratigraphic sequence is recorded (Abell, 1993): <ul style="list-style-type: none"> • Weathered volcanic mantle: Major aquifer on the island, porous but clayey. The upper water table on Norfolk sits within the weathered mantle. • Basaltic lavas: Heterogeneous water-bearing systems, dominated by water movement through fractures, joints and bedding. • Vertical movement of groundwater through fractures in the basalt likely form localised, semi-confined aquifers within tuff beds and fragmented layers. The heterogeneous nature of basaltic aquifers results in a complex groundwater flow regime. In general, groundwater flow follows, to a subdued degree, topographic features, discharging to surface water bodies and further towards the coastline. |
| Terrestrial Environments | Limited on-site flora and fauna are present due to the highly modified nature of the Airport environment. Prior to European settlement, Norfolk Island was dominated by subtropical rainforest and native flora of which over 30% is endemic (CSIRO, 2020). A large proportion of the island has been cleared for farmland used for grazing or cropping, with intact native communities being largely restricted to the 6.5 km ² Norfolk Island National Park centred around Mount Bates and Mount Pitt. |

2.3 PFAS Source Areas

The PSI and DSI identified 17 confirmed or potential PFAS source areas across the island. Generally, the most significant source areas ('Group 1 Source Areas') were locations where there was repeated application of foams and concentrate.

Group 1 Source Areas were confirmed through soil testing and include the following, located on the airport:

- PFAS Source (PS) Area 01: The former fire station and foam shed.
- PS Area 02: Flushing out area in the northeast corner of the site.
- PS Area 03: The former drill ground south-west of the former fire station in the northeast portion of the site. This area is now utilised by the waste management centre, which includes a composting facility.
- PS Area 04: Current fire drill area along the northern site boundary.
- PS Area 05: The airport maintenance depot where general maintenance of fire trucks historically occurred.
- PS Area 06: The current fire station.



Other source areas that were considered to be less significant based on frequency of AFFF application were identified and grouped as follows:

Group 2 Source Areas (PS Areas 7-11) – Areas where legacy AFFF concentrate and / or foam was used or stored more than once, but with less frequent rates of application than Group 1. Also includes secondary source areas like the wastewater treatment plant on the Airport (PS11), which is considered likely to be a higher source of PFAS flux than the other Group 2 Source Areas.

Group 3 Source Areas (PS Areas 12-15) – Areas where a single application of foams occurred due to an incident or a one-off event.

Group 4 Source Areas (PS Areas 16 & 17) – Areas where no AFFF is known to have been used, however water containing elevated concentrations of PFAS used.

Key source areas are indicated on **Figure 2-1** below and appended **Figure 2A** and **2B** and further information on potential PS Areas is provided in the DSI.



Figure 2-1: PFAS Source Areas

2.4 Water Sources and Use

A water use survey was conducted during the DSI (Senversa, 2021b) to identify potential PFAS migration and exposure pathways across the island. The understanding of current water use is refined during each subsequent monitoring event through consultation with the DITRDCA on-island team, Norfolk Island Regional Council (NIRC) and private property owners contacted for sampling. The Year 4 monitoring event date was announced in the local newspaper. Available key information on water sources and use is summarised below, which supersedes past understanding presented in earlier reports.

2.4.1 Groundwater Source

Council-provided survey data indicates that there are 228 active groundwater bores, 38 dry bores and 10 “contaminated” bores across the island (Senversa, 2021a). Other sources indicate approximately 450 bores exist across the island (Abell, 1993). It is understood that not all bores on the island are registered with the NIRC or surveyed for elevation or location.

Groundwater is known to be extracted for limited drinking water supplied by water carters, and for stock watering (chickens and cows), refer to **Sections 2.4.3** and **2.4.4**. There is no evidence to suggest that groundwater is extracted for recreational purposes (e.g., to fill a swimming pool).



On the Airport, there is one known groundwater well that was not in use at the time of the investigation. Immediately off-site, a second 'Airport Bore' is operational. Water from the bore is pumped into a concrete holding tank, then filtered through a POET filter, and finally supplied for a range of uses via either pipework or the Ben Christian Down Pipe near the waste management centre access track just off Douglas Drive (see further discussion in **Section 2.4.2**). The down pipe was previously referred to as the Airport Truck Fill and was locked at the time of the Year 4 monitoring event.

2.4.2 Airport Bore Water Use

Information on the Airport Bore water treatment and use has been provided progressively by NIRC and updated during the Year 4 monitoring event, summarised in **Table 2-3**. Due to the anecdotal nature of some information, pipework connection records sometimes contradict across monitoring events and PFAS analysis results have been assessed in **Section 5.3** to confirm the understanding presented below. Not all locations presented below were monitored during the Year 4 monitoring event, having previously been assessed during the DSI (Senversa, 2021c).

Since at least June 2024, the Airport Bore water has been pumped to the concrete holding tank then filtered via a POET filter prior to use. The filter was previously located at the Fire Station where it was used to filter water used for equipment flushing. NIRC tests the treated water quarterly to confirm filter efficacy. Results of the NIRC testing are further discussed in **Section 5.3**.

Table 2-3: Airport Bore Water Use Summary

| Area / Sample Location | Historical Water Source | Current Water Source | Description |
|--|-------------------------|----------------------|--|
| AIRPORT (ONSITE) | | | |
| Airport Bore Tank | Airport Bore | Airport Bore | Airport Bore sampled from concrete tank adjacent former council offices. Use as holding tank before water is currently treated via POET and historically pumped to uses below. |
| A-TANK 1 | Airport Bore | Rainwater | Open water pit within building now filled via rainwater. |
| Airport tanks A_TANK 2 and A_TANK 3 (new tanks since 2020 PSI sampling) | Rainwater | Rainwater | Tanks have been connected to buildings within the airport precinct. |
| Airport Terminal Bathrooms and drinking fountains | Airport Bore | Rainwater | Historically sourced from Airport Bore, now sourced from new airport rainwater tanks. Plumbing has not been replaced. |
| Maintenance Shed at Airport (A_TAP3) | Airport Bore | Rainwater | Water now sourced from new rainwater tanks. |
| Former Fire Station - Mechanical Shed adjacent airport terminal and Gate 1 (A_TAP4) | Airport Bore | Airport Bore | Airport bore water may still be connected based on PFAS concentrations detected, however non-use warnings in place and the kitchen was not in use during the 2025 monitoring event (NIRC, 2025). |
| BoM Office (A_TAP5) | Airport Bore | Rainwater | Plumbing has not been replaced. |
| Electricity Depot | Airport Bore | Rainwater | Plumbing has not been replaced. |
| Waste Management Centre | Airport Bore | Rainwater | Plumbing has not been replaced. |
| WWTP | Airport Bore | Treated Airport Bore | Piped water used to flush the water treatment plant. |



| Area / Sample Location | Historical Water Source | Current Water Source | Description |
|--|-------------------------|----------------------------------|--|
| Current Fire Station taps (kitchen, laundry, toilets) (FRE_TAP1, TAP3, TAP5) | Airport Bore | Rainwater, desalinated water | Water supply transferred to rainwater. Connection was then mistakenly made to rainwater tanks that contained a PFAS-impacted inner liners sometime between June 2023 and June 2024. Following this discovery, temporary plumbing was installed for the kitchen, the bathrooms and the laundry in July 2024 connected to a desalinated water tank. Reconnection to relined rainwater tanks planned. |
| Current Fire Station Emergency Management Norfolk Island (EMNI) kitchen taps (FRE_TAP6) | Airport Bore | Rainwater | It is understood from discussions with NIRC that water is supplied by the relined rainwater tanks noted above. The plumbing has not been replaced. |
| Fire Hydrants on Airport (FRE_TAP2) | Airport Bore | Treated Airport Bore & rainwater | Piped under runway to two concrete underground storage tanks. Used for firefighting, fire systems testing, firefighting training and fire truck flush outs. More detail provided below. |
| Former Council Offices at Airport (COUNCIL_TAP1, TAP2) | Airport Bore | Rainwater | Historically sourced from Airport Bore, now sourced from new airport rainwater tanks. The offices were not in use during the 2025 monitoring event. |
| HOSPITAL (OFFSITE) | | | |
| Hospital Tank - concrete underground (PWS_HOSP_TANK1) | Airport Bore | Rainwater | Historically sourced from airport bore via underground line and filling of tanks. All tanks are connected to hospital master tank and filled with rainwater collected from hospital roof. |
| Hospital Tank - all other tanks (PWS_TANK2-TANK8) | Rainwater | Rainwater | All tanks are connected (feed in) to master tank PWS_HOSP_TANK1 |
| Taps (PWS_HOSP_TAP1-TAP21) | Airport Bore | Rainwater | Through PWS_HOSP_TANK1. Plumbing and filters were replaced in late 2020. |
| WIDER NORFOLK ISLAND INVESTIGATION AREAS | | | |
| Council Works Depot (DEPOT_TANK1,TANK2 and TANK3) | Airport Bore | Rainwater | Underground concrete Tank1 and Tank2 previously supplied by Airport Bore Water. Now supplied by rainwater and used to supply the kitchen tap and for emergency firefighting. New Tank3 installed at rear of depot for rainwater and used for truck washing at the mechanical sheds in the works depot. |
| Council Works Depot kitchen taps (DEPOT_TAP1) | Airport Bore | Rainwater | Historically sourced from Airport Bore, now sourced from Tank2. |
| Ben Christian Down Pipe | Airport Bore | Treated Airport Bore | Airport Bore water was publicly accessible via the down pipe until 2020. The down pipe is now locked and in use by NIRC for the public uses described in this table. |
| Public Toilets | Airport Bore | Treated Airport Bore | Filled via water carter from Ben Christian Down Pipe during low rainfall. |
| Quarry Dust Suppression | Unknown | Treated Airport Bore | NIRC advised in 2025 that treated Airport Bore water, carted from the Down Pipe, is utilised for this purpose. The duration of use is unknown. |
| Roads Depot Gravel Washing for Road Maintenance | Unknown | Treated Airport Bore | NIRC advised in 2025 that treated Airport Bore water, carted from the Down Pipe, is utilised for this purpose. The duration of use is unknown. |



Along with firefighting, the underground fire station tanks are used for fire systems testing, fire truck flush outs and firefighting training. It is understood flush outs of the fire trucks occurred up to three times a week and historically took place in the unsealed area to the south of the former fire station (PS02) where it would runoff towards Mission Creek. Currently, the fire station uses approximately 15,000 L of water once per fortnight for a live fire training test (NIRC, 2020).

2.4.3 Potable Drinking and Stock Water Use

Potable drinking and stock water use are understood to remain largely unchanged since the water use survey was undertaken in 2020.

Bore water is not widely consumed in times of high rainfall and potable and stock watering water are generally supplied by rainwater tanks. In times of drought, when tank water is not readily available, bore water may be extracted for drinking water purposes (NIRC, 2018). In 2019 in response to extreme water shortages on-island, an Australian Defence Force desalination plant was installed and operated for an initial 6-week period before being re-established in 2020 to assist with suitable potable water supply for the Airport Runway Reseal Project. The plant was permanently re-installed in 2024 to provide an alternate water supply during dry periods.

Water carting from groundwater bores is understood to remain operational and was investigated as part of the PSI. Prior to the Year 4 monitoring event, NIRC advised that four water carters were in operation. Limited information on the location and nature of the water carting water sources is available due to the locations of assets being kept largely confidential by the water carters.

Bore water is known to be extracted for stock watering (cows) on at least one property in the Mission Creek catchment. Mission Creek surface water is known to be currently pumped for stock watering (cows) on at least one property and historically for chickens on one property (ceased in 2023). Water extracted from Watermill Creek is also understood to be used for livestock watering between the Airport and the Duck Dam. Both creeks are accessible to cattle on multiple properties and utilised for stock watering, including publicly accessible Mission Pool utilised by road side cattle herds.

NIRC advised residents currently have access to water from one public standpipe adjacent to Headstone Creek (Headstone Dam). This water is understood to be used for non-potable uses (including stock watering).

2.4.4 Irrigation Water Use

Irrigation water use is understood to remain largely unchanged since the water use survey was undertaken in 2020.

Irrigation water is understood to not be used on-airport, however grass on-site may be affected by rainfall runoff over impacted soils and over areas of historical AFFF use (NIRC, 2020).

Irrigation water derived from bores and pumped surface water is used across the island for small commercial and private residential gardens. CSIRO (2020) estimated approximately 10.8 ha of cultivated land is used for commercial food production, up to 75% of which may be irrigated.

An additional 5 ha of land is estimated to be used for medium to large scale vegetable gardens. It is unknown to what extent these gardens are irrigated and the source of water across the island more broadly (CSIRO, 2020). The water use survey and OMP monitoring have focused on Mission and Watermill Creek Catchments where elevated PFAS concentrations have historically been identified. One property was previously identified in these catchments where water was pumped from Mission Creek to irrigate home produce but has been discontinued since at least 2023. One property was also identified in the Mission Creek Catchment where bore water was in use for home produce irrigation during low rainfall, and this water use was active during the 2025 monitoring event.

It is understood that water is not widely used for irrigation of grassed paddocks (i.e., for livestock grazing) on the island.



3.0 Sampling and Analysis Approach

3.1 Sampling Analysis and Quality Plan

The SAQP details the data quality objectives (DQOs), data quality indicators (DQIs) and assessment methodology of the monitoring, included in **Appendix A**. Deviations from the SAQP are summarised in the table below.

Table 3-1: Summary of deviations from the SAQP

| Item | Relevant Site Information |
|------------------------------------|---|
| Moved Sample Location | <ul style="list-style-type: none"> MC_OMP04a was collected slightly upstream of usual location MC_OMP04 due to thick vegetation. Similarly, MC_OMP05a was collected slightly upstream of usual location MC_OMP05. MC_OMP07a was collected upstream of MC_OMP07 across Douglas Drive as no access was provided to the MC_OMP07 and MC_OMP08 access property. |
| Sample Not Collected | No access was provided to MC_OMP08 access property, and this sample was not collected. Samples MC_OMP07a and MC_OMP09 up and downstream of this location were collected. |
| Alternate Sample Location | <ul style="list-style-type: none"> As in the Year 2 and 3 Monitoring Events, ID013_BORE was sampled in lieu of ID013_SW01, as the property owner advised that the bore was now being used instead of the pumped creek water (SW01) for fruit tree and garden irrigation. ID013_BORE had previously been disused due to high salinity. No access was provided to ID015_BORE. ID019_BORE, not currently in use, was sampled in lieu of this location. |
| Sample Not Analysed | DEPOT_TANK3 was not analysed as it was confirmed with NIRC that this tank is supplied only with rainwater and has not recorded PFAS above the laboratory limit of reporting (LOR) during the previous three monitoring events. |
| Additional Samples Analysed | In addition to the OMP SAQP scope of works, additional monitoring was undertaken as part of the Year 4 event, detailed in Table 3-4 . |

Biota and soil sampling undertaken during the Year 4 monitoring event were not included in the SAQP methodology. The sampling methodology is detailed below.

Table 3-2: Additional Sampling Methodology

| Activity | Description and Further Information |
|----------------------------------|--|
| Biota Sample Collection | <p>Biota samples were collected in consultation with the property manager (NIRC or the property owner as applicable) to sample plant material meeting the following criteria:</p> <ul style="list-style-type: none"> Most prevalent species or for home produce, with the highest consumption potential, present in target area. Distribution of species such that differences in PFAS uptake are captured. For example, a representative selection of leafy, fruiting and root produce may be sampled. <p>Biota samples were collected with a gloved hand and placed directly into laboratory supplied sample jars using disposable nitrile gloves. Large samples, e.g. tomato, were first cut with a knife, decontaminated prior to use. A dry sample weight of 10 g was required for laboratory analysis.</p> |
| Biota Sample Preservation | To meet biosecurity import requirements, biota samples were preserved by desiccation in silica beads directly in the sample jar in a ratio of one part plant material to ten parts silica beads. |
| Soil Sample Collection | Soil samples were collected from the near-surface using a gloved hand with the aid of a small hand trowel/shovel and transferred into sample jars using disposable nitrile gloves. The trowel was decontaminated between sampling locations. |



| Activity | Description and Further Information |
|---------------------------------|---|
| Soil Sample Preservation | Refer to water sample preservation presented in the SAQP, included in Appendix A . |

3.2 Monitoring Conducted

Sampling locations, excluding samples collected on private properties, are indicated on **Figure 1**.

Australian Laboratory Services Pty Ltd (ALS) was the primary analytical laboratory for soil, sediment, water and biota samples and Envirolab was the secondary laboratory. Both laboratories are National Association of Testing Authorities (NATA) accredited for the analyses conducted. All samples were analysed for the extended PFAS suite of 28 analytes.

Laboratory results and field water quality observations were uploaded to the Esdat³ database.

Table 3-3: PFAS OMP Year 4 Sample Location Summary

| Sample Purpose | Sample Location | Number of Locations | Sample IDs | Sample Media |
|-------------------------------|---|---------------------|---|--------------------|
| Creek Sampling | Mission Creek | 11 | WWII_DAM, MC_OMP01 to MC_OMP07a, MC_OMP09 to MC_OMP11 | Surface water |
| | Watermill Creek | 5 | WC_OMP01 to WC_OMP05 | |
| | Cascade Creek | 1 | COCKPIT_SW01 | |
| | Headstone Creek | 1 | PWS_HEAD_DAM | |
| Irrigation Water | Mission Creek Catchment groundwater | 2 | ID013_BORE ^{2, 3} ID016_BORE ² | Point of use water |
| Stock Water | Mission Creek Catchment groundwater and surface water | 2 ¹ | ID014_BORE ⁴ ID019_BORE ² (MC_OMP09 and MC_OMP10 ¹) | |
| Managed Water Supplies | On-Airport | 5 | AIRPORT_BORE ^{2, 5} FRE_TAP1 FRE_TAP2 A_TAP1 A_TAP4 | |
| | Off-Airport | 3 | DEPOT_TAP1 DEPOT_TANK1 DEPOT_TANK2 | |
| Total Primary Samples | | 30 | | |

Table notes:

- There are 5 samples relevant for stock watering, however only two unique samples (ID014_BORE and ID015_BORE, used for cattle watering) not also collected for another purpose. MC_OMP08, MC_OMP09 and MC_OMP10 (creek locations with possible cattle access in the absence of management) are included in the total sample numbers for creek sampling.
- Groundwater location.
- ID013_BORE analysed in lieu of ID013_SW01, refer **Section 3.1**.
- Pumped surface water location.

³ Environmental data management software.



5. AIRPORT_BORE sample is collected from the above-ground concrete holding tank (pre-treatment) adjacent to the POET filtration system and former council offices on the airport.

6. Green text indicates samples analysed for trace PFAS analysis, perfluorooctane sulfonate (PFOS) LOR 0.002 micrograms per litre (µg/L).

In addition to the OMP SAQP scope of works, additional monitoring was undertaken as part of the Year 4 event, detailed in **Table 3-4**. The additional monitoring was undertaken to address the following identified data gaps:

- Updated PFAS drinking water guidelines (DWG) have been released in the Australian Drinking Water Guidelines (ADWG) in June 2025. The 2025 revised DWG for PFOS is below the LOR utilised in previous analysis, further discussed in **Section 3.3.3**. As such, where PFOS was previously not detected in water used for drinking water, these samples have been re-analysed for trace level PFAS, indicated in green in **Table 3-3** and **Table 3-4**.
- Updated human health investigation levels for soil have been released in the National Environmental Management Plan (NEMP) in March 2025. The revised PFOS Human Health Investigation Level (HIL) for residential use with accessible soil (HIL-A) is 0.003 milligrams per kilogram (mg/kg). Soil PFOS concentrations off-airport commonly exceeded this revised investigation level, including at two residential properties with an identified PFAS source. One of these two properties was identified to grow produce for home consumption. Further soil and produce sampling was undertaken at this property.
- Two previously identified PFAS source areas at the Ball Bay fuel storage area and Ben Christian down pipe supplied by the Airport Bore were further investigated to better understand whether PFAS impacts to soil requiring management were present. Sampling was not undertaken with the objective of delineation of impacts and was designed to detect the presence of gross contamination requiring further action.
- NIRC advised that Duck Dam has previously (prior to 2020) been routinely cleared of vegetation and sediment to reduce risk to cattle and maintain the structure. Clearing has been delayed since this date due to uncertainty with respect to vegetation and sediment PFAS concentrations and required management. Preliminary PFAS analysis was undertaken to inform management of these waste materials following removal.

Table 3-4: OMP Additional Sample Location Summary

| Sample Purpose | Sample Location | Sample Matrix | Details | Sample IDs |
|------------------------|---|---------------|---|-------------|
| Managed Water Supplies | Airport terminal departure lounge drinking fountain | Water | Supplied by rainwater tanks, previously supplied by the airport bore. | A_TAP15 |
| | BoM Station kitchen | | Supplied by rainwater tanks, previously supplied by the airport bore. | A_TAP5 |
| | Electricity Depot kitchen | | Supplied by rainwater tanks, previously supplied by the airport bore. | ELEC_TAP1 |
| | Waste Management Centre kitchen | | Supplied by rainwater tanks, previously supplied by the airport bore. | WASTE_TAP1 |
| | Council Office kitchen | | Supplied by rainwater tanks and new plumbing. This sample was collected as a control/background sample for the trace PFAS analysis. | COUNCIL_TAP |
| | Fire Station | | EMNI kitchen tap. Supplied by rainwater, previously supplied by airport bore water. | FRE_TAP6 |



| Sample Purpose | Sample Location | Sample Matrix | Details | Sample IDs |
|------------------------------|--|---------------|---|---|
| | Hospital | | Hospital taps previously sampled following filter and plumbing replacement. Re-sampled during the Year 4 monitoring event at a lower LOR to capture the revised drinking water guideline thresholds. Locations selected to align with previous monitoring and targeting main water use points. Hospital advised that the dialysis room is no longer in use. | PWS_HOSP_TAP1 PWS_HOSP_TAP3 PWS_HOSP_TAP4 PWS_HOSP_TAP6 PWS_HOSP_TAP8 |
| Potable Water Use | Water carting bore water | Water | Two of the four identified water carters authorised testing of bore water sources. Both WC-03_BORE and WC-04 samples were collected from taps connected directly to pumped bores. This is to distinguish from WC-03 water which was also collected in 2021 from a water carting truck and tank filled with carted water. ² | WC-03_BORE WC-04 |
| Soil and Sediment | PFAS source area on residential property | Soil | Soil tested within vegetable garden of residential property. | ID008_SS2 |
| | Ben Christian down pipe | | Soil below the down pipe analysed given historical truck filling with PFAS impacted water at this location. | A_TRUCKFILL_SS1 |
| | Ball Bay fuel storage area | | Soil within the firefighting water retention basin and adjacent the two PFAS drum storage areas. | BALL_SS3 BALL_SS4 BALL_SS5 |
| | Duck Dam | Sediment | Sediment from Duck Dam was analysed to manage proposed desilting activities. | WC_OMP04_DUCKDAM |
| Biota | PFAS source area on residential property | Biota | Representative samples analysed from vegetable garden including tomato, zucchini, silver beet and parsley. | I008_BIOTA1 to ID008_BIOTA4 |
| | Duck Dam | | Predominant vegetation from Duck Dam, water hyacinth, was analysed to manage proposed clearing activities. | WC_BIOTA1 |
| Total Primary Samples | | | | 24 |

Table notes:

- Green text indicates samples analysed for trace PFAS analysis to LOR 0.002 µg/L.
- WC-03_TRUCK and WC-03_TANK samples were not analysed.



3.3 Decision Framework

3.3.1 Trigger Values

The analytical results collected as part of the monitoring program have been used to assess whether the currently selected management actions should change. A decision framework has been developed in the OMP to define the conditions under which further assessment of the appropriateness of the current management measures is required (Senversa, 2021g). This decision framework includes trigger values:

- **Upper Trigger Values (UTVs)** are defined for use where risks are currently assessed to be low and acceptable. Where concentrations previously found to be associated with low and acceptable risks increase to be above the UTVs, review of the risk profile is required to assess if additional management is required.
- **Lower Trigger Values (LTVs)** are defined for use where exposures currently require management. Where concentrations previously found to be associated with potentially elevated risks decrease to be below the LTVs, review of the risk profile is required to assess if management measures can be reduced.

Where trigger values reference the ADWG Health-Based Guideline Values (HBGV), the 2020 NEMP⁴ values have been retained for consistency across the initial OMP implementation period. The revised 2025 DWG have been considered separately, detailed in **Section 3.3.3**.

Table 3-5: Upper Trigger Values

| Sampling Medium | Upper Trigger Value (µg/L) | | | Rationale |
|---|----------------------------|-------|------------|---|
| | PFOS | PFHxS | PFHxS+PFOS | |
| Mission Creek Catchment Water Used for Irrigation | 4.2 | 2.5 | - | The risk to consumers of irrigated produce associated with the previously measured range in concentrations at ID013 (1.4 - 2.8 µg/L PFOS; 1.4 - 1.5 µg/L Perfluorohexane sulfonate (PFHxS)) is assessed to be low and acceptable. Given the conservatism in the assessment, small variations above the previously measured range are considered unlikely to alter the risk profile. Trigger values approximately 50% above the upper end of the assessed range are adopted as indicative of a requirement to assess whether the potential risks have increased, and if further management is required. The triggers can be applied at property ID016, where water is used for irrigation. |
| Surface Water from Other Creeks | 0.5 | 1.3 | - | Outside of Mission Creek, surface water concentrations of up to 0.29 µg/L PFOS and 0.85 µg/L PFHxS have previously been measured. The HHERA assessed risks to consumers of produce irrigated with this water and risks to consumers of livestock products where this water is used for stock watering, as low and acceptable. Given the conservatism in the assessment, small variations above the currently measured range are considered unlikely to alter the risk profile. Trigger values approximately 50% above the upper end of the assessed range are adopted as indicative of a requirement to assess whether the potential risks have increased, and if further management is required. It is noted that the adopted trigger values are more stringent than the majority of the conservative screening levels for stock water and irrigation pathways, and within 50% of the most stringent values. Given the conservatism in the HHERA screening levels, it is assessed that provided concentrations remain below the triggers, risks will remain low and acceptable, regardless of water usage for irrigation or stock watering. |
| Surface Water from Cascade | - | - | 0.07 | PFAS concentrations previously measured in Cascade Creek and Headstone Creek were below the HBGV for drinking water, and no |

⁴ Heads of Environment Protection Authority (HEPA) (2020), *PFAS National Environmental Management Plan 2.0*.



| Sampling Medium | Upper Trigger Value (µg/L) | | | Rationale |
|-------------------------|----------------------------|-------|------------|--|
| | PFOS | PFHxS | PFHxS+PFOS | |
| Creek / Headstone Creek | | | | management measures are currently in place for this water. The drinking water HBGV is selected as the UTV. If concentrations increase above this level, further assessment of the requirement for management of water use is required. |

Table 3-6: Lower Trigger Levels

| Sampling Medium | Lower Trigger Value (µg/L) | | | Rationale |
|---|----------------------------|-------|------------|--|
| | PFOS | PFHxS | PFHxS+PFOS | |
| Domestic water use: Reticulated water supplies at public facilities | | | | The drinking water HBGV (2020) has been selected as the LTV. Where concentrations are below this level, ongoing management of water use (including for sensitive use as drinking/domestic water) is unlikely to be required. |
| Surface water from Mission Creek | - | - | 0.07 | |
| Surface water from Watermill Creek | | | | |
| Groundwater at Airport (Airport Bore) | | | | Where water is used for drinking water, the revised HBGV (2025) should be applied. |
| Mission Creek water used for cattle stock watering prior to management | 0.33 | 1.2 | - | The conservative beef cattle stock watering screening levels adopted in the HHERA are adopted as the LTV; if concentrations remain consistently below these values, ongoing management is unlikely to be required. |
| Mission Creek water used for chicken stock watering prior to management | 0.9 | 1.3 | - | The conservative chicken stock watering screening levels adopted in the HHERA are adopted as the LTV; if concentrations remain consistently below these values, ongoing management is unlikely to be required. |

3.3.2 Decision Trees

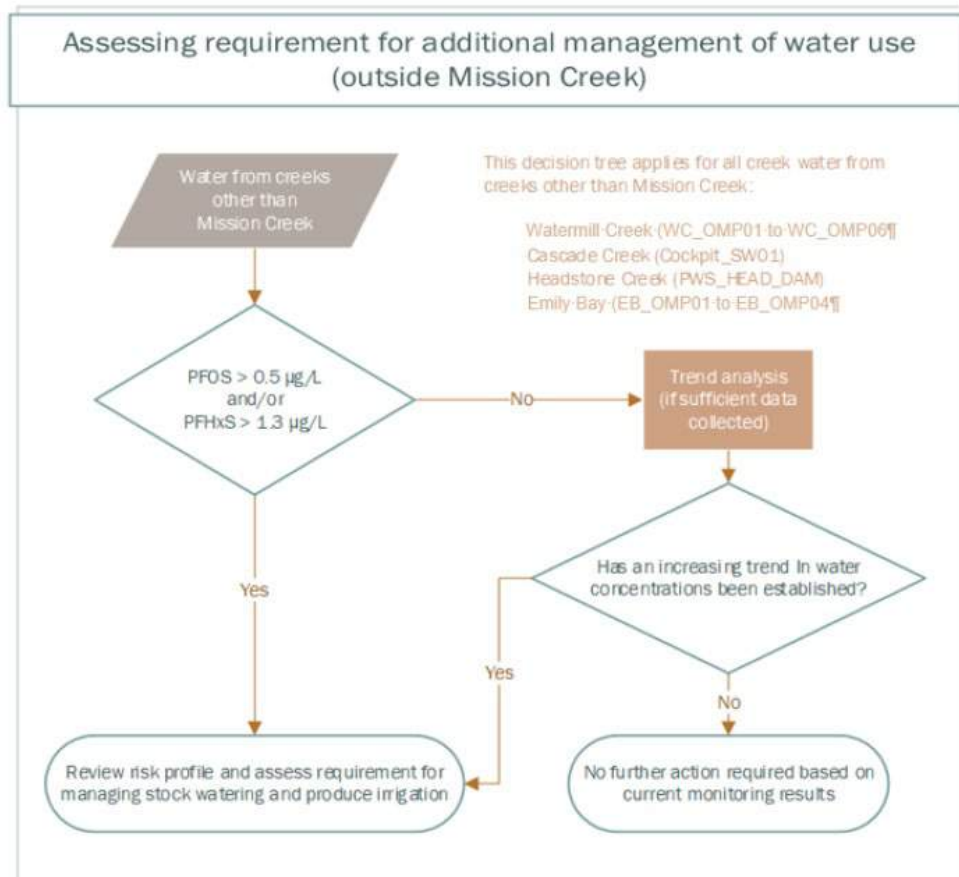
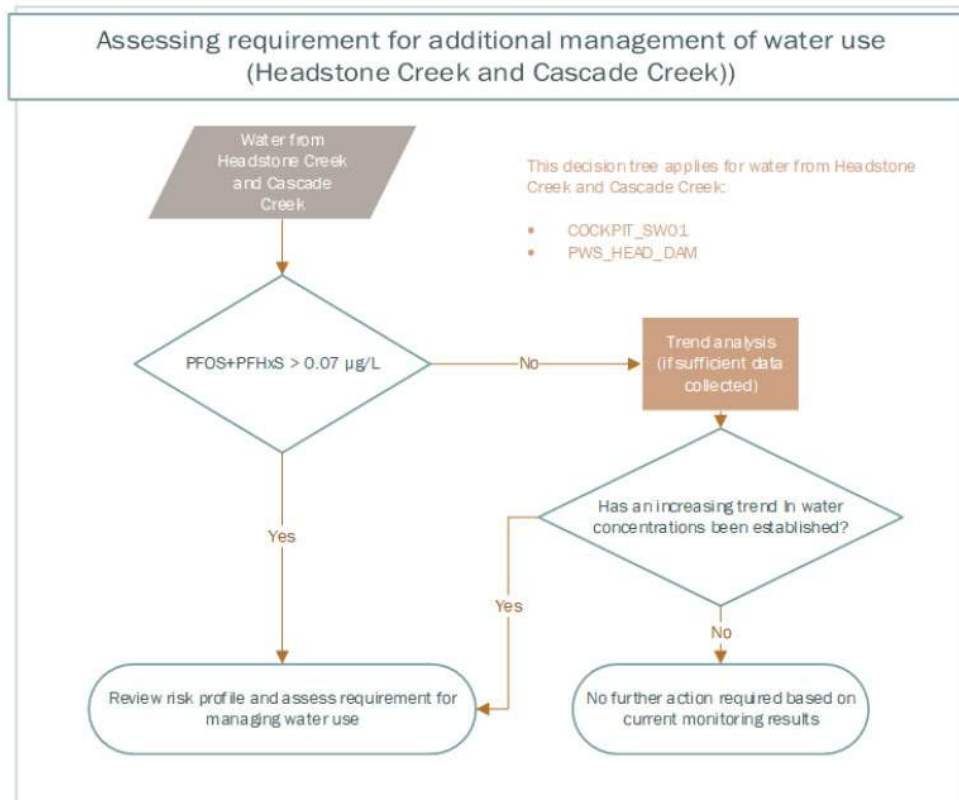
The flowcharts (“decision trees”) presented on the following pages detail the decision framework process for the different samples collected in the PFAS OMP.

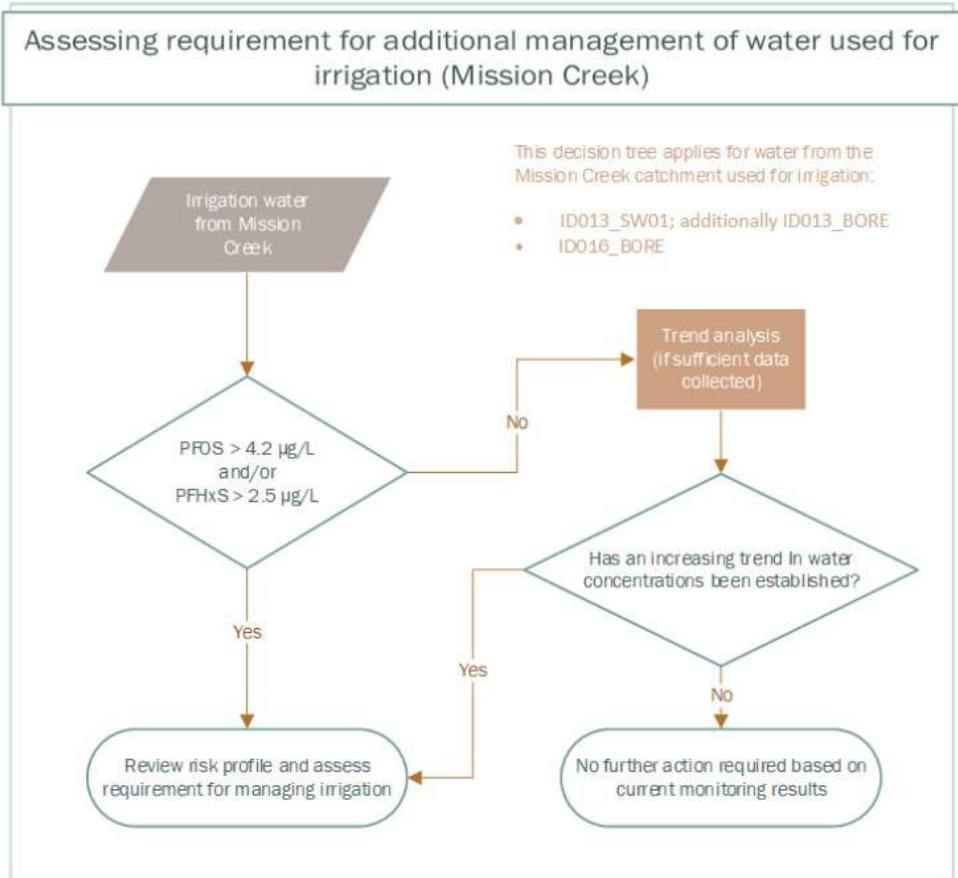
Separate decision trees have been presented for each water use and actions to take in the case that either:

- **The UTVs are exceeded**, requiring an assessment of the requirement for additional management of risks currently assessed to be low and acceptable; or
- **The LTVs are not met**, requiring an assessment of whether current management is still required.



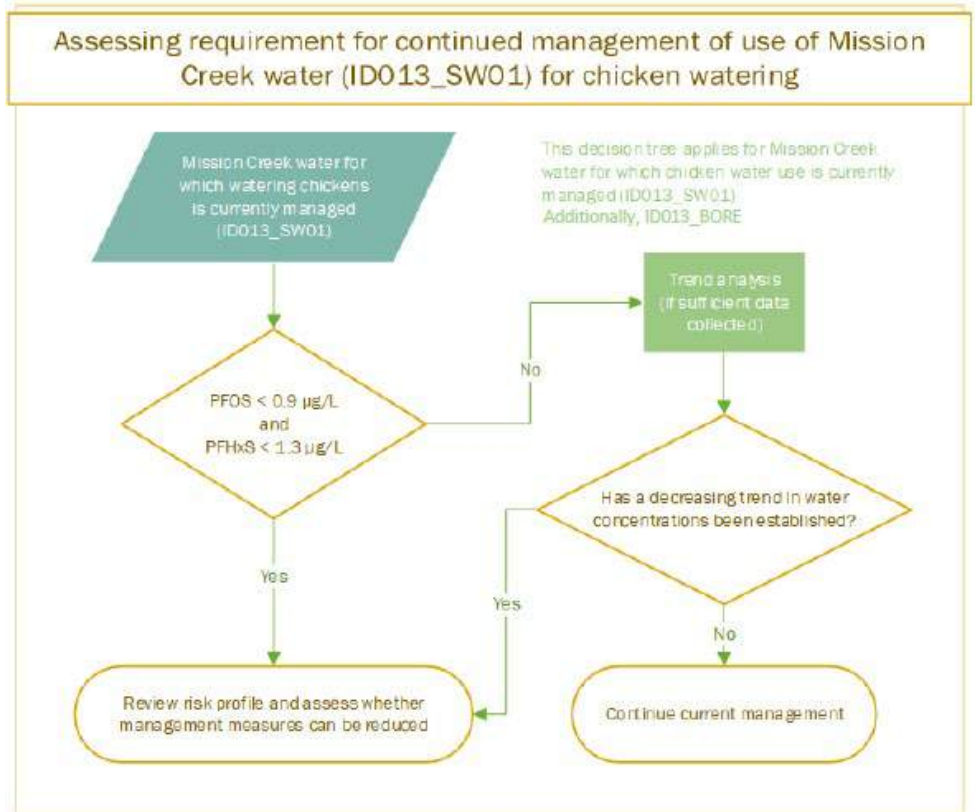
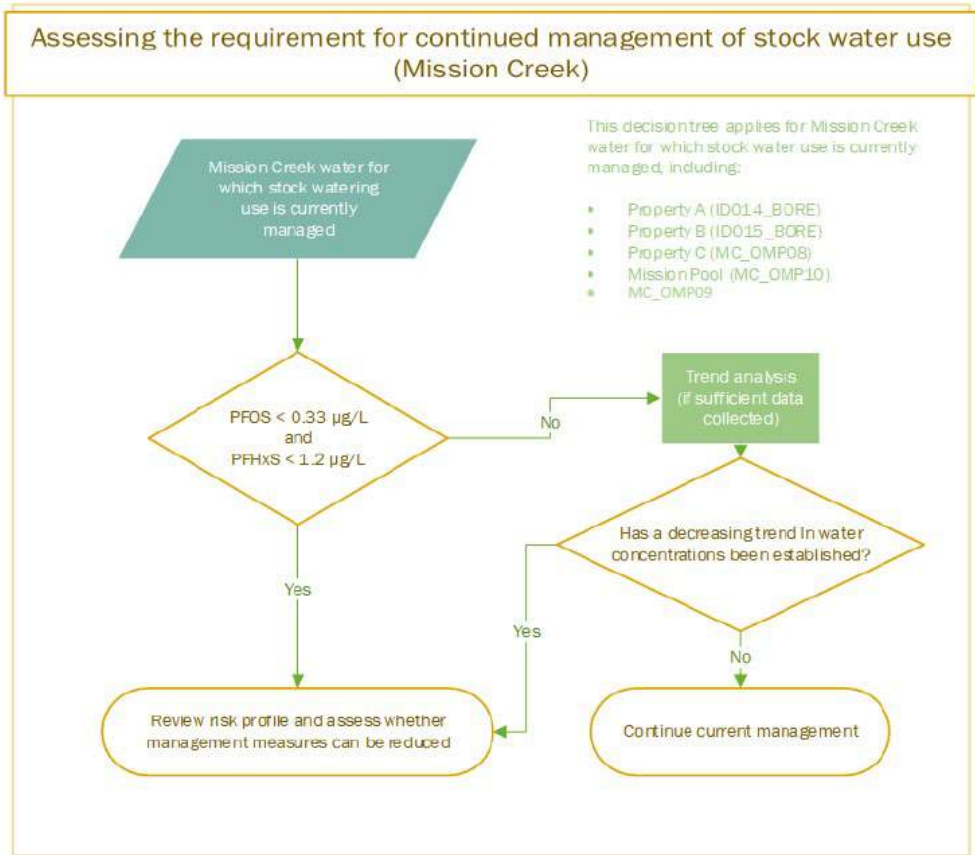
3.3.2.1 Decision Trees: Use of UTVs for Exposures Currently Assessed to be Low and Acceptable

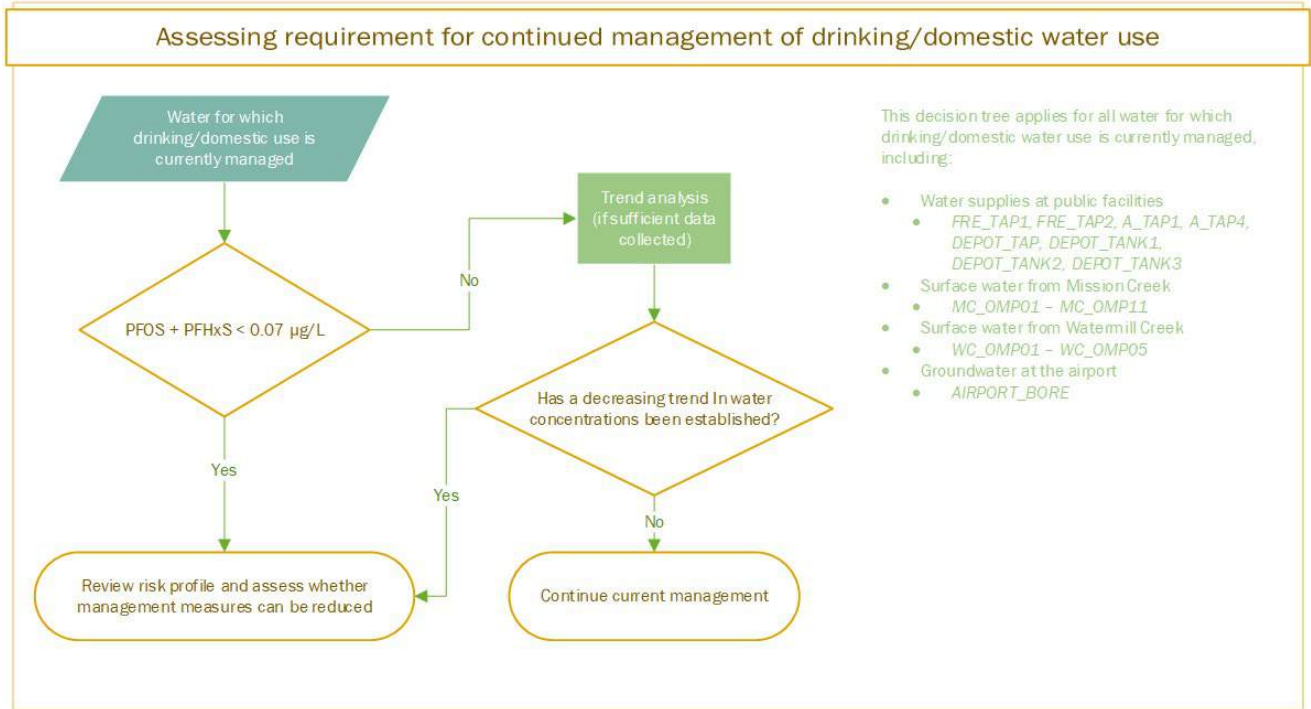






3.3.2.2 Decision Trees: Use of LTVs for Exposures Which Currently Require Management





3.3.3 Additional Screening Criteria

In addition to comparing to the UTVs/LTVs, results have also been compared to a range of screening levels to provide clarity around the potential risks associated with the range of potential exposure pathways. These screening levels have been adopted with reference to those adopted in the DSI, but (where necessary) updated to align with the latest version of the guidance, sourced from the PFAS NEMP 3.0 (HEPA, 2025), and NHMRC⁵ 2025 Australian Drinking Water Guidelines (ADWG) (as relevant), and to include project-specific screening levels where derived as part of the HHERA. These criteria are summarised in **Table 3-8**. Further information on the adopted criteria is provided in the DSI (Senversa, 2021b). The revised DWG are summarised in **Table 3-7** below. This update responds to recent PFAS toxicity studies, and the recommended values are more stringent, and more closely aligned with drinking water values in other jurisdictions.

Table 3-7: Drinking water guidelines

| PFAS | DWG (µg/L) | |
|-------|------------|-----------------------|
| | Current | Previous |
| PFOS | 0.008 | 0.07 (sum PFOS+PFHxS) |
| PFHxS | 0.03 | |
| PFOA | 0.2 | 0.56 |
| PFBS | 1 | - |

PFOA- Perfluorooctanoic acid
 PFBS- Perfluorobutanesulfonic acid

⁵ National Health and Medical Research Council



The DWG have changed as a result of updating the toxicity data (i.e. the “acceptable intake” of PFAS, which is the daily PFAS intake which is not expected to result in health effects). This updated toxicity data is potentially relevant to all PFAS exposures (e.g. food consumption, recreational water exposure and soil contact), not only to a pathway of exposure to PFAS via drinking water. However, the scope of the DWG derivation is for drinking water only; other regulatory agencies are responsible for regulation of pathways other than drinking water (e.g. Food Standards Australia and New Zealand [FSANZ] is the regulatory agency responsible for food). Agencies other than ADWG will need to review and accept the updated toxicity data before guideline values for pathways other than drinking water are updated. **The guideline values for PFAS exposures other than through drinking water have not yet been updated** to incorporate the revised toxicity data; as such the previous guideline values are currently applicable.⁶

Table 3-8: Additional Screening Criteria

| Land/Water Use | Adopted Screening Criteria | | | | | Source |
|---|--|--|---------------------|-------------|---------------------|--|
| | PFOA ¹ | PFOS | PFHxS | PFHxS+P FOS | PFBS | |
| SURFACE WATER AND GROUNDWATER | | | | | | |
| Aquatic Ecosystems⁴ | 19 µg/L (99% protection) 220 µg/L (95% protection) 632 µg/L (90% protection) | 0.00023 µg/L (99% protection) 0.13 µg/L (95% protection) 2 µg/L (90% protection) | - | - | - | HEPA (2025) |
| Primary and/or Secondary Contact Recreation | 10 µg/L | 2 µg/L ³ | 2 µg/L ³ | 2 µg/L | - | NHMRC (2019) ⁵ |
| Aesthetic Enjoyment | To be assessed based on observations of odour and/or visual amenity impact (noting that aesthetic impacts have not been noted for PFAS impacted water during site investigations). | | | | | - |
| Cultural And Spiritual Values (Indigenous and/or Non-Indigenous) | No specific guidelines available, considered that criteria for other land uses will also be protective of this use. | | | | | - |
| Drinking (Potable) Water | 0.2 µg/L | 0.008 µg/L | 0.03 µg/L | - | 1 µg/L ² | NHMRC (2025). |
| Agriculture (Stock Watering) | - | 0.33 µg/L | 1.2 µg/L | - | - | Relevant screening levels for this land use are not available. HHERA (Senversa, 2021c) derived screening levels for cattle watering adopted. These levels are also protective of chicken watering. |

⁶ It is noted that in the Public Consultation draft of the updated Australian Recreational Water Quality Guidelines (NHMRC, 2026), the NHMRC toxicity data used in the DWG derivation is referenced in the derivation of recreational screening levels for PFAS; this provides an indication that the NHMRC toxicity data may also be applicable to recreational exposure pathways in the future (i.e. in the event this guidance is finalised unchanged).



| Land/Water Use | Adopted Screening Criteria | | | | | Source |
|--|--|---|-----------------------------------|-----------------------------------|------|--|
| | PFOA ¹ | PFOS | PFHxS | PFHxS+P FOS | PFBS | |
| Irrigation | - | 1.4 µg/L | 0.8 µg/L | - | - | Relevant screening levels for this land use are not available. HHERA (Senversa, 2021c) derived screening levels for vegetable irrigation adopted, which are more stringent than fruit irrigation screening levels. |
| Aquaculture Human Consumption of Fish, Crustacea and Molluscs | As the Creeks on Norfolk Island are largely ephemeral with water flowing only during rainfall events, this land use is not considered relevant to this investigation. Furthermore, there was no evidence of human consumption of freshwater Fish, Crustacea, and Molluscs from water bodies or within the Mission Creek Catchment. | | | | | - |
| Industrial and Commercial Use | No generic screening criteria for these uses are available, however, criteria for other land uses relevant to human and animal health (including potable water supply, primary contact recreation and stock watering) are considered relevant and will be considered in assessing impacts to this land use. | | | | | - |
| SOIL AND SEDIMENT | | | | | | |
| Human Health | 0.06 mg/kg (sensitive) | Refer to PFHxS+PFOS ³ | Refer to PFHxS+P FOS ³ | 0.003 mg/kg (sensitive) | - | HEPA (2025) |
| • Sensitive Use (including growing of home produce) | 10 mg/kg (recreation / open space) | | | 1 mg/kg (recreation / open space) | | |
| • Recreation / Open Space Use | | | | | | |
| Human Health Commercial Use Industrial Use | 50 mg/kg | Refer to PFHxS+PFOS ³ | Refer to PFHxS+P FOS ³ | 20 mg/kg | - | |
| Maintenance of Ecosystems | 0.003 mg/kg (secondary poisoning / bioaccumulation) | 0.003 mg/kg (secondary poisoning / bioaccumulation) | - | - | - | |
| | 10 mg/kg (direct toxicity) | 1 mg/kg (direct toxicity) | | | | |
| Production of Food, Fibre and Flora | 0.06 mg/kg | Refer to PFHxS+PFOS ³ | Refer to PFHxS+P FOS ³ | 0.003 mg/kg | - | HEPA (2025) HIL-A sensitive use criteria adopted in absence of agricultural guidelines |
| Aesthetics | PFAS are not considered to be relevant indicators for this land use, and it has not been considered further in the DSI. | | | | | - |



| Land/Water Use | Adopted Screening Criteria | | | | | Source |
|--|---|---|-------|---|------|---|
| | PFOA ¹ | PFOS | PFHxS | PFHxS+P FOS | PFBS | |
| Buildings and Structures | PFAS are not considered to be relevant indicators for this land use, and it has not been considered further in the DSI. However, PFAS is reported to adsorb and desorb from permeable materials such as concrete, potentially representing source of PFAS that should be considered in the conduct of the works. | | | | | - |
| BIOTA | | | | | | |
| Food for Human Consumption - Fruits | 0.0051 mg/kg | 0.001 mg/kg (0.0006 mg/kg, however LOR 0.001 mg/kg) | - | 0.001 mg/kg (0.0006 mg/kg, however LOR 0.001 mg/kg) | - | FSANZ (2017) Trigger Points, which are conservative screening levels and not regulatory limits. |
| Food for Human Consumption - Vegetables | 0.0088 mg/kg | 0.0011 mg/kg | - | 0.0011 mg/kg | - | FSANZ (2017) Trigger Points, which are conservative screening levels and not regulatory limits. |

Table Notes:

1. PFOA- perfluorooctanoic acid.
2. PFBS has been included as a criterion is now available, however it is not considered a CoPC at this site.
3. Where the criteria refer to the sum of PFOS and PFHxS, this means concentrations of PFOS only, PFHxS only, and the sum of the two (HEPA, 2025).
4. For bioaccumulative contaminants, which include many PFAS, the framework specifies that the 99% species protection DGV should be used in:
 - assessing toxicity and bioaccumulation in high conservation value ecosystems
 - assessing bioaccumulation in slightly to moderately disturbed ecosystems.
 Similarly, the 95% species protection DGV has been applied to highly disturbed systems.
5. It is noted that in the Public Consultation draft of the updated Australian Recreational Water Quality Guidelines (NHMRC, 2026), the NHMRC toxicity data used in the DWG derivation is referenced in the derivation of recreational screening levels for PFAS; this provides an indication that the NHMRC toxicity data may also be applicable to recreational exposure pathways in the future (i.e. in the event this guidance is finalised unchanged).



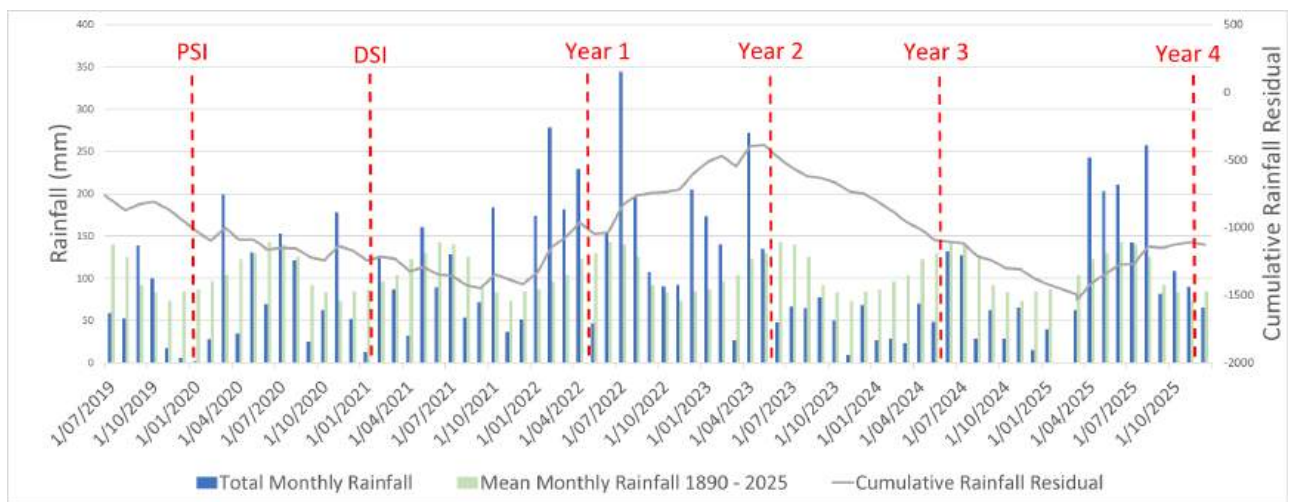
4.0 Results

4.1 Surface and Groundwater Results

4.1.1 Rainfall

Rainfall conditions and the presence of surface water varied significantly between the PSI sampling in January 2020 and the OMP monitoring period. **Figure 4-1** below displays the total monthly rainfall between 2019 and 2025 and rainfall residual. Above average rainfall was recorded prior to the Year 1, Year 2 and Year 4 monitoring events; below average rainfall was recorded prior to the PSI, DSI and Year 3 monitoring events.

Figure 4-1: Total Monthly Rainfall



Rainfall can be expected to have competing impacts on observed PFAS concentrations, including:

- PFAS concentration increase where high rainfall infiltrates PFAS in soil source areas and allows increased migration of PFAS to surface water and groundwater.
- PFAS concentration increase where low rainfall results in evaporation of stagnant water bodies, concentrating PFAS in residual water.
- PFAS concentration decrease where high rainfall dilutes PFAS concentrations in surface water and groundwater.

Upper and lower Mission Creek PFHxS+PFOS concentrations are graphed with total rainfall recorded in the preceding month below. There appears to be a very weak positive correlation (R^2 0.011 to 0.14) between increased rainfall and increased PFHxS+PFOS concentration in the majority of locations. A weak to moderate negative correlation (R^2 0.27 to 0.33) is apparent closest to source areas at WWII_DAM and the Airport Bore. Overall, rainfall is not assessed to have a significant impact on PFAS concentrations monitored.

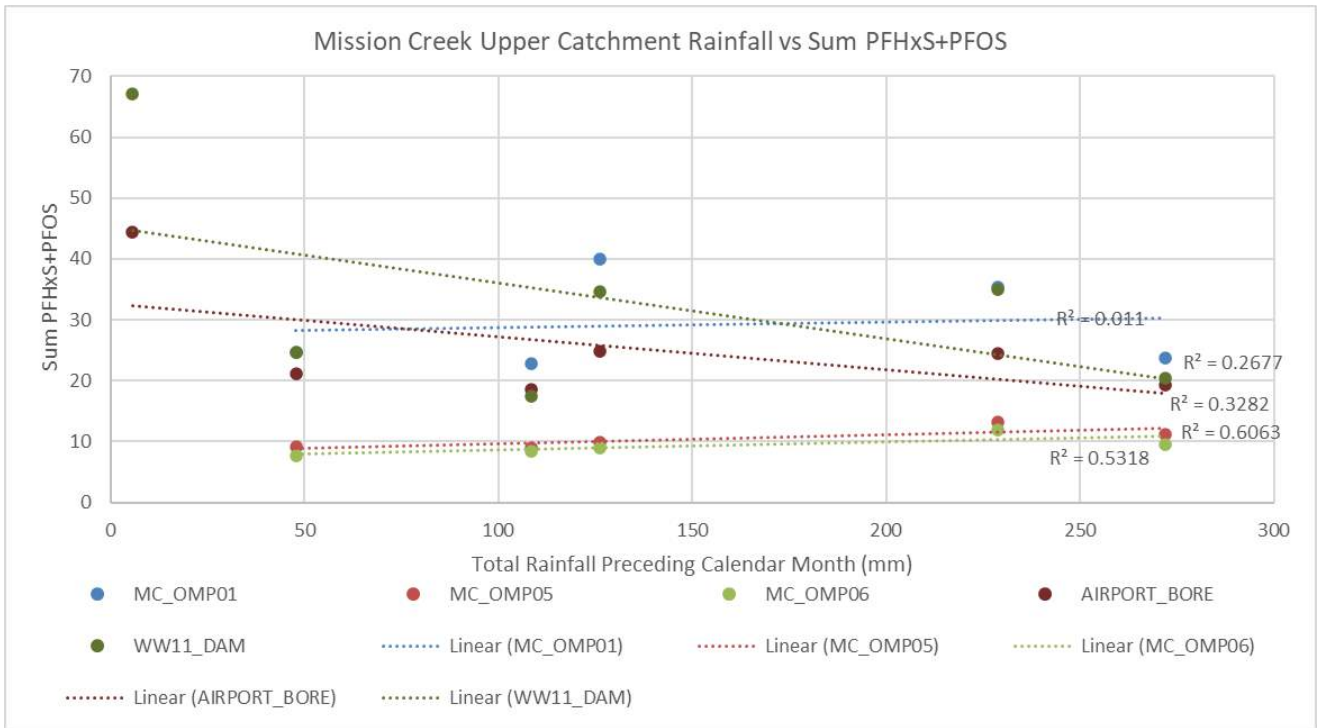


Figure 3: Rainfall Vs PFAS Concentration Upper Mission Creek

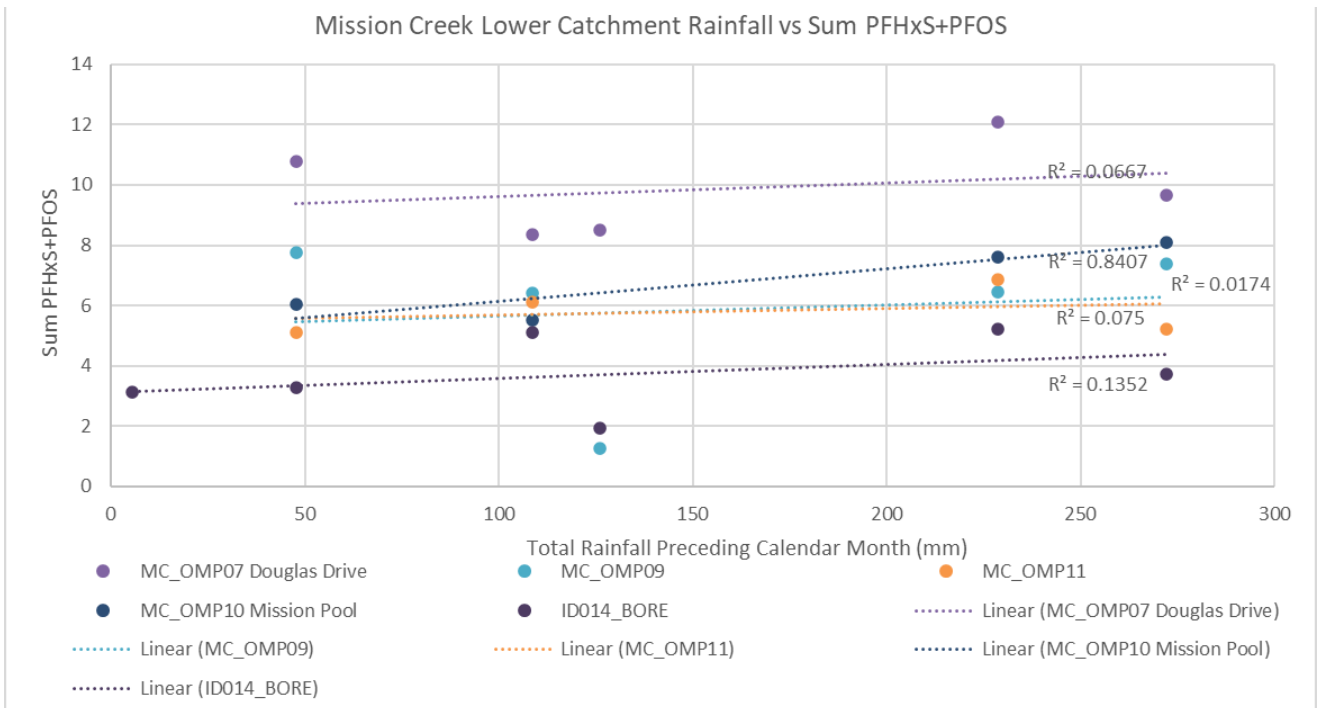


Figure 4: Rainfall Vs PFAS Concentration Lower Mission Creek



4.1.2 Surface Water and Groundwater Field Observations

Water was present at all sampling locations during the monitoring event. Surface water and groundwater field observations are presented in **Appendix C** and summarised in **Table 4-1** below. It is noted that a number of samples were opportunistically collected and field observations recorded in **Appendix C**, however laboratory analysis was not completed for all of these samples. This data has been included in **Appendix C** for completeness.

Table 4-1: Summary of Water Field Observations

| Sample location | | Mission Creek ¹ | Watermill Creek ² | Headstone Creek ³ | Cascade Creek ⁴ | Ground-Water ⁵ | Public Taps and Tanks ⁶ |
|---|---------------|----------------------------|------------------------------|------------------------------|----------------------------|---------------------------|------------------------------------|
| Dissolved Oxygen (DO) [mg/L] | Min | 0.2 | 0.4 | - | - | 3.2 | 4.5 |
| | Max | 7.2 | 8.3 | - | - | 7.7 | 7.8 |
| | Median | 5.3 | 3.9 | 0.7 | 7.0 | 5.5 | 6.1 |
| Electrical Conductivity (EC) [micro siemens per centimetre [µS/cm]] | Min | 348 | 253 | - | - | 208 | 17 |
| | Max | 661 | 508 | - | - | 819 | 735 |
| | Median | 414 | 358 | 457 | 357 | 491 | 32 |
| pH | Min | 5.9 | 5.9 | - | - | 4.7 | 5.2 |
| | Max | 7.0 | 6.7 | - | - | 6.5 | 7.3 |
| | Median | 6.5 | 6.4 | 6.4 | 6.5 | 5.6 | 6.2 |
| Redox Potential [millivolts [mV]] | Min | -465 | -373 | - | - | -292 | -366 |
| | Max | -129 | -228 | - | - | -69 | 31 |
| | Median | -171 | -279 | -260 | -264 | -205 | -253 |
| Temperature [°C] | Min | 19 | 19 | - | - | 21 | 13 |
| | Max | 26 | 23 | - | - | 23 | 29 |
| | Median | 21 | 20 | 22 | 21 | 21 | 23 |

Table notes:

1. Mission Creek: MC_OMP01 to 07, MC_OMP09 to 11, WW11_DAM and ID014_BORE.
2. Watermill Creek: WC_OMP01 to 5.
3. Headstone Creek: PWS_HEAD_DAM.
4. Cascade Creek: Cockpit_SW01.
5. Groundwater: AIRPORT_BORE, ID013_BORE, ID016_BORE, ID019_BORE, WC-03_BORE and WC-04.
6. Public taps and tanks excluding those supplied by the Airport Bore: FRE_TAP2, public toilets, A_TAP4.



Recorded surface water field parameters were generally consistent with the previous monitoring event in 2024. Surface water generally displayed aerobic conditions and an EC within the expected range for fresh water. The pH was neutral to slightly acidic. The redox potential was more reducing than previously recorded, generally moderately reducing.

The fire station kitchen tap recorded an elevated EC of 735 us/cm, consistent with the change in supply to desalinated water.

Groundwater parameters were generally consistent with the previous monitoring event in 2024. Groundwater generally displayed aerobic conditions and low salinity (EC between 208 and 819 $\mu\text{S}/\text{cm}$). The pH was slightly acidic (4.7 to 6.5) and moderately reducing conditions were recorded.

4.1.3 Surface Water and Groundwater Laboratory Results

Surface water and groundwater analytical results have been compared against the adopted upper and lower trigger values to assess whether the currently selected management actions should change. Results are presented in appended **Tables 1 to 6** and summarised in **Table 4-2** and **Table 4-3** below. Water concentrations are mapped on appended **Figures 3** and **4**.

Table 4-2: Water UTV Results

| Sampling Medium | Upper Trigger Value ($\mu\text{g}/\text{L}$) | | | No. Samples | No. Detections | No. Above UTV |
|---|--|-------|------------|-------------|--------------------------------------|---------------|
| | PFOS | PFHxS | PFHxS+PFOS | | | |
| Mission Creek Catchment Groundwater Used for Irrigation: ID013_BORE, ID016_BORE | 4.2 | 2.5 | - | 2 | PFOS: 0 PFHxS: 1 PFHxS+PFOS: 1 | 0 |
| Surface Water from Other Creeks: WC_OMP01 to 05 | 0.5 | 1.3 | - | 5 | PFOS: 5 PFHxS: 5 PFHxS+PFOS: 5 | 0 |
| Surface Water from Cascade Creek / Headstone Creek: Cockpit_SW01, PWS_HEAD_DAM | - | - | 0.07 | 2 | PFOS: 1 PFHxS: 1 PFHxS+PFOS: 1 | 0 |

Table 4-3: Water LTV Results

| Sampling Medium | Lower Trigger Value ($\mu\text{g}/\text{L}$) | | | No. Samples | No. Detections | No. Below LTV |
|---|--|-------|------------|-------------|----------------|---------------|
| | PFOS | PFHxS | PFHxS+PFOS | | | |
| Reticulated Water Supplies at Public Facilities: A_TAP1, A_TAP4, A_TAP5, A_TAP15, FRE_TAP1, FRE_TAP2, FRE_TAP6, DEPOT_TAP, DEPOT_TANK1 and 2, COUNCIL_TAP1, hospital taps, WASTE_TAP, ELEC_TAP, water carting water Surface Water from Mission Creek: MC_OMP01 to 07a and MC_OMP09 to 11, WW11_DAM Surface Water from Watermill Creek: WC_OMP01 to 05 Groundwater at Airport: AIRPORT_BORE | - | - | 0.07 | 37 | PFHxS+PFOS: 21 | 20 |



| Sampling Medium | Lower Trigger Value (µg/L) | | | No. Samples | No. Detections | No. Below LTV |
|--|----------------------------|-------|------------|-------------|---------------------|---------------|
| | PFOS | PFHxS | PFHxS+PFOS | | | |
| Mission Creek Water Used for Cattle Stock Watering Prior to Management: ID014_BORE, ID019_BORE, MC_OMP09 and 10. | 0.33 | 1.2 | - | 4 | PFOS: 4 PFHxS: 4 | 1 |
| Mission Creek Water Used for Chicken Stock Watering Prior to Management: ID013_BORE | 0.9 | 1.3 | - | 1 | PFOS: 0 PFHxS: 0 | 1 |

Laboratory results have also been compared against the additional screening criteria, presented in **Table 7** (Year 4 and historical data). Laboratory certificates of analysis are provided in **Appendix D**. A summary of detections and exceedances of the drinking water and recreational water use criteria is provided below.

Table 4-4: Water Additional Screening Criteria Results

| Sample location | Assessment Criteria (µg/L) | Number of locations Detected / Number of locations Exceeding Criteria | | | | | | |
|-----------------------------|----------------------------|--|-------------------------------------|-------------------------------------|-----------------------------------|----------------------------------|--|-----|
| | | Mission Creek ¹ n=12 | Watermill Creek ² n=5 | Headstone Creek ³ n=1 | Cascade Creek ⁴ n=1 | Ground-Water ⁵ n=6 | Public Taps and Tanks ⁶ n=16 | |
| Drinking Water – Health | PFOA | 0.2 | 12/6 | 1/0 | 0/0 | 0/0 | 1/1 | 0/0 |
| | PFOS | 0.008 | 12/12 | 5/5 | 0/0 | 1/1 | 3/2 | 0/0 |
| | PFHxS | 0.03 | 12/12 | 5/5 | 0/0 | 1/0 | 4/3 | 0/0 |
| | PFBS | 1 | 12/0 | 2/0 | 0/0 | 0/0 | 3/0 | 0/0 |
| Recreational Water - Health | PFOA | 10 | 12/0 | 1/0 | 0/0 | 0/0 | 1/1 | 0/0 |
| | PFHxS+PFOS | 2 | 12/10 | 5/0 | 0/0 | 1/0 | 4/1 | 0/0 |

Table notes:

n: number of samples analysed.

1. Mission Creek: (12 samples) MC_OMP01 to 07, MC_OMP09 to 11, WW11_DAM and ID014_BORE.

2. Watermill Creek: (5 samples) WC_OMP01 to 5.

3. Headstone Creek: (1 sample) PWS_HEAD_DAM. The LOR analysed at this location was above the DWG for PFOS.

4. Cascade Creek: (1 sample) Cockpit_SW01.

5. Groundwater: (6 samples) AIRPORT_BORE, ID013_BORE, ID016_BORE, ID019_BORE, WC-03_BORE and WC-04.

6. Public taps and tanks excluding those supplied by the Airport Bore FRE_TAP2, public toilets, A_TAP4 (16 samples).

4.2 Soil and sediment Results

Soil and sediment laboratory results have been compared against the additional screening criteria, presented in **Table 8**. Laboratory certificates of analysis are provided in **Appendix D**. Discussion on the findings is presented in **Section 5.0**.



4.3 Biota Results

Biota laboratory results have been compared against the additional screening criteria, presented in **Table 9**. Laboratory certificates of analysis are provided in **Appendix D**. Discussion on the findings is presented in **Section 5.0**.

4.4 Quality Assurance and Quality Control

The data quality assurance and quality control (QAQC) procedures adopted by Senversa provide a consistent approach to evaluation of whether the data quality objectives required by the project have been achieved. The process focuses on assessment of the useability of the data in terms of accuracy and reliability in forming conclusions on the condition of the element of the environment being investigated.

The methodology and results of the QAQC assessment are presented in **Appendix B**.

While a small number of results were outside specified acceptance criteria, these were not considered to significantly impact the quality or representativeness of the data, and the majority of results indicated that the precision and accuracy of the data was within acceptable limits. The results are therefore considered to be representative of chemical concentrations in the environmental media sampled at the time of sampling, and suitable to be used for their intended purpose in forming conclusions relating to the contamination status of water, soil, sediment and biota.

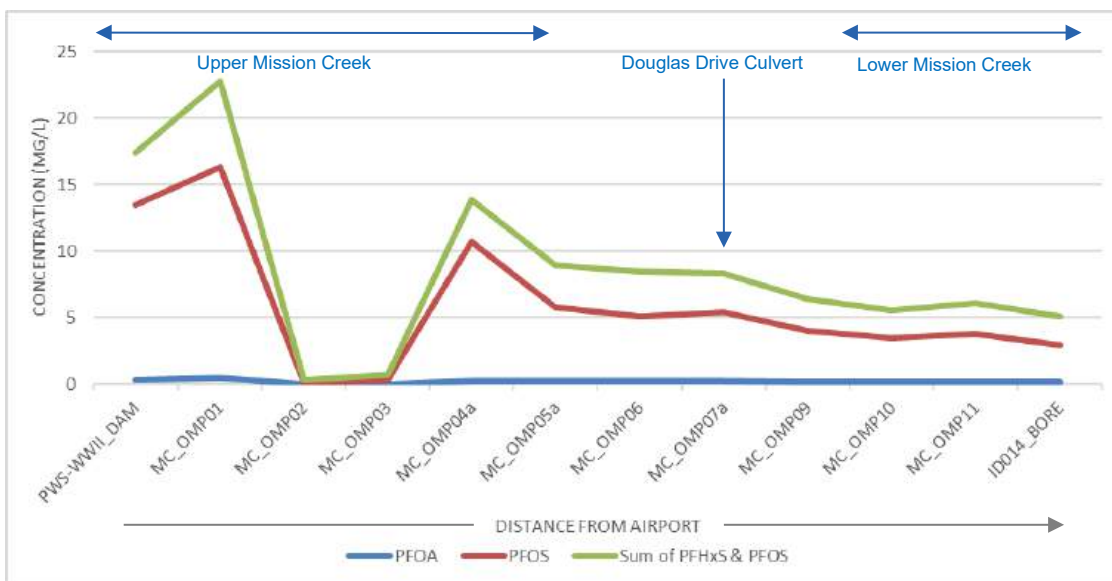


5.0 Findings

5.1 Nature and Extent of PFAS in Creek Water

5.1.1 Mission Creek

PFAS were detected in all Mission Creek samples analysed, consistent with historical results. Surface water samples from the Mission Creek catchment showed the highest concentrations at locations closest to source zones at the Airport (WWII Dam and MC_OMP01) and generally decreased with distance from the Airport, consistent with historical results. MC_OMP02 and MC_OMP03 were the exception, which reported low levels of PFAS due to river confluence with less contaminated streams, as noted in previous monitoring events. PFOA, PFOS and PFHxS+PFOS concentrations along



Mission Creek with distance from the Airport are shown on **Figure 5-1** below.

Figure 5-1: Surface Water PFAS Concentrations in Mission Creek



5.1.1.1 Mission Creek Temporal Trends

Mission Creek PFHxS+PFOS concentrations from 2020 to 2025 are plotted on **Figure 5-2** and **Figure 5-3** below.

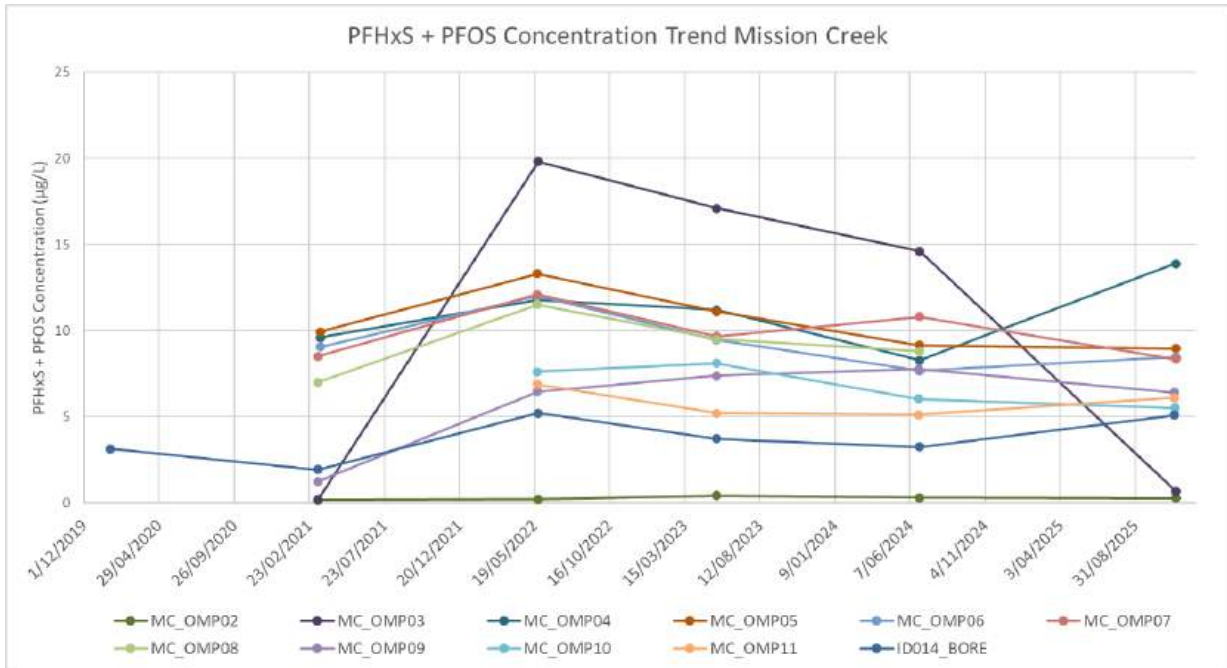


Figure 5-2: Mission Creek PFHxS+PFOS Concentration Trends

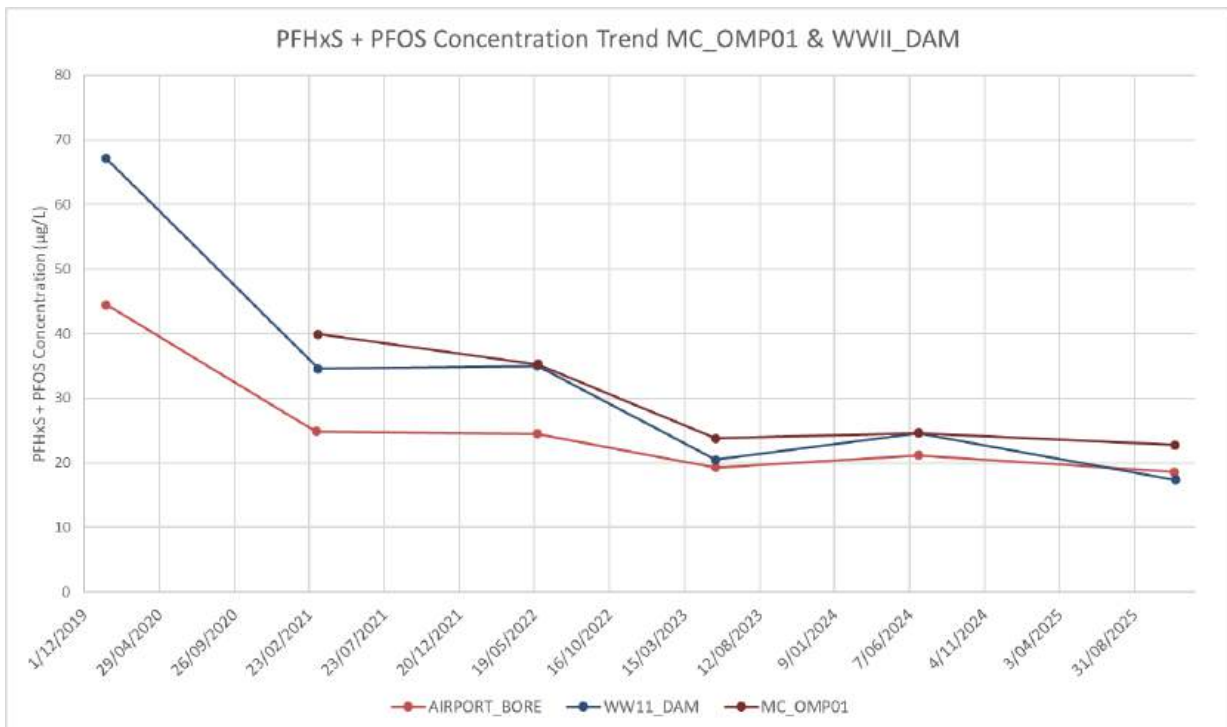


Figure 5-3: MC_OMP01 & WWII_DAM PFHxS+PFOS Concentration Trends

The decrease in PFAS concentrations at MC_OMP03 relative to 2024 is considered to be due to the high rainfall prior to the monitoring event. This causes connection of the braided river streams in this section of the creek from the less contaminated stream behind the waste management centre and the more contaminated stream originating at the WWII_DAM sample location.



A Mann Kendall trend analysis was undertaken on the data collected to-date to assess statistically significant changes in concentrations over time, presented in **Appendix E**. In general, the Mission Creek data showed no significant upwards or downwards trends in PFOS, PFOA or PFHxS+PFOS concentrations between 2020 and 2025. However, a statistically significant downwards trend was noted for these three analytes at the WWII Dam and Airport Bore, as well as PFOA at MC_OMP07 and PFHxS+PFOS at MC_OMP01. Trends are generally more discernible at higher concentration ranges due to a reduction in measurement uncertainty and the broader numerical range. The observed downwards trends in the upper portion of Mission Creek where the highest PFAS concentrations have been reported are consistent with this pattern.

The primary AFFF product historically in use at the airport from 1980 to 2004 was reportedly 3M Lightwater™, followed by Ansulite® until 2015 (Senversa, 2021a). Both compounds are known to contain several PFAS compounds, including PFOS and PFOA, however Ansulite® contains trace amounts (AirServices, 2007). Consistent with this reported historical use, in Australian sites where legacy AFFF has been utilised (including on Norfolk Island Airport), PFOS and PFHxS are predominantly the most widespread PFAS detected. The observed reduction in PFOS, Sum PFHxS+PFOS and PFOA concentrations is therefore consistent with cessation of AFFF use on-site and contaminant mass reduction in the source areas as rainfall flushes through the system over time.

5.1.1.2 Mission Creek Assessment Against Screening Criteria

The Year 4 monitoring event PFAS concentrations in Mission Creek remained above the LTVs (indicating that current management measures around the use of water for drinking / domestic use and stock watering should continue). No UTV have been set as management measures are required.

No change to current management actions is required based on assessment of the Year 4 monitoring results against these trigger values, i.e. management measures and/or further assessment are required.

All Mission Creek samples analysed exceeded the drinking water assessment criteria. All but two samples also exceeded the recreational water use and vegetable irrigation assessment criteria. Current use of Mission Creek water for these purposes has not been identified.

5.1.2 Watermill Creek

Within the Watermill Creek catchment, the highest PFAS concentration in surface water (WC_OMP01, PFHxS+PFOS: 1.01 µg/L) was identified immediately downstream of the Airport maintenance depot, consistent with the previous monitoring event. This concentration remained above the adopted PFOS human health drinking water criterion of 0.008 µg/L, but below the criteria for recreational use (2 µg/L). PFAS concentrations decreased with distance from the Airport, as indicated on **Figure 5-4** below.

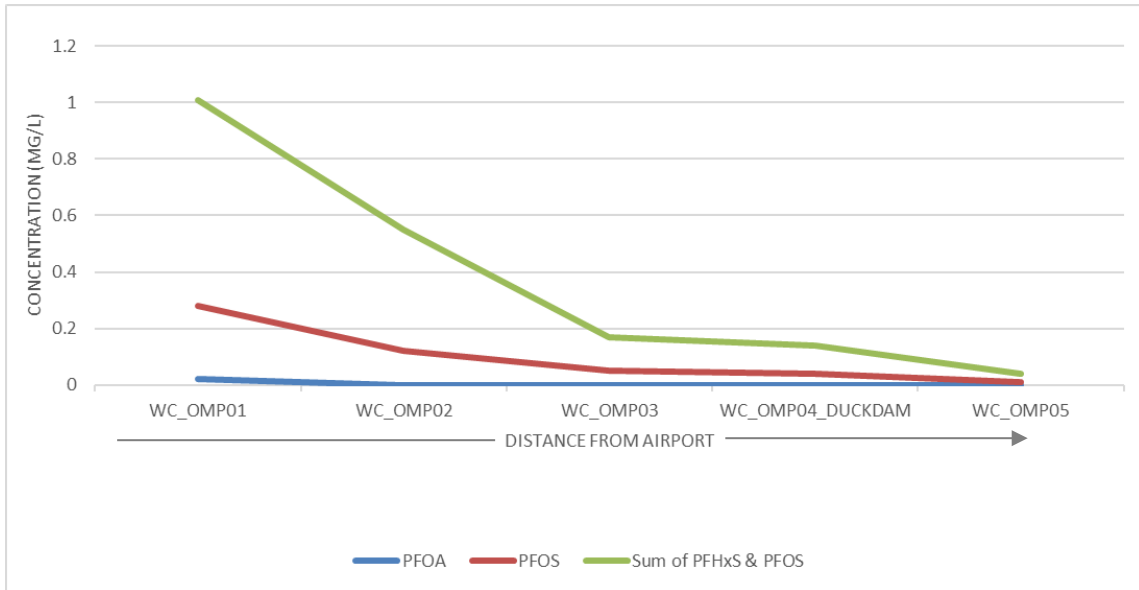


Figure 5-4: Surface Water PFAS Concentrations in Watermill Creek

5.1.2.1 Watermill Creek Temporal Trends

Watermill Creek PFHxS+PFOS concentrations from 2020 to 2025 are plotted on **Figure 5-5** below.

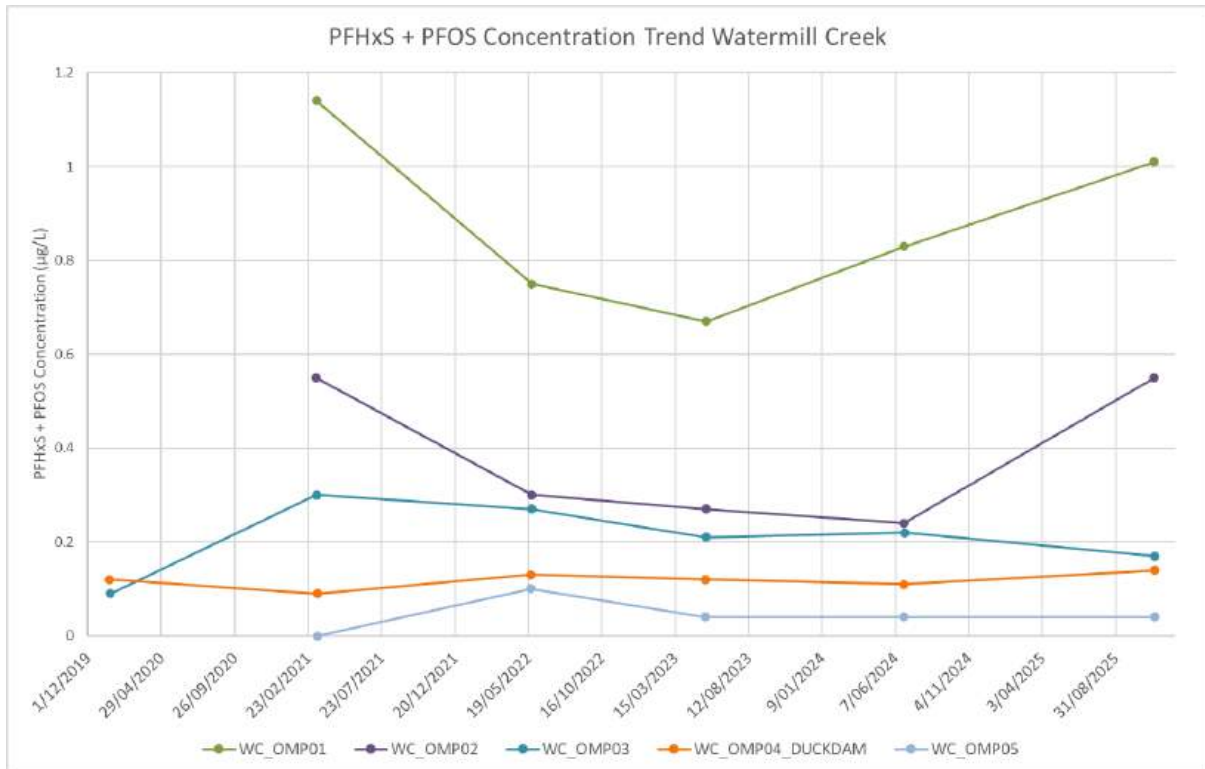


Figure 5-5: Watermill Creek PFHxS+PFOS Concentration Trends



A Mann Kendal trend analysis was undertaken on the data collected to-date to assess statistically significant changes in concentrations over time, presented in **Appendix E**. Reported PFAS concentrations in Watermill Creek displayed no statistically significant temporal trends. However, the time plot above indicates PFAS concentrations closest to the airport (WC_OMP01 and 2) increased between 2024 and 2025. The concentration of PFHxS+PFOS also marginally increased in Duck Dam and was the highest recorded to-date. Consultation with NIRC including the airport and fire station has not indicated a change in site management that may have resulted in an increased PFAS flux into Watermill Creek and it is noted that the 2025 reported PFAS concentrations remain within the same order of magnitude as historical results. Ongoing monitoring of Watermill Creek is recommended to monitor this trend.

5.1.2.2 Watermill Creek Assessment Against Screening Criteria

The Year 4 monitoring event PFAS concentrations in Watermill Creek remained below the UTVs (indicating that no additional management measures are required) and generally above the LTVs (indicating that current management measures around the use of water for drinking / domestic use should continue). Specifically, as concentrations remain below the UTVs, this indicates that the risks from other water uses (including stock watering and produce irrigation) remain low and acceptable.

No change to current management actions is required based on assessment of the Year 4 monitoring results against these trigger values.

All Watermill Creek samples analysed exceeded the drinking water assessment criteria, however, no samples exceeded the recreational water use, vegetable irrigation or stock watering assessment criteria.

5.1.3 Cascade Creek and Headstone Creek

Cockpit_SW01 reported a low-level concentration of PFOS (0.01 µg/L) and PFHxS+PFOS (0.02 µg/L). These concentrations were higher than the 2023 results, when PFAS was not detected, however are consistent with results reported in other years since 2020. The reported concentrations were above the PFOS 99% ecological protection guideline value (0.00023 µg/L) and DWG (0.008 µg/L) but remained below the recreational criterion (2 µg/L). It is noted that the creek waters are not utilised for drinking.

PWS_HEAD_DAM PFAS concentrations were reported below the LOR, sampled from the end of Headstone Creek. This is a reduction from low level PFOS previously reported in 2020 and is consistent with the 2022 to 2024 results.

The Year 4 monitoring event PFAS concentrations reported in both Cascade and Headstone Creeks were below the UTVs and no implementation of management actions is required based on assessment against these trigger values.



5.2 Nature and Extent of PFAS in Groundwater

5.2.1 Groundwater Temporal Trends

The reported concentration of PFHxS+PFOS in groundwater collected from both the Airport Bore and ID_016 bore located in the Mission Creek catchment decreased in 2025 relative to 2024. PFAS was reported below the laboratory LOR in ID013 bore consistent with previous years. No access was provided to ID015 bore, however ID019 bore was sampled for the first time in the lower Mission Creek Catchment. PFOS and PFHxS were detected in ID019_BORE above the DWG, but below the recreational water use criteria, broadly consistent with concentrations previously reported in ID015 bore. An overall decreasing trend is indicated at the Airport Bore in **Figure 5-6**.

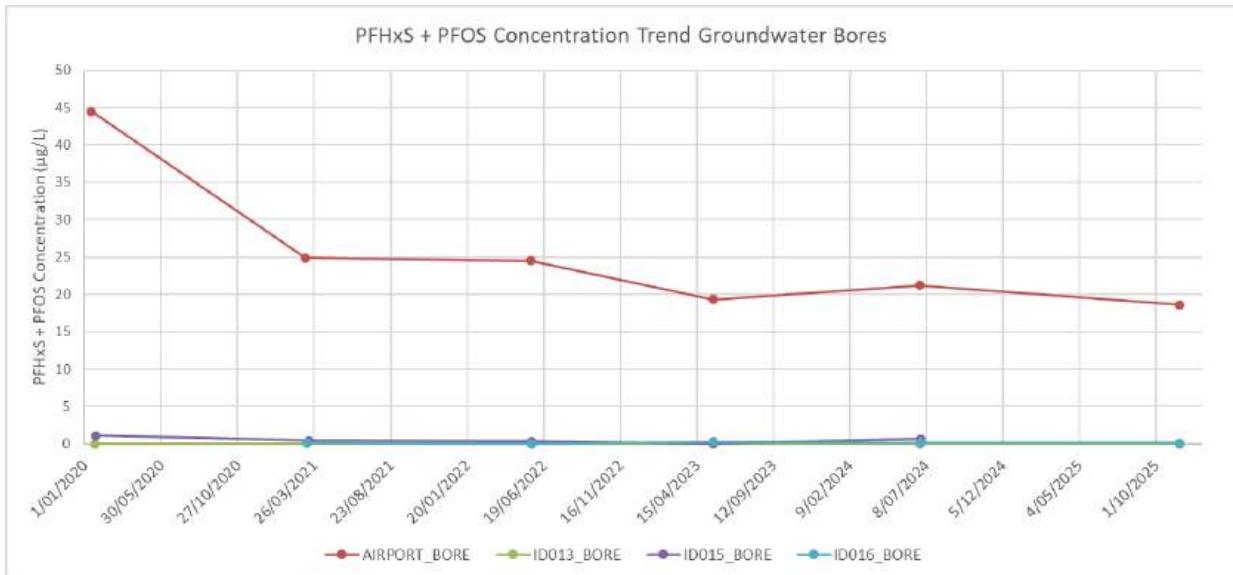


Figure 5-6: Groundwater PFHxS+PFOS Concentration Trends

Mann Kendal trend analysis results for the Airport Bore are discussed in **Section 5.1.1** and included in **Appendix E**. Other groundwater data showed no significant upwards or downwards trends in PFOS, PFOA or PFHxS+PFOS concentrations between 2020 and 2025.

Groundwater-surface water connectivity has not been directly assessed in the Mission Creek Catchment as part of the PFAS investigation. It is noted that the upper section of the Mission Creek Catchment from the airport (WWII_DAM) to Douglas Drive (MC_OMP06) has maintained flow during investigations since 2020. Conversely, the lower section from Douglas Drive to the creek's discharge at the ocean was largely dry during PSI and DSI investigations, with some small sections of ponded, discontinuous water. These observations indicate that the upper section of the catchment may act as a gaining creek (groundwater predominantly discharges to the creek), and the lower section a losing creek (groundwater is predominately recharged by the creek), in particular around Mission Pool. This is consistent with the observed concurrent decrease in PFAS concentrations in both the upper section of Mission Creek (WWII_DAM and MC_OMP01) and the Airport Bore.

5.2.2 Groundwater Assessment Against Screening Criteria

Groundwater assessment against the screening criteria is provided in **Section 5.3** below, relevant to each point of use.



5.3 Nature and Extent of PFAS in Point of Use Water

5.3.1 Water Used for Irrigation

During the 2020 and 2021 monitoring events, PFAS was identified in a private water supply ID013_SW01 (pumped from Mission Creek) used for fruit and vegetable produce irrigation. In monitoring from 2023 to 2025, the pump for ID013_SW01 was no longer in use for irrigation, replaced by groundwater bore ID013_BORE and rainwater. Consistent with previous results, the 2025 PFAS concentrations in ID013_BORE were reported below the LOR.

PFAS was also detected in private Mission Creek catchment groundwater bore ID016_BORE, used for fruit and vegetable irrigation. Reported Sum PFHxS+PFOS concentrations in ID016_BORE have ranged from 0.02 µg/L to 0.29 µg/L between March 2021 and November 2025, reported below the drinking water screening criteria in 2022, and above in the remaining monitoring rounds. In 2025, the concentration of PFHxS+PFOS decreased relative to the 2024 concentration, however no statistically significant concentration trend was observed (**Appendix E**).

The November 2025 reported PFAS concentrations in water used for irrigation at these two locations were below the UTVs for irrigation pathways. These water sources can continue to be used for produce irrigation without management measures based on the current monitoring results.

Water from other creeks may also be used for irrigation. PFAS concentrations in 2025 in other creeks also remained below the UTVs for Watermill, Headstone and Cascade Creeks, protective of this use. As such, the risks of irrigation from creeks other than Mission Creek are assessed to remain low and acceptable, and management is not required.

Screening of Mission Creek water against the HHERA derived vegetable irrigation screening levels indicates this water remains unsuitable for this use. Both PFOS and PFHxS exceeded the screening levels along the length of the creek (with the exception of MC_OMP02 and MC_OMP03).

5.3.2 Water Used for Chicken Watering

PFAS concentrations in ID013_SW01 remained above the LTVs for chicken watering in May 2022 when last monitored, consistent with previous monitoring rounds, although it is noted that the concentrations had increased in 2021. The risk assessment completed as part of the HHERA (based on the previously measured concentrations) indicated that management measures are required to manage the potential exposures of consumers of chicken eggs where chickens drink the water from ID013_SW01. **The requirement for management remains unchanged based on the currently measured concentrations in water pumped from Mission Creek.** As the landholder indicated that since May 2022 this water is no longer used for chicken watering, this pathway is currently managed via the use of rainwater and/or bore water. Continued management via use of rainwater and/or bore water is required.

From May 2023, ID013_BORE has been sampled as a replacement location for ID013_SW01. Reported PFAS concentrations in ID013_BORE were below the LOR in January 2020 and remained below the LOR in May 2023 to 2025. Given the consistent below LOR results reported at ID013_BORE, the need for ongoing monitoring should be assessed in the PMP update.

Screening of surface water in other creeks against the HHERA derived stock watering screening levels (protective of chicken watering) indicates this water remains suitable for this use.



5.3.3 Water Used for Cattle Stock Watering

There are several properties in the Mission Creek catchment where water impacted by PFAS is known to be used, or is potentially used, for watering cattle. Additionally, there are properties in the Watermill / Town Creek catchment where surface water impacted by PFAS is potentially used for watering cattle.

Mission Creek catchment stock watering PFHxS+PFOS concentration trends are indicated on **Figure 5-7** below.

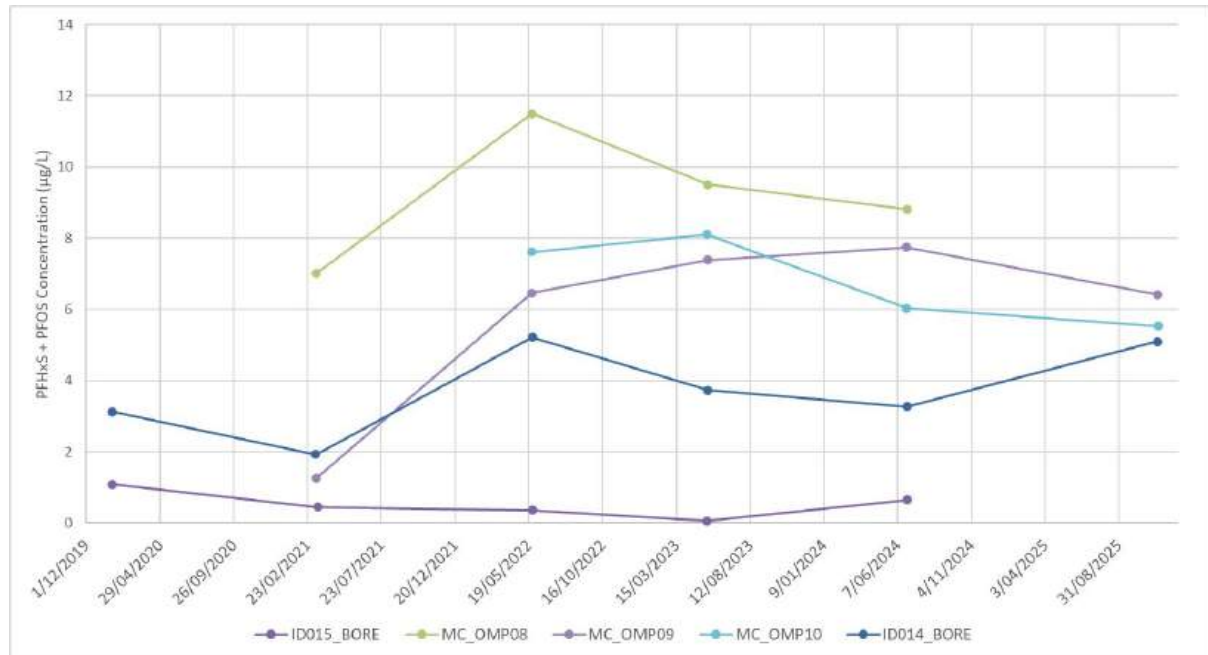


Figure 5-7: Mission Creek Catchment Stock Watering PFHxS+PFOS Concentration Trends

The risk assessment completed as part of the HHERA (based on the previously measured concentrations from 2021) indicated that management measures are required to manage the potential exposure of home consumers or public consumers of cattle products where the cattle have access to Mission Creek water for drinking. The November 2025 reported PFAS concentrations in surface water used for stock watering remained above the LTVs, consistent with previous years. **The requirement for management therefore remains unchanged based on the currently measured concentrations.**

Further to this requirement, PFAS concentrations have increased in a number of cattle watering locations since the HHERA in 2021. The continued measurement of concentrations above those measured in 2021 may indicate an increased PFAS exposure potential for consumers of cattle products, emphasising the requirement for management for both home consumers and public consumers.

Mission Creek Catchment groundwater at ID019, and ID015 when last measured in 2024, were below the LTVs for both PFOS and PFHxS. The ID015 bore PFOS concentration was reported above the Mission Creek stock watering LTV (0.33 µg/L) in 2020 and since then remained below this level. A reduction in management measures at this location should be considered in the updated PMP. No stock watering management is recommended at ID19 bore.

Water from Watermill Creek is also understood to be used for cattle watering. The HHERA assessed the risks from the use of water from Watermill Creek for stock water to be low and acceptable, and that management measures for this use were therefore not required. Concentrations in 2025 remained below the UTVs protective of this use. As such, the risks are assessed to remain low and acceptable, and management for stock watering in the Watermill Creek catchment is not required.



5.3.4 Water Used for Drinking

The laboratory results of drinking water sources are compared to historical results in the table below. Concentrations above the 2025 DWG are indicated in red. As noted in **Section 3.3**, the LTV for domestic water use reference the 2020 NEMP HBGV and these values have been retained for consistency across the monitoring events. To assess the suitability of water use for drinking, the revised 2025 DWG have been applied.

Reference should be made to **Section 2.4.2** for more information on water sources and information collection.

Table 5-1: Comparison of Historical and 2025 PFAS Concentrations in Drinking Water

| Area / Sample Location | Historical Water Source | Max Historical Concentration PFOS (µg/L) ¹ | Current Water Source | Most Recent Concentration PFOS (µg/L) Nov 2025 | Comments |
|--|-------------------------|---|----------------------|--|--|
| Airport Departure Lounge Fountain (A_TAP15) | Airport Bore | NM | Rainwater Tanks | <0.002 | Historically sourced from Airport Bore, now sourced from new airport rainwater tanks. Plumbing has not been replaced but appears to have been effectively flushed. |
| Airport Maintenance Shed Kitchen (A_TAP4) | Airport Bore | 15.2 | Airport Bore | 0.148 | Airport bore water may still be connected, however non-use warnings in place and the kitchen was not in use during the 2025 monitoring event. Continues to exceed DWG. |
| Fire Station kitchen (FRE_TAP1) | Airport Bore | 5.49 | Desalinated water | <0.002 | Temporarily supplied by desalinated water in new plumbing. Planned future connection to relined rainwater tanks. |
| EMNI Kitchen Tap (FRE_TAP6) | Airport Bore | NM | Rainwater Tanks | <0.002 | It is understood from discussions with NIRC that water is supplied by the relined rainwater tanks noted above. |
| Waste Centre Kitchen Tap (WASTE_TAP1) | Airport Bore | NM | Rainwater Tanks | <0.002 | Airport Bore water no longer connected. Currently connected to new rainwater tanks. Plumbing not replaced, but appears to have been effectively flushed. |
| Electricity Depot Kitchen Tap (ELEC_TAP) | Airport Bore | NM | Rainwater Tanks | <0.002 | Airport Bore water no longer connected. Currently connected to new rainwater tanks. Plumbing not replaced, but appears to have been effectively flushed. |



| Area / Sample Location | Historical Water Source | Max Historical Concentration PFOS (µg/L) ¹ | Current Water Source | Most Recent Concentration PFOS (µg/L) Nov 2025 | Comments |
|--|-------------------------|---|----------------------|--|--|
| BoM Station Kitchen Tap (A_TAP5) | Airport Bore | <0.01 | Rainwater Tanks | <0.002 | Airport Bore water no longer connected. Currently connected to new rainwater tanks. Plumbing not replaced, but appears to have been effectively flushed. |
| Hospital Taps (PWS_HOSP_TAP1, PWS_HOSP_TAP4, PWS_HOSP_TAP6) | Airport Bore | <0.01 – 0.46 | Rainwater Tanks | <0.002 | Plumbing and filters have been replaced since Airport Bore water supply ceased. |
| Council Works Depot (DEPOT_TANK2) | Airport Bore | 5.54 | Rainwater Tanks | <0.01 | NIRC advised Tank 2 supplies the Council Depot kitchen tap. |
| Council Works Depot Kitchen Tap (DEPOT_TAP1) | Airport Bore | 5.46 | Rainwater Tanks | <0.002 | - |
| Council Kitchen Tap (COUNCIL_TAP1) | NA | NA | Rainwater Tanks | <0.002 | No suspected historical or current PFAS source. |
| Water carting water (WC-03) | Groundwater | <0.01 | Groundwater | 0.004 | - |
| Water carting water (WC-04) | Groundwater | NM | Groundwater | <0.002 | - |

Table Notes: 1. Concentration when first monitored by Senversa 2020-2021. 2: NM: Not measured. 3: NA: Not applicable.

PFAS concentrations were generally reported below the LOR and water at each of these locations is considered suitable for drinking. The exception is at the airport maintenance shed kitchen (A_TAP4), where concentrations remain above the DWG. Although the source of water at this tap could not be confirmed, discussions with NIRC and the reported concentrations indicate the tap is likely still connected to un-treated Airport Bore water. NIRC advised that this tap is not in use and 'do no use' signage is in place.

PFAS concentrations reported in water carting sources analysed were below the DWG. No PFAS were detected in sample WC-04 above the LOR. PFOS was detected in sample WC-03 in 2025 50% below the DWG (0.004 µg/L). Other PFAS for which DWG are available (PFHxS, PFOA, PFBS) were also detected at concentrations well below the DWG (50% of the DWG or lower). It is noted however, that other PFAS compounds (for which DWG are unavailable) were identified at higher concentrations, with the sum concentration of perfluoroalkane sulfonic acids totalling 0.037 µg/L. Monitoring of this location in 2020 reported PFAS below the LOR, noting that the LOR at that time was higher, above the concentrations reported in 2025.

While DWG for these other PFAS are unavailable, further assessment is warranted to provide confidence that the water can be safely used for drinking. This could include repeat monitoring and further risk assessment.

PFAS concentrations were detected in the fire station kitchen tap during the Year 3 ongoing monitoring event. While the concentrations reported were below the health-based drinking water criteria for regulated PFAS (PFOS, PFHxS and PFOA), some other non-regulated PFAS were identified, including 8:2 fluorotelomer sulfonic acid (FTS). These other PFAS compounds are found in newer generation AFFF (e.g. Ansulite) the use of which has also ceased at the fire station (as of 2021, refer to Senversa 2021b). Given these identifications of other PFAS, and the need for a precautionary



approach which considers all PFAS identified, alternative drinking and washing water were supplied by NIRC and further assessment was undertaken to identify the source, completed by others. It was identified that relining of the rainwater tanks is required due to contamination during the fire truck cleaning program. While relining and validation works are completed, NIRC have advised that a desalination water tank and temporary plumbing have been installed at the fire station connected to the kitchen, laundry and bathroom taps. Results of the 2025 monitoring at the kitchen tap were below the LOR and indicate the temporary water supply and plumbing are effective.

5.3.5 Other Water Uses

The laboratory results of public water sources not used for drinking water are compared to historical results in the table below. Concentrations above the recreational water use assessment criterion for PFOS (2 µg/L) are indicated in red for reference, noting that this value is adopted for indicative purposes, and may not be applicable to / protective of all listed uses for these waters.

Table 5-2: Comparison of Historical and 2025 PFAS Concentrations in Public Water Use

| Area / Sample Location | Historical Water Source | PFOS (µg/L) Historical Water Source | Current Water Source | PFOS (µg/L) Nov 2025 | Comments |
|---|-------------------------|-------------------------------------|------------------------------------|----------------------|---|
| Airport Bore Tank (AIRPORT_BORE) | Airport Bore | 33.1 | Airport Bore | 12.5 | Historically used as a holding tank before water was pumped to the fire station and terminal. Currently tank holds untreated Airport Bore water. Water is later filtered by POET filter prior to use. No sample was analysed immediately following filtration. |
| Ben Christian Down Pipe (AIRPORT_TRUCKFILL) | Airport Bore | 14.0 | Treated Airport Bore | NM | Not measured in 2025 or post POET installation. Water used to fill public toilet tanks, at the water treatment facility, to suppress dust at the quarry and wash gravel at the road depot. Latest NIRC testing results post-filtration in May 2025 0.053 µg/L. |
| Airport Terminal Bathrooms (A_TAP1) | Airport Bore | 0.02 | Rainwater Tanks | <0.002 | Airport Bore water no longer connected. Currently connected to new Airport rainwater tanks. Plumbing not replaced, but appears to have been effectively flushed. 'Do not drink' signage in place and used for hand washing only. |
| Public Bathrooms (PWS_CAS_TOILETS, PWS_EB_TOILETS, PWS_HEAD_TOILETS) | Airport Bore, rainwater | 20.8 – 21.2 | Treated Airport Bore, rainwater | NM | Not measured in 2025. In 2024, post POET installation, PFOS <0.01 – 0.77 µg/L reported in Emily Bay, Cockpit Waterfall and Headstone reserve toilet taps. This is below the adopted recreational screening level. Airport bore water supply varies with rainwater availability. |
| Fire Hydrants on Airport (FRE_TAP2) | Airport Bore | 15.0 | Treated Airport Bore and rainwater | 0.03 | Used for firefighting, training and truck flushing. |
| Hospital Laundry Tap (PWS_HOSP_TAP8) | Airport Bore | NM | Hospital bore water | <0.002 | Plumbing and filters have been replaced since airport bore water supply ceased. |
| Council Works Depot (DEPOT_TANK1, TANK3) | Airport Bore | <0.01 | Rainwater Tanks | <0.01 | NIRC advised Tank 1 used for firefighting. New Tank3 installed at rear of depot for rainwater and used for truck washing at the mechanical sheds in the works depot. |



| Area / Sample Location | Historical Water Source | PFOS (µg/L) Historical Water Source | Current Water Source | PFOS (µg/L) Nov 2025 | Comments |
|---------------------------|-------------------------|-------------------------------------|----------------------|----------------------|--|
| Duck Dam Down Pipe | Duck Dam | NM | Duck Dam | NM | Approximated from Duck Dam concentration, PFOS 0.04 µg/L in 2025. This is below the adopted recreational screening level. Water used to wash gravel off-site at the roads depot. |

Of the 37 reticulated water supply locations sampled, 20 were below the LTVs for Sum PFHxS+PFOS. Where this water is used for drinking, further assessment has been made against the 2025 DWG (refer to **Section 5.3.4**). Where water is used for other uses, management measures may be reduced and further assessment is provided below.

There is limited potential human exposure to POET-treated Airport Bore water for the supply indicated by NIRC (public bathroom taps, wastewater treatment plant, firefighting and training, quarry dust suppression and gravel washing) given the current understanding of management measures:

- 'Do not drink' signs are installed at public bathroom taps across the island.
- Both solids and liquids from the wastewater treatment plant are pumped through a pipeline to an outfall pipe at Headstone Cliff directly into the ocean.

The potential for exposure during non-potable water use including hand washing, firefighting and training, dust suppression, gravel washing and vehicle washing is considered relatively low given the frequency and duration of exposure, the limited potential for PFAS adsorption through the skin and the non-volatile nature of PFAS. The concentrations measured since 2021 are below those assessed during the HHERA, which concluded that the risks associated with firefighters contacting this water for fire systems testing, training and firefighting (prior to management) is assessed **low and acceptable**.

On this basis, the human health risks associated with the ongoing use of this water for the known uses are assessed to be low, and these health risks are currently adequately managed. It should be noted however, that introduction of PFAS to previously unimpacted areas should be limited where possible.

Continued testing of the POET-treated water is required to assess the ongoing effectiveness of the filtration system and whether further management is warranted. Furthermore, further assessment of risk would be required prior to utilisation of this treated water for other purposes.

5.4 Nature and Extent of PFAS in Soil and Biota

5.4.1 Duck Dam

Sediment and water hyacinth from Duck Dam were analysed for PFAS to assess management options for material removed from the dam for routine dam maintenance. Assessment of reuse options for PFAS-contaminated material will be based on the principles that reuse must not lead to an unacceptable risk to the environment and/or human health, or an increase in the level of risk at or near the location in which it is used. NIRC indicated that at present, vegetation removed from the Kingston wetland is composted at the Waste Management Centre and either stockpiled at the current drill ground or supplied across the island as a soil ameliorant. To assess whether this management option is suitable for Duck Dam vegetation and sediment, PFAS concentrations have been compared to the HIL-A assessment criteria protective of *residential land use with accessible soil* in the scenario that compost is applied to residential properties.



Regarding the plant material, the measured concentration (0.001 mg/kg PFOS, at the limit of detection) was below soil screening levels that apply in people's back yards (i.e. HIL-A of 0.003 mg/kg; ecological criterion of 0.003 mg/kg). The HIL-A considers backyard exposures including the uptake of PFAS into home-grown produce, so is relevant for compost. On this basis, the risk associated with composting of this vegetation and application to land is considered to be low. It should be noted however, that introduction of PFAS to previously unimpacted areas (e.g. residential backyards) should be limited where possible.

Regarding the sediment material, the measured PFOS concentration (0.01 mg/kg PFOS) was above current soil screening levels that apply in people's back yards (i.e. HIL-A of 0.003 mg/kg PFOS+PFHxS; ecological criterion of 0.003 mg/kg PFOS). Given the relatively low magnitude of the exceedance, small volumes of sediment adhered to plant material is likely to be acceptable. However, adding a significant volume of sediment, for example sediment removed during dam dredging, with low levels of PFAS to areas that have even lower or no levels of PFAS should be considered only in consultation with the relevant environmental regulator in exceptional circumstances where there is no feasible, practicable alternative. Significant volumes of sediment removal from the dam should therefore either be avoided or appropriately managed to avoid migration of PFAS. Appropriate management may include storage at a site with known existing PFAS contamination where it can be demonstrated that application to that land will result in no increased level of risk.

It is noted that this assessment of material re-use is preliminary in nature and does not constitute a site-specific risk assessment. Should sediment be removed from the dam during clearing, further sampling including leachability analysis and a site-specific risk assessment should be completed to inform suitable management measures.

5.4.2 Ball Bay

Soil analysed from the AFFF storage areas of the Ball Bay Fuel Storage Depot reported detectable PFAS in each of the three sampled locations. Concentrations of PFOS exceeded the ecological criterion of 0.003 mg/kg, however were below the human health commercial criterion of 20 mg/kg. As no vegetation is present on the depot itself, this area is unlikely to provide good ecological habitat, or to support a significant proportion of the diet of terrestrial ecological receptors. These factors mean that risks to terrestrial ecological receptors on the depot are likely to be low. To manage the ongoing risk of stored AFFF leakage into the surrounding environment, it is understood that the AFFF will be removed once a suitable alternative is available on-island.

5.4.3 Airport Down Pipe

Soil from below the Ben Christian down pipe was analysed to assess the impact of historical airport bore water supply to this asset and current POET-treated water supply i.e. spills and overtopping of trucks during filling. Concentrations of PFOS (0.0841 mg/kg) exceeded the ecological criterion of 0.003 mg/kg, however were below the commercial criterion of 20 mg/kg. Given the low magnitude of the exceedance, this concentration is considered to pose a negligible risk to human and ecological health. To reduce the risk of ongoing PFAS introduction to soil from impacted groundwater, ongoing filtration of the airport bore water with the POET should be maintained.

5.4.4 Home Grown Produce

For the following home grown produce consumption pathways, risks were assessed in the DSI or HHERA to be negligible, and further assessment was not required:

- One soil sample from ID013 (Mission Creek catchment) exceeded residential use criteria indicating contamination at this property may require management if the land is for residential uses (with home grown produce). However, no PFAS greater than the LOR was reported in produce from this property.
- An exceedance of residential criteria was identified in sediments from the creek bed behind the depot; this is not residential land but limited produce is collected from this area. The HHERA assessed the risks to be low.



- No exceedances of residential screening criteria were reported at all other off-site locations (including residential properties where potential sources were identified), therefore the PFAS identified was assessed to not present an unacceptable risk for residential land use.

Additional sampling was undertaken during the Year 4 monitoring event to further investigate home grown produce at property ID008 where known AFFF application was previously recorded. PFAS was detected in soil in 2021, below the adopted residential assessment criteria protective of home grown produce consumption, as noted above. Re-analysis in a different area of the garden in 2025 indicated a PFOS concentration marginally above the assessment criteria (0.0052 mg/kg compared to 0.003 mg/kg threshold).

Analysis of representative produce in this area, parsley, silver beet, zucchini and tomato, reported PFAS concentrations in each sample except silver beet. The concentration of PFOS in the tomato sample (0.001 mg/kg) exceeded the FSANZ trigger point for further investigation (0.001 mg/kg, the limit of detection). Additionally, the laboratory-reported sum of perfluoroalkane carboxylic acids in the parsley sample was 0.036 mg/kg. No trigger point has been set for these summed compounds; however it is noted that the concentration of PFOA (the single perfluoroalkane carboxylic acid compound for which a trigger point is available) was below its respective trigger point.

No chickens were kept for egg or meat consumption at this property.

It is noted that the FSANZ (2017) trigger points apply a set of conservative assumptions, including that the consumers are aged 2 to 6 years, have a high consumption rate of fruits and vegetables, and that 100% of an individual's fruit or vegetable intake comes from the contaminated source. Given these assumptions and the PFAS detections reported, further assessment is warranted which considers the nature of the produce in which exceedances were identified and likely consumption rates.

PFAS accumulates in humans over long periods of exposure to PFAS-affected sources, such as food or water. Given this, while we work to undertake further assessment and (if necessary) develop management measures, continuing consumption of this home-produce would not have a significant impact on long-term exposure PFAS exposure and risk. Notwithstanding this, we note that reducing or limiting the consumption of home produce from this property is an effective way for the residents to reduce their current exposure to PFAS. Recommendations are included in **Section 6.2**.



6.0 Conceptual Site Model

6.1 Conceptual Site Model Summary

A conceptual summary of the linkages between the main PFAS source areas, pathways and identified receptors is provided below, with further information provided in the DSI report (Senversa, 2021b). No changes to the source areas, pathways or potential receptors were identified as a result of the Year 4 monitoring event.

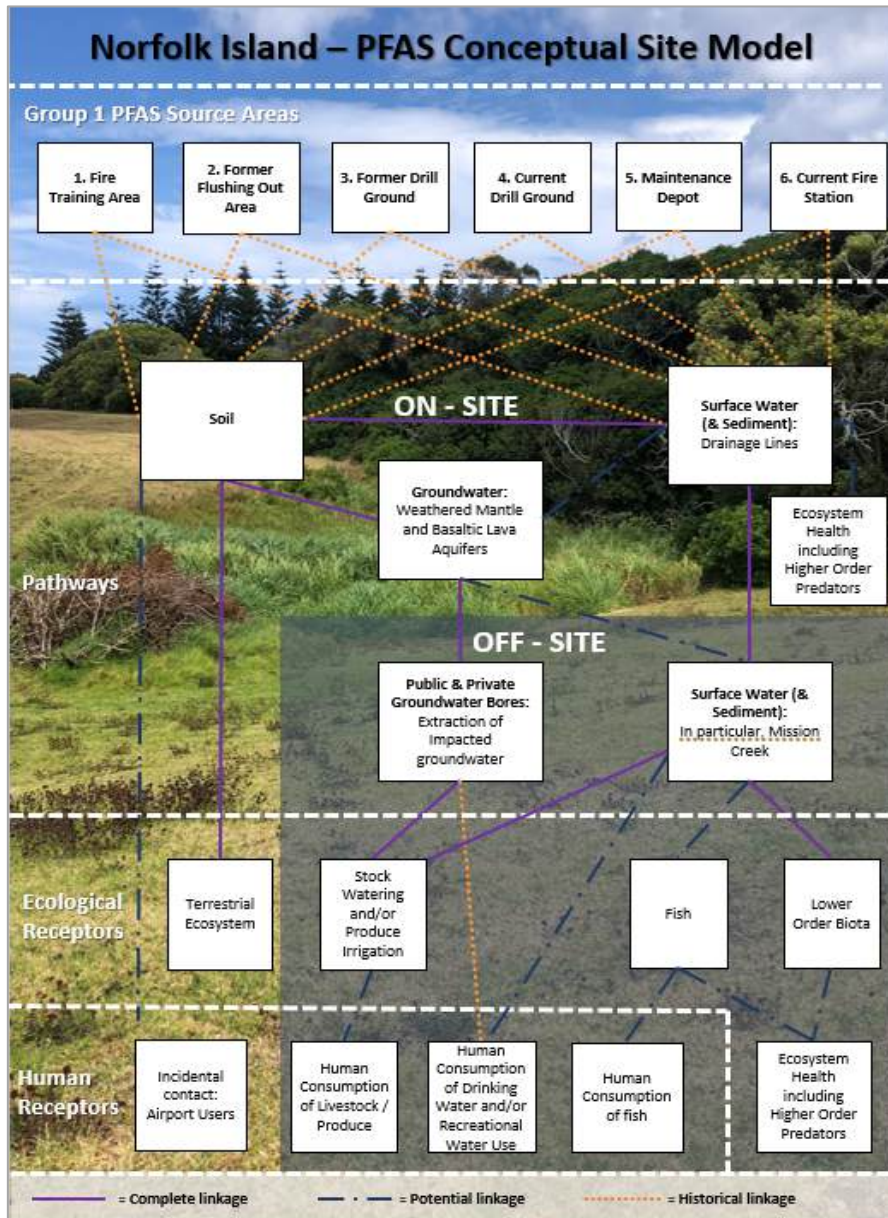


Figure 6-1: PFAS Conceptual Site Model Flow Chart



6.2 Assessment of Management Actions

Following the identification of PFAS in groundwater in late 2019, DITRD/CSA have worked with NIRC to undertake a number of management actions aimed at reducing the potential for exposure to the identified PFAS within the on-island environment both on-Airport and off-Airport.

Use of Legacy AFFF for training ceased in 2015 and emergency use ceased in 2022. Legacy AFFF foams are no longer used, and a fluorine free foam has since been introduced at the airport. The fire trucks have been cleaned and tested for PFAS and are now reported free of Legacy AFFF. AFFF continues to be stored at Ball Bay Fuel Depot in case of emergency and replacement is planned when an alternative product is available on-island.

Source management options are assessed in the PMP (Senversa, 2021e). Additional management actions already in place and recommended as a result of the November 2025 monitoring results are summarised in **Table 6-1** below.

A future reduction in monitoring may be warranted at locations where the LTVs were not met. This will be assessed in the updated PMP.

Table 6-1: PFAS Management Actions

| Risk Identification | Do Existing Management Measures Mitigate Risks? | Recommended Additional or Ongoing Controls |
|---|---|---|
| <p>1. Mission Creek: Home consumption or public consumption of cattle, chicken eggs or other animal products where the animal drinks water sourced from Mission Creek.</p> | <p>Yes, though further measures warranted for long-term effective management</p> <p>Advice provided in results letters to continue not using water for chicken watering.</p> <p>Advice provided in results letters that further assessment / management warranted to ensure ongoing effective mitigation of risks associated with the consumption of livestock products where cattle drink water sourced from Mission Creek. The letters also indicated that PFAS accumulates in humans over long periods of exposure to PFAS-affected sources, such as food or water and (given this), while further assessment and management approaches are completed, continuing with current practices would not have a significant impact on long-term exposure.</p> | <p>The findings from this monitoring round confirm the previous recommendations for further management of the use of water from Mission Creek for watering cattle:</p> <ul style="list-style-type: none"> Further assessment and/or management of cattle access to PFAS impacted water sources required. PFAS concentrations during this monitoring event remained consistent with those measured in 2022, and above those measured in earlier sampling (2020 and 2021), following which it was assessed that further assessment/management was warranted. Alternatively, measures to manage human exposure (e.g. livestock product consumption advice) could be considered. <p>For chickens at ID013, PFAS was <LOR in the currently used water supply (ID013_BORE).</p> <p>Previous advice to livestock farmers remains unchanged and use of water from Mission Creek (e.g. ID013_SW01) for watering chickens should not be recommended.</p> |
| <p>2. Mission Creek Catchment: use of surface water or groundwater for any extractive use (other than livestock watering)</p> | <p>Yes</p> <p>There are no current unacceptable exposures identified; and advice has been provided not to use water for drinking / domestic use.</p> <p>ID013_SW01 water no longer in use.</p> | <p>Continued advice not to use surface water for produce irrigation required.</p> <p>Continued advice to not use surface water or groundwater for drinking / domestic use required.</p> |



| Risk Identification | Do Existing Management Measures Mitigate Risks? | Recommended Additional or Ongoing Controls |
|--|---|--|
| <p>3. Groundwater near airport: use of groundwater from or nearby the Airport for any extractive use.</p> | <p>Yes</p> <p>There are no current extractive uses of water identified, with the exception of uses assessed to be associated with low and acceptable risks.</p> <p>The POET filtration system was in use at the Airport Bore at the time of the Monitoring Event and treated water utilised by NIRC for various public uses. Ben Christian Down Pipe was locked at the time of the Monitoring Event and 'do not drink' signs were observed at public toilet taps.</p> | <p>NIRC advised that routine testing of the POET-treated water is undertaken. While there is low exposure potential for the known uses of this water, continued testing of the treated water will be required to assess the ongoing effectiveness of the POET filtration system and whether further management is warranted. In order to demonstrate effective management, the off-site transfer of PFAS via use of this water should be minimised to the extent practicable, and this requires that the filtration system continues to be effective in removing PFAS (i.e. functioning optimally and removing PFAS in line with design specifications).</p> <p>PFAS detected in water carting supply was below the DWG, however confirmatory sampling is recommended.</p> |
| <p>4. Drinking or washing water at public facilities formerly supplied by the Airport Bore and now supplied by alternative water supply including: the fire station, other on-Airport buildings, hospital, and council works depot.</p> | <p>Yes</p> <p>Sampling undertaken at public facilities indicate that replacement of the PFAS impacted water source, Airport Bore, with alternative water supplies (rainwater and desalinated water) has been successful in reducing PFAS concentrations to levels below the guidance values. This includes replacement of the hospital pipework and filters (NIRC, 2021).</p> <p>This means it is safe to use the water, including for sensitive uses such as drinking and eating.</p> <p>Water in the airport maintenance shed kitchen remains unsuitable for drinking and 'do not drink' signs should be maintained.</p> | <p>Continued controls are required such that PFAS impacted water (e.g., Airport Bore) is not used to supply drinking water while above HBGV. This includes the lock on the Ben Christian Down Pipe and signage at public bathroom taps and the airport maintenance shed kitchen.</p> |
| <p>5. Watermill Creek Catchment: use of surface water or groundwater for drinking water or domestic use.</p> | <p>Yes</p> <p>No current use of water for drinking water or domestic use identified, and advice has been provided not to use water for drinking / domestic use.</p> | <p>Continued ongoing monitoring of PFAS concentrations in Watermill Creek, with a view to revising advice if concentrations decrease below the guidance value in the future.</p> |
| <p>6. Exposures to freshwater aquatic ecosystems.</p> | <p>Pending source management</p> | <p>Continued ongoing monitoring of PFAS concentrations, with a view to ecological risk revision if concentrations decrease below guideline values in the future.</p> |
| <p>8. Consumption of produce grown in PFAS source areas.</p> | <p>Further assessment warranted</p> <p>Advice will be provided to relevant landholders, and additional assessment undertaken to determine the requirement for management measures.</p> | <p>-</p> |



7.0 Conclusions

Senversa undertook the PFAS OMP Year 4 monitoring event from 17 to 21 November 2025 in general accordance with the SAQP. The following findings were made addressing the objectives outlined in **Section 1.2**:

Objective 1: Trends in PFAS concentrations in the environment

- Reported concentrations of PFAS in the Airport Bore and the upper Mission Creek catchment surface water generally decreased between 2020 and November 2025, representing a statistically significant downwards trend for PFOA, PFOS and PFHxS+PFOS at the Airport Bore and WWII_DAM, and for PFHxS+PFOS at MC_OMP01.
- PFAS concentrations have been variable over time in both the remaining Mission Creek surface water locations and Watermill Creek. Approximately half of the sampled locations reported PFAS concentrations marginally higher in 2025 than 2024, however most remained within the historical range. Highest-to-date concentrations were recorded at ID014 in the Mission Creek Catchment and Watermill Creek Duck Dam for selected PFAS, however the increases above previous maximum concentrations were marginal.
- PFAS concentrations in surface water generally decreased with increased distance from source areas.

Data Gap Assessment

- PFAS were detected in home grown produce grown in PFAS-impacted soil and waterway biota and sediments, requiring further assessment.
- Concentrations in water carting water were below the adopted assessment criteria. However, detections of PFAS for which guidelines are unavailable require further assessment.
- PFAS were detected in soil at source areas Ball Bay fuel depot and below the Ben Christian Down Pipe, below the commercial land use assessment criteria.

Objective 2: The Effectiveness of the Selected Management Options in Managing Current Risks

Reported PFAS concentrations were below the UTVs at all sample locations. A number of point of use sample locations reported PFAS concentrations below the LTVs.

The selected management options are considered appropriate for the purpose of managing current risks. In addition, the following changes in management options and potential further assessments should be considered:

- Stock watering: Further assessment and/or management of cattle access to PFAS impacted water sources requires further consideration. PFAS concentrations during this monitoring event remained consistent with those measured in 2022, and above those measured in earlier sampling (2020 and 2021), following which it was assessed that further assessment/management was warranted. Alternatively, measures to manage human exposure (e.g. livestock product consumption advice) could be considered.
- Airport Bore water use: The POET filtration system has been successfully installed on the Airport Bore and NIRC advised that routine testing of the treated water is undertaken. While there is low exposure potential for the known uses of this water, continued testing of the treated water will be required to assess the ongoing effectiveness of the POET filtration system and whether further management is warranted. The efficacy of the POET filter at reducing PFAS concentrations in the Airport Bore water and suitable concentration threshold limits will require assessment prior to utilisation of this treated water for other purposes.
- Bore water carting: Further assessment of the quality of water carting water sources used for potable water supply is recommended to confirm detected PFAS concentrations and facilitate a more detailed assessment including consideration of PFAS compounds for which no DWG are available. It is noted that no unacceptable risk has been identified to-date.



- Home produce growing: Where PFAS was detected in home grown produce, further assessment is warranted which considers the nature of the produce in which exceedances were identified and likely consumption rates. This will allow the requirement for management to be determined.
- Biota and sediment re-use: Based on a preliminary assessment, the re-use of biota from Duck Dam, e.g. for compost, is considered to pose a low and acceptable risk. The re-use of sediment should be avoided or appropriately managed to avoid increasing the level of risk on less-impacted sites.

A future reduction in monitoring may be warranted at locations where the LTVs were not met; this will be assessed in the revised PMP.

Objective 3: Has the Change in Conditions Resulted in a Change in the Risk Profile

The change in PFAS concentrations has not resulted in a potentially unacceptable change to the risk profile and therefore no change in management controls is required.



8.0 Principles and Limitations of Investigation

8.1 General Principles and Limitations of Investigation

The following principles are an integral part of site contamination assessment practices and are intended to be referred to in resolving any ambiguity or exercising such discretion as is accorded the user or site assessor.

Table 8-1: Principles and Limitations of Investigation

| Area | Field Observations and Analytical Results |
|---|--|
| Elimination of Uncertainty | Some uncertainty is inherent in all site investigations. Furthermore, any sample, either surface or subsurface, taken for chemical testing may or may not be representative of a larger population or area. Professional judgment and interpretation are inherent in the process, and even when exercised in accordance with objective scientific principles, uncertainty is inevitable. Additional assessment beyond that which was reasonably undertaken may reduce the uncertainty. |
| Failure to Detect | Even when site investigation work is executed competently and in accordance with the appropriate Australian guidance, such as the National Environmental Protection (Assessment of Site Contamination) Amendment Measure ('the NEPM'), it must be recognised that certain conditions present especially difficult target analyte detection problems. Such conditions may include, but are not limited to, complex geological settings, unusual or generally poorly understood behaviour and fate characteristics of certain substances, complex, discontinuous, random, or heterogeneous distributions of existing target analytes, physical impediments to investigation imposed by the location of services, structures and other man-made objects, and the inherent limitations of assessment technologies. |
| Limitations of Information | The effectiveness of any site investigation may be compromised by limitations or defects in the information used to define the objectives and scope of the investigation, including inability to obtain information concerning historic site uses or prior site assessment activities despite the efforts of the user and assessor to obtain such information. Information received during preparation of this report from third parties or anecdotal sources, such as the sources of PFAS identified, was not able to be independently verified by Defence records. |
| Chemical Analysis Error | Chemical testing methods have inherent uncertainties and limitations. Senversa routinely seeks to require the laboratory to report any potential or actual problems experienced, or non-routine events which may have occurred during the testing, so that such problems can be considered in evaluating the data. |
| Level of Assessment | The investigation herein should not be considered to be an exhaustive assessment of environmental conditions on a property. There is a point at which the effort of information obtained and the time required to obtain it outweigh the benefit of the information gained and, in the context of private transactions and contractual responsibilities, may become a material detriment to the orderly conduct of business. If the presence of target analytes is confirmed on a property, the extent of further assessment is a function of the degree of confidence required and the degree of uncertainty acceptable in relation to the objectives of the assessment. |
| Comparison with Subsequent Inquiry | The justification and adequacy of the investigation findings in light of the findings of a subsequent inquiry should be evaluated based on the reasonableness of judgments made at the time and under the circumstances in which they were made. |
| Data Useability | Investigation data generally only represent the site conditions at the time the data were generated. Therefore, the usability of data collected as part of this investigation may have a finite lifetime depending on the application and use being made of the data. In all respects, a future reader of this report should evaluate whether previously generated data are appropriate for any subsequent use beyond the original purpose for which they were collected, or are otherwise subject to lifetime limits imposed by other laws, regulations or regulatory policies. |
| Nature of Advice | The investigation works herein are intended to develop and present sound, scientifically valid data concerning actual site conditions. Senversa does not seek or purport to provide legal or business advice. |



9.0 References

- Abell, R S & Falkland A C, 1991. *The hydrogeology of Norfolk Island South Pacific Ocean*. Department of Primary Industries and Energy Bureau of Mineral Resources, Geology and Geophysics, Bulletin 234.
- Abell, R S, 1993. *Aquifer vulnerability on small volcanic islands in the southwest Pacific region – an example from Norfolk Island*. AGSO Journal of Australian Geology & Geophysics, 14 (2/3), pp 123-133.
- Australian Government – Geoscience Australia, 2020 – Norfolk Island. Accessed 21 February 2020.
- BoM, 2025. *Climate statistics for Australian locations: Summary Statistics Norfolk Island Aero*.
- CSIRO (2020) *Norfolk Island Water Resource Assessment Hydrology Report. A summary report from the CSIRO Norfolk Island Water Resource Assessment*, CSIRO, Australia.
- FSANZ, 2017. *Perfluorinated chemicals in food. Food Standards Australia New Zealand*. April 2017.
- HEPA, 2020. *PFAS National Environmental Management Plan (PFAS NEMP) 2.0*.
- HEPA, 2025. *PFAS National Environmental Management Plan (PFAS NEMP) 3.0*.
- NHMRC, 2019. *Guidance on Per and Polyfluoroalkyl substances (PFAS) in Recreational Water*. National Health and Medical Research Council, Canberra. Publication EH45, Canberra.
- NHMRC, 2025. *National Water Quality Management Strategy, Australian Drinking Water Quality Guidelines 6 2011*, Version 4.0 updated June 2025.
- NIRC, 2018. *2018 - 2023 Norfolk Island Environment Strategy* (Norfolk Island Environment Strategy). November 2018.
- Senversa, 2020a. *Preliminary Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*.
- Senversa, 2021a. *Preliminary Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*, revision 2, dated 3 February 2021.
- Senversa, 2021b. *Detailed Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*, revision 5, dated 12 November 2021.
- Senversa, 2021c. *Human Health and Ecological Risk Assessment (PFAS), Norfolk Island Airport*, revision 3, dated 12 November 2021.
- Senversa 2021d. *Ongoing Monitoring Plan, Norfolk Island Airport*, revision 0, dated 24 November 2021.
- Senversa 2021e. *PFAS Management Plan, Norfolk Island Airport*, revision 1, dated 10 December 2021.
- Senversa, 2022a. *PFAS Sampling and Analysis Quality Plan – Year 1 Ongoing Monitoring*, revision 1, dated 3 May 2022.
- Senversa, 2022b. *PFAS Ongoing Monitoring Plan – Year 1 Monitoring Report, Norfolk Island Airport*, revision 1, dated 14 October 2022.
- Senversa, 2023. *PFAS Ongoing Monitoring Plan – Year 2 Monitoring Report, Norfolk Island Airport*, revision 1, dated 11 October 2023.
- Senversa, 2024. *PFAS Ongoing Monitoring Plan – Year 3 Monitoring Report, Norfolk Island Airport*, revision 1, dated 16 October 2024.



Figures

Figure 1: Monitoring Plan Locations

Figure 2A: Site Layout and On-Site PFAS Source Areas

Figure 2B: Off-Site PFAS Source Areas

Figure 3: Surface Water and Groundwater Concentrations PFOS – Ecological Guidelines

Figure 4: Surface Water and Groundwater Concentrations PFOS – Human Health Guidelines

Figure 5: Soil, Sediment and Biota Concentrations PFOS – Ecological and Human Health Guidelines

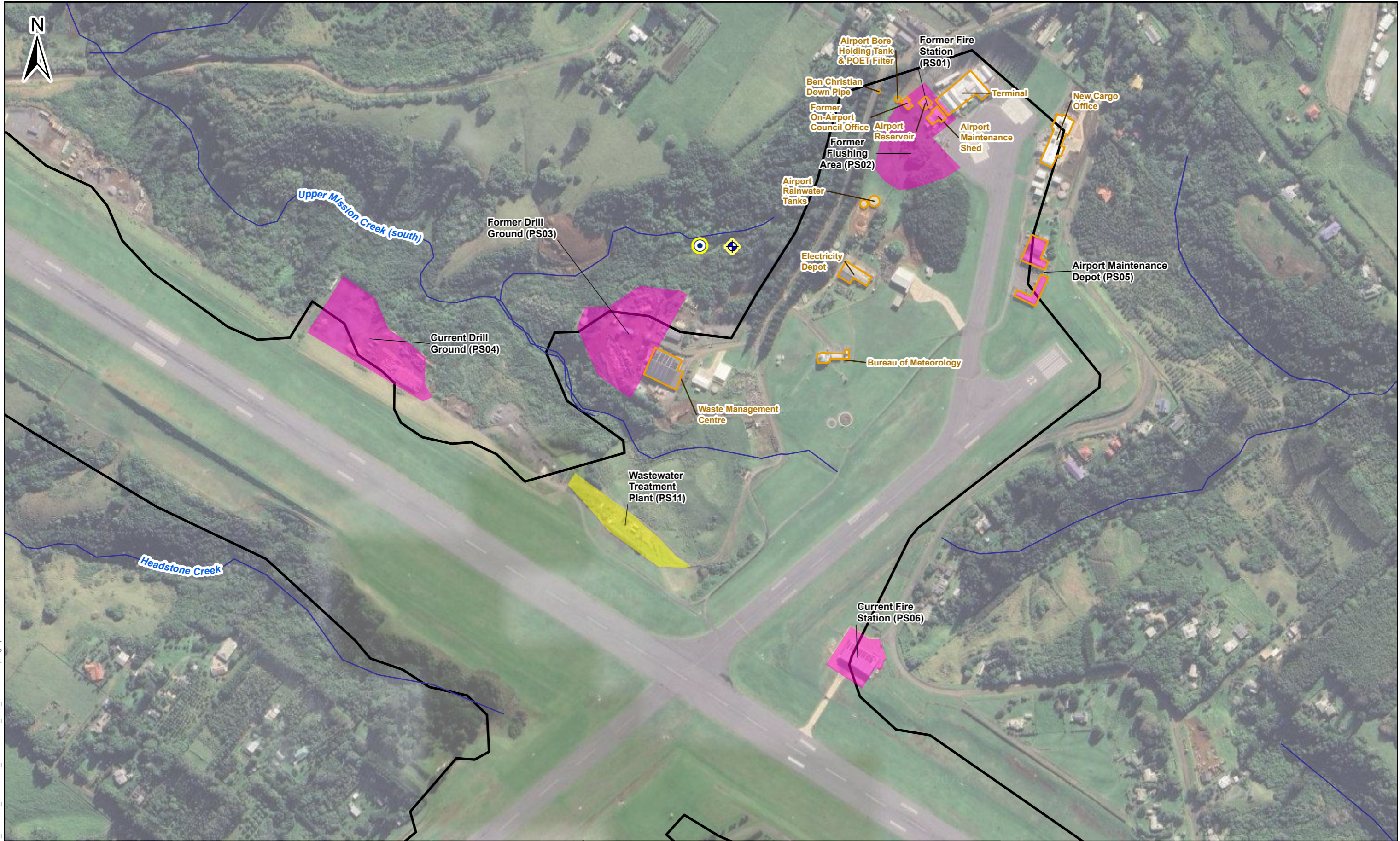


| Legend | |
|--------|---|
| | Surface Soil |
| | Biota |
| | Groundwater |
| | Tap Water |
| | Pump |
| | Stock Water (Cow) |
| | Stock Water (Vegetables and Chicken) |
| | Surface Water and Sediment |
| | Tap Water |
| | Biota |
| | Toilet Water |
| | Waterways |
| | National Park |
| | Surface Soil |
| | Mountain Peak |
| | Kingston and Arthurs Vale Historic Area (KAVHA) |
| | Approximate Extent of Surface Water Catchments |
| | Headstone Creek |
| | Mission Creek |
| | Mt Pitt / Broken Bridge Creek |
| | Rocky Point Creek |
| | Stockyard Creek |
| | Town Creek / Watemill Creek |
| | Approximate Airport Boundary |

| | | | |
|---|---|-----------|---------------|
| Created: | A. Murray | Date: | 24/02/2026 |
| Reviewed: | M. Agnew | Revision: | 0 |
| Approved: | M. Agnew | Scale: | 1:30,000 (A3) |
| File: | C17776_040_F001_Ongoing_Monitoring_Plan_Locations | | |
| | | | |
| Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere | | | |

| | |
|-------------------|---|
| Figure No: | 1 |
| Title: | Ongoing Monitoring Plan Locations |
| Project: | OMP Year 4 Monitoring Report (2025) |
| Location: | Norfolk International Airport |
| Client: | Department of Infrastructure, Transport, Regional Development and Communication |

Path: S:\01_Jobs\1_NSW_Jobs\17776_DITCRD_NORFOLK_DSIC\17776_040_Year 4 Sampling.aprx



Legend

- Site Feature
- PFAS Source Areas (Potential and Confirmed)
- Group 1 Source Area
- Group 2 Source Area
- Airport Bore
- WWII Dam
- Approximate Airport Boundary
- Waterways



| | | | |
|---|------------------------------|-----------|--------------|
| Created: | A. Murray | Date: | 24/02/2026 |
| Reviewed: | M. Agnew | Revision: | 0 |
| Approved: | M. Agnew | Scale: | 1:5,000 (A3) |
| File: | C17776_040_F002a_Site Layout | | |
| | | | |
| Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere | | | |

Figure No: 2a
Title: Site Layout and PFAS Source Areas Onsite

Project: OMP Year 4 Monitoring Report (2025)
 Location: Norfolk International Airport
 Client: Department of Infrastructure, Transport, Regional Development and Communication



St Barnabas Chapel (PS08)

Current Drill Ground (PS04)

Wastewater Treatment Plant (PS11)

Current Fire Station (PS06)

Former Drill Ground (PS03)

Former Fire Station (PS01)

Former Flushing Area (PS02)

Maintenance Depot (PS05)

Public toilets filled by airport bore (PS16)

Headstone Burning Area (PS15)

Council works depot (PS09)

Perfumery (PS14)

Private residence on Webb Adams Road (PS12)





Paradise Hotel (PS13)





Ball Bay refuelling area (PS10)





Common Oval (PS07)

Public toilets filled by airport bore (PS16)

Legend

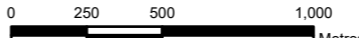
-  Airport Bore
-  WWII Dam
-  Mountain Peak
-  Elevation Contours (approximate)

-  Creek
-  Approximate Airport Boundary
- PFAS Source Areas (Potential and Confirmed)**
-  Group 1 Source Area
-  Group 2 Source Area

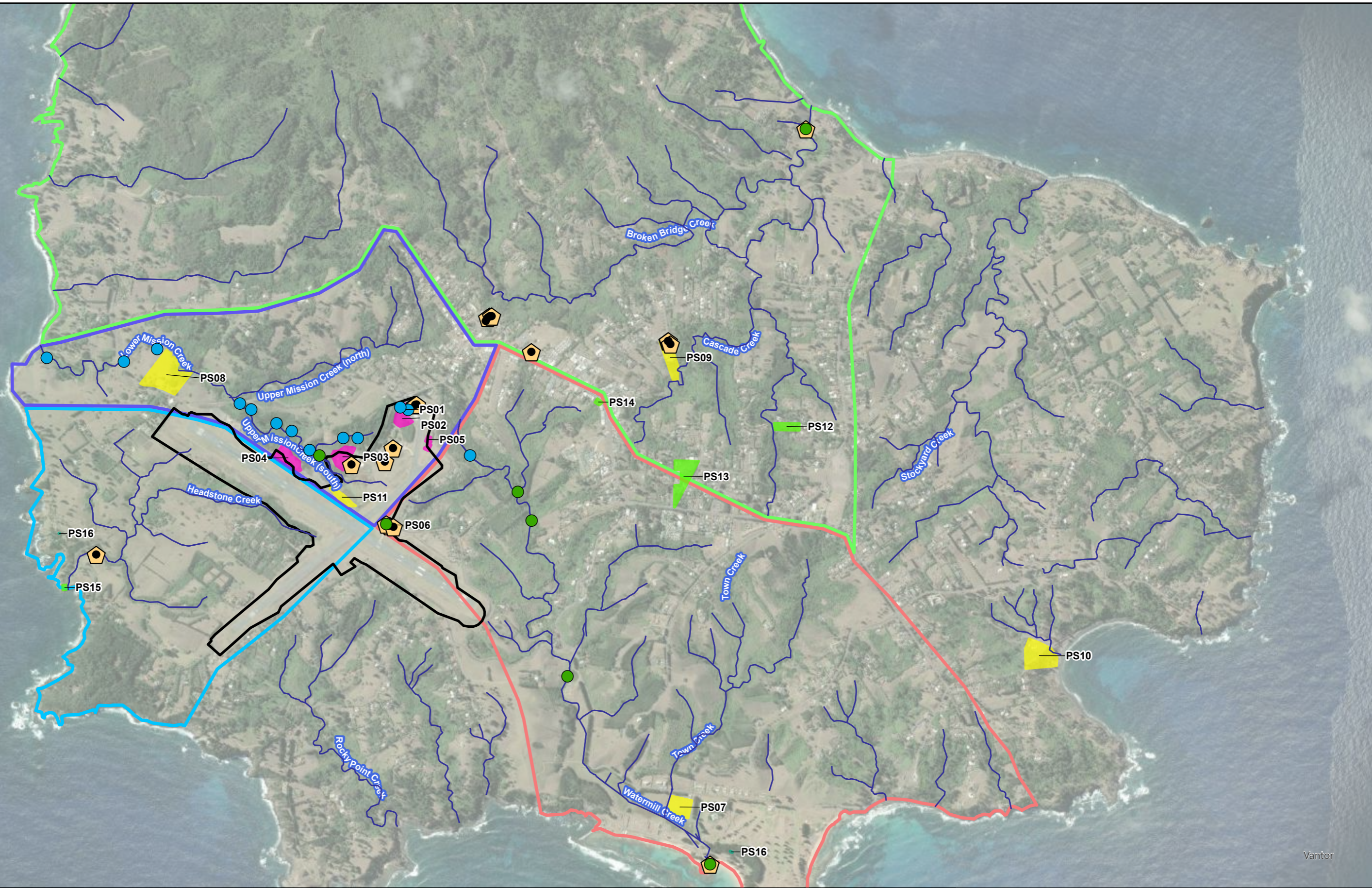
-  Group 3 Source Area
-  Group 4 Source Area
- Areas of Significance**
-  National Park
-  Vale Historic Area (KAVHA)



Aerial Imagery: Esri World Imagery

| | | | |
|---|--|-----------|---------------|
| Created: | J. Currie | Date: | 24/02/2026 |
| Reviewed: | M. Agnew | Revision: | 2 |
| Approved: | M. Agnew | Scale: | 1:25,000 (A3) |
| File: | C17776_040_F002b_Site Layout, PFAS_Offsite | | |
|  | | | |
| Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere | | | |

| | |
|-------------------|---|
| Figure No: | 2b |
| Title: | Site Layout and PFAS Source Areas Offsite |
| Project: | OMP Year 4 Monitoring Report (2025) |
| Location: | Norfolk International Airport |
| Client: | Department of Infrastructure, Transport, Regional Development and Communication |



Vantor

Path: S:\01_Jobs\1_NSW_Jobs\1_NSW_Jobs\17776_040_Year 4 Sampling.aprx



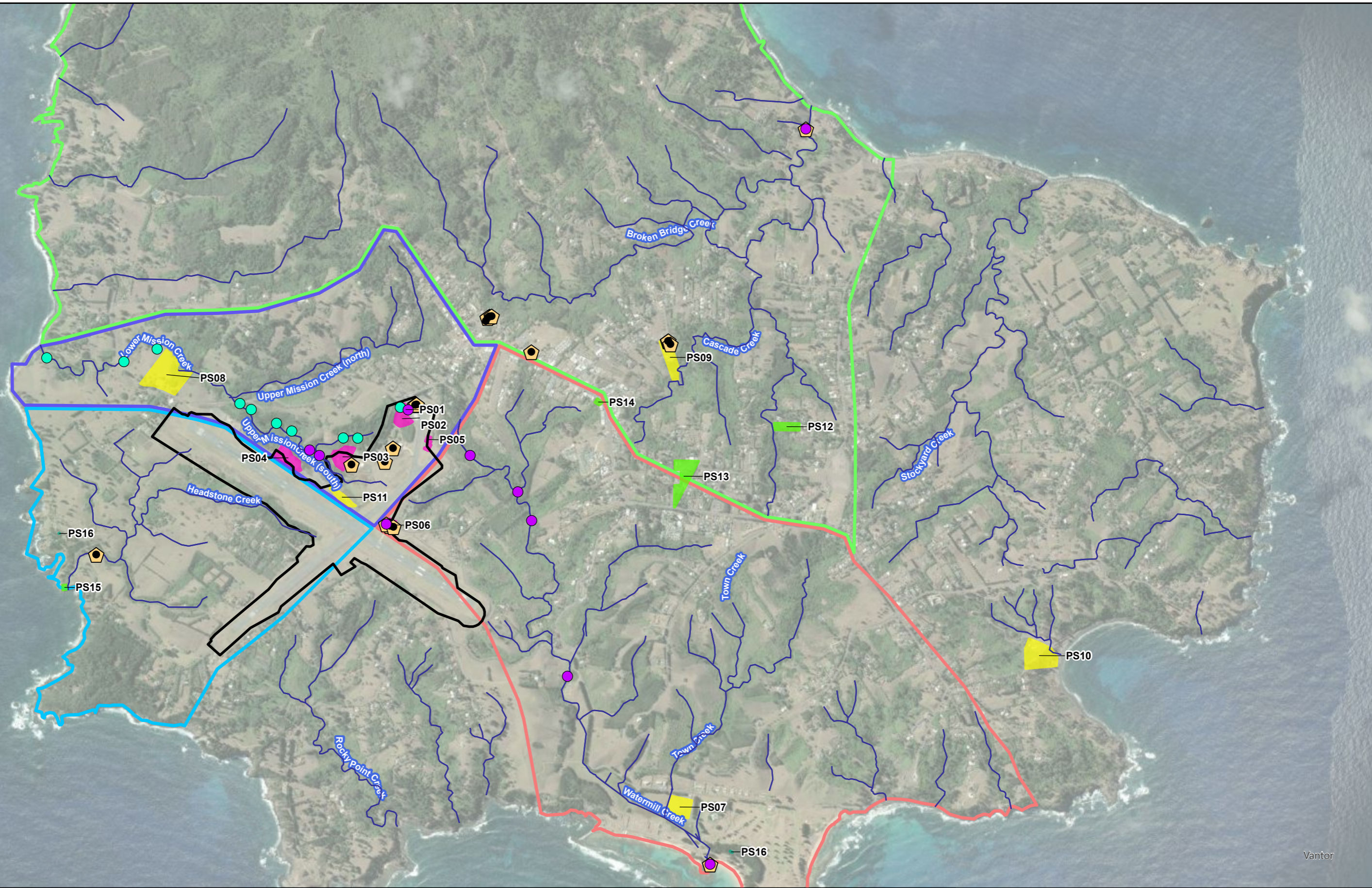
- Legend**
- Approximate Airport Boundary
 - Waterways
 - Approximate Extent of Surface Water Catchments
 - Headstone Creek
 - Mission Creek
 - Mt Pitt / Broken Bridge Creek
 - Town Creek / Watermill Creek
 - PFAS Source Areas (Potential and Confirmed)
 - Group 1 Source Area
 - Group 2 Source Area

- Group 3 Source Area
- Group 4 Source Area
- PFOS Exceedances - Ecological
 - Above Aquatic ecosystems DGV - 90% protection (0.13 µg/L).
 - Above Aquatic ecosystems DGV - 95% and 99% protection (0.00023 µg/L).
 - Below LOR
 - Below LTV

Aerial Imagery : Esri World Imagery

| | | | |
|---|---------------------------|-----------|---------------|
| Created: | A. Murray | Date: | 24/02/2026 |
| Reviewed: | M. Agnew | Revision: | 0 |
| Approved: | M. Agnew | Scale: | 1:25,000 (A3) |
| File: | C17776_040_F003_Water Eco | | |
| | | | |
| Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere | | | |

| | |
|-------------------|--|
| Figure No: | 3 |
| Title: | Surface Water and Groundwater Concentrations PFOS - Ecological Guidelines |
| Project: | OMP Year 4 Monitoring Report (2025) |
| Location: | Norfolk International Airport |
| Client: | Department of Infrastructure, Transport, Regional Development and Communication |



Vantor

Legend

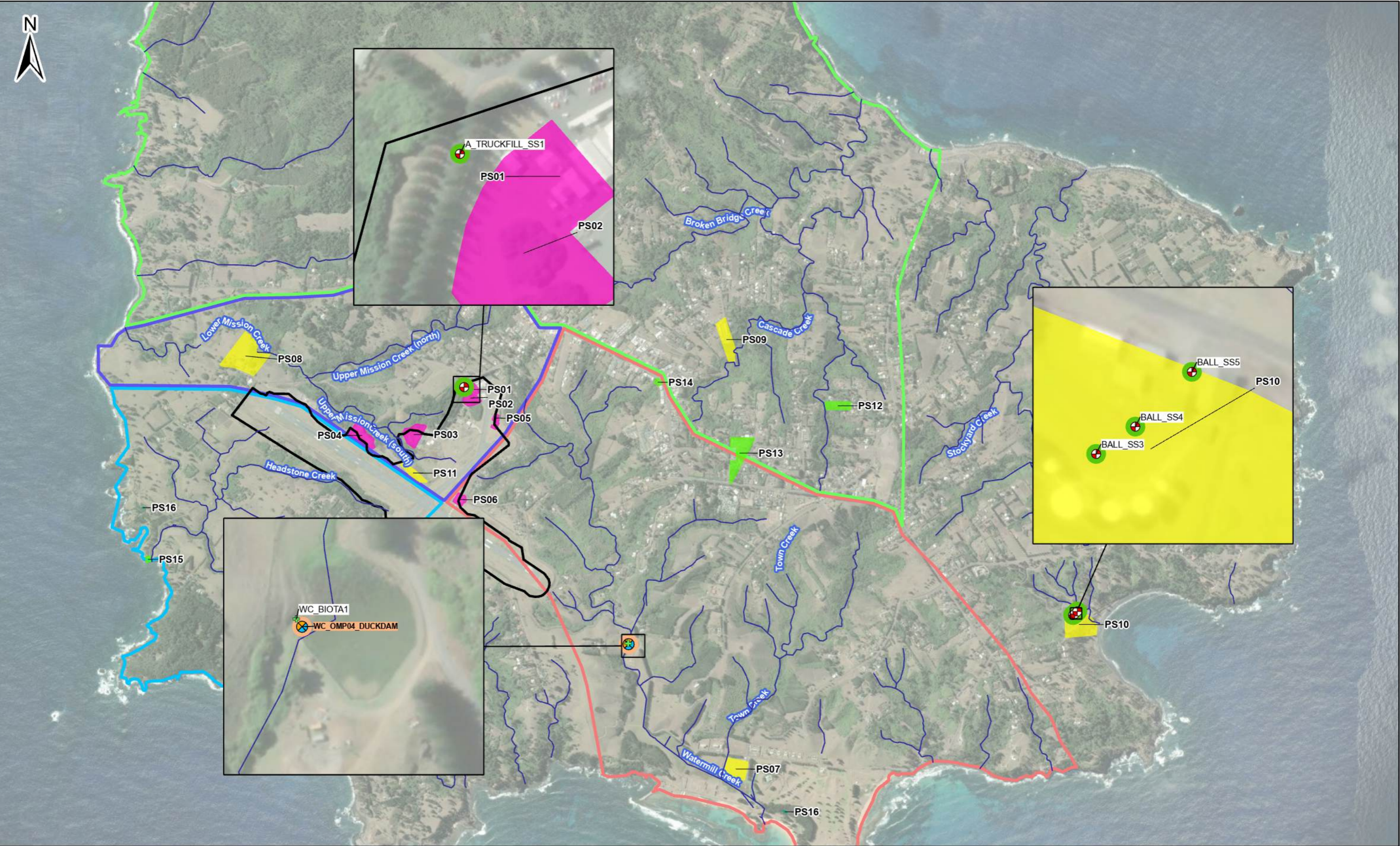
- Approximate Airport Boundary
- Waterways
- Approximate Extent of Surface Water Catchments
 - Headstone Creek
 - Mission Creek
 - Mt Pitt / Broken Bridge Creek
 - Town Creek / Watermill Creek
- PFAS Source Areas (Potential and Confirmed)
 - Group 1 Source Area
 - Group 2 Source Area
 - Group 3 Source Area
 - Group 4 Source Area
- PFOS Exceedances - Human Health
 - Above Recreational Water - Health (2µg/L)
 - Above Drinking Water - Health (0.008µg/L)
 - Below LOR
 - Below LTV

Aerial Imagery : Esri World Imagery

| | | | |
|---|--------------------------|-----------|---------------|
| Created: | A. Murray | Date: | 24/02/2026 |
| Reviewed: | M. Agnew | Revision: | 0 |
| Approved: | M. Agnew | Scale: | 1:25,000 (A3) |
| File: | C17776_040_F004_Water HH | | |
| | | | |
| Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere | | | |

| | |
|-------------------|--|
| Figure No: | 4 |
| Title: | Surface Water and Groundwater Concentrations PFOS - Human Health Guidelines |
| Project: | OMP Year 4 Monitoring Report (2025) |
| Location: | Norfolk International Airport |
| Client: | Department of Infrastructure, Transport, Regional Development and Communication |

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Legend

- + Biota
- Surface Soil
- ⊗ Surface Water And Sediment
- PFOS exceedances for applicable land use setting
- Above Soil Human Health Commercial (20mg/kg)
- Above Soil Maintenance of Ecosystems & Human Health Residential (0.003 mg/kg)
- Above Soil Maintenance of Ecosystems Commercial (0.003 mg/kg)
- Waterways
- Approximate Airport Boundary
- Approximate Extent of Surface Water Catchments
- Headstone Creek
- Mission Creek
- Mt Pitt / Broken Bridge Creek
- Town Creek / Watermill Creek
- PFAS Source Areas (Potential and Confirmed)
- Group 1 Source Area
- Group 2 Source Area
- Group 3 Source Area
- Group 4 Source Area

| | | | |
|-----------|---|-----------|---------------|
| Created: | A. Murray | Date: | 27/02/2026 |
| Reviewed: | M. Agnew | Revision: | 0 |
| Approved: | M. Agnew | Scale: | 1:25,000 (A3) |
| File: | C17776_040_F005_Soil Sediment Biota Exceedances | | |

0 250 500 1,000 Metres

Coordinate System: WGS 1984 Web Mercator Auxiliary Sphere

Figure No: 5

Title: Soil, Sediment and Biota Concentrations PFOS - Ecological and Human Health Guidelines

Project: OMP Year 4 Monitoring Report (2025)

Location: Norfolk International Airport

Client: Department of Infrastructure, Transport, Regional Development and Communication



Tables

Table 1: Upper Trigger Values: Mission Creek Irrigation Water

Table 2: Upper Trigger Values: Other Creeks Water

Table 3: Upper Trigger Values: Cascade and Headstone Creeks Water

Table 4: Lower Trigger Values: Public Water Use

Table 5: Lower Trigger Values: Cattle Stock Watering

Table 6: Lower Trigger Values Chicken Watering

Table 7: Historical Water Analytical Results

Table 8: Soil and Sediment Analytical Results

Table 9: Biota Analytical Results

Table 1: Upper Trigger Values: Mission Creek Irrigation Water
DITRDCA Norfolk PFAS



| | | |
|-----------------------|------------|------------|
| Location Code | ID013_BORE | ID016_BORE |
| Field ID | ID013_BORE | ID016_BORE |
| Date | 17/11/2025 | 17/11/2025 |
| Sample Type | Normal | Normal |
| Lab Report No. | ES2537583 | ES2537583 |

| | Unit | EQL | Upper Trigger Value - Mission Creek | | |
|---|------|-------|--|-------|-------|
| (n:2) Fluorotelomer Sulfonic Acids | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | <0.01 | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | 4.2 ^{#1} | <0.01 | <0.01 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | 2.5 ^{#1} | <0.01 | 0.09 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | <0.01 | 0.09 |
| Perfluoroalkyl Sulfonamides | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEIFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.02 | <0.02 |
| PFAS | | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - | - |
| Sum of PFAS | µg/L | 0.002 | | <0.01 | 0.09 |

Comments

#1 Mission Creek water used for irrigation

Table 2: Upper Trigger Values: Other Creeks Water
DITRDCA Norfolk PFAS



| | | | Location Code | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCKDAM | WC_OMP05 |
|---|------|-------|------------------------------------|------------|------------|------------|------------------|------------|
| | | | Field ID | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCKDAM | WC_OMP05 |
| | | | Date | 17/11/2025 | 17/11/2025 | 18/11/2025 | 18/11/2025 | 18/11/2025 |
| | | | Sample Type | Normal | Normal | Normal | Normal | Normal |
| | | | Lab Report No. | ES2537583 | ES2537583 | ES2537583 | ES2537583 | ES2537583 |
| | Unit | EQL | Upper Trigger Value - Other Creeks | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | 0.12 | 0.06 | <0.02 | <0.02 | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | 0.04 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | - | - | - | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | 0.02 | <0.01 | <0.01 | <0.01 | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | 0.5 ^{#1} | 0.28 | 0.12 | 0.05 | 0.04 | 0.01 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | 0.10 | 0.07 | <0.02 | <0.02 | <0.02 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | 1.3 ^{#1} | 0.73 | 0.43 | 0.12 | 0.10 | 0.03 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | 0.11 | 0.06 | <0.02 | <0.02 | <0.02 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | 1.01 | 0.55 | 0.17 | 0.14 | 0.04 |
| Perfluoroalkyl Sulfonamides | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEIFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| PFAS | | | | | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - | - | - | - | - |
| Sum of PFAS | µg/L | 0.002 | | 1.44 | 0.76 | 0.17 | 0.14 | 0.04 |

Comments

#1 Surface water from other creeks.

Table 3: Upper Trigger Values: Cascade and Headstone Creeks Water
DITRDCSA Norfolk PFAS



| | | | Location Code | Cockpit_SW01 | PWS_HEAD_DAM |
|---|------|-------|---|--------------|--------------|
| | | | Field ID | COCKPIT_SW01 | PWS_HEAD_DAM |
| | | | Date | 18/11/2025 | 19/11/2025 |
| | | | Sample Type | Normal | Normal |
| | | | Lab Report No. | ES2537583 | ES2537583 |
| | Unit | EQL | Upper Trigger Value - Cascade and Headstone Creek | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | <0.01 | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | | 0.01 | <0.01 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | | 0.01 | <0.01 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | <0.02 | <0.02 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | 0.07 ^{#1} | 0.02 | <0.01 |
| Perfluoroalkyl Sulfonamides | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.05 | <0.05 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.02 | <0.02 |
| PFAS | | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - | - |
| Sum of PFAS | µg/L | 0.002 | | 0.02 | <0.01 |

Comments

#1 Surface water from Cascade Creek / Headstone Creek

Table 4: Lower Trigger Values: Public Water Use
DITRDCSA Norfolk PFAS



| | | | | Location Code | WC-04 | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCK DAM | WC_OMP05 |
|---|------|-------|------------------------------|----------------|------------|------------|------------|------------|-------------------|------------|
| | | | | Field ID | WC-04 | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCK DAM | WC_OMP05 |
| | | | | Date | 18/11/2025 | 17/11/2025 | 17/11/2025 | 18/11/2025 | 18/11/2025 | 18/11/2025 |
| | | | | Sample Type | Normal | Normal | Normal | Normal | Normal | Normal |
| | | | | Lab Report No. | ES2537583 | ES2537583 | ES2537583 | ES2537583 | ES2537583 | ES2537583 |
| | Unit | EQL | Lower Trigger Value - Public | | | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FIS) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | <0.002 | 0.12 | 0.06 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | <0.002 | 0.04 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | <0.005 | - | - | - | - | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.01 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | <0.002 | 0.02 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | | <0.002 | 0.28 | 0.12 | 0.05 | 0.04 | 0.01 | |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | <0.002 | 0.10 | 0.07 | <0.02 | <0.02 | <0.02 | |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | | <0.002 | 0.73 | 0.43 | 0.12 | 0.10 | 0.03 | |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | <0.002 | 0.11 | 0.06 | <0.02 | <0.02 | <0.02 | |
| Sum of PFHxS and PFOS | µg/L | 0.002 | 0.07 ^{#1} | <0.002 | 1.01 | 0.55 | 0.17 | 0.14 | 0.04 | |
| Perfluoroalkyl Sulfonamides | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEtFOSAA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.005 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.002 | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| PFAS | | | | | | | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - | - | - | - | - | - | |
| Sum of PFAS | µg/L | 0.002 | | <0.002 | 1.44 | 0.76 | 0.17 | 0.14 | 0.04 | |

Comments

- #1 Reticulated water supplies at public facilities (Risk ID 4)
 - Surface water from Mission Creek (Risk ID 2)
 - Surface water from Watermill Creek (Risk ID 5)
 - Groundwater at airport (airport bore) (Risk ID 3)

Table 5: Lower Trigger Values: Cattle Stock Watering
DITRDCSA Norfolk PFAS



| | | | Location Code | ID014_BORE | ID019_BORE | MC_OMP09 | MC_OMP10 |
|---|------|-------|--------------------------------------|------------|------------|------------|------------|
| | | | Field ID | ID014_BORE | ID019_BORE | MC_OMP09 | MC_OMP10 |
| | | | Date | 17/11/2025 | 20/11/2025 | 17/11/2025 | 19/11/2025 |
| | | | Sample Type | Normal | Normal | Normal | Normal |
| | | | Lab Report No. | ES2537583 | ES2537583 | ES2537583 | ES2537583 |
| | Unit | EQL | Lower Trigger Value - Stock Watering | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | 0.35 | 0.02 | 0.35 | 0.31 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | 0.11 | <0.02 | 0.10 | 0.10 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | - | - | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | 0.06 | <0.02 | 0.06 | 0.06 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.1 | <0.1 | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | 0.13 | <0.01 | 0.14 | 0.14 |
| Perfluoroalkane Sulfonic Acids | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | 0.33 ^{#1} | 2.90 | 0.07 | 3.99 | 3.49 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | 0.26 | 0.02 | 0.33 | 0.26 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | 1.2 ^{#1} | 2.20 | 0.12 | 2.43 | 2.04 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | 0.13 | <0.02 | 0.14 | 0.12 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | 0.31 | 0.03 | 0.31 | 0.24 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | 5.10 | 0.19 | 6.42 | 5.53 |
| Perfluoroalkyl Sulfonamides | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEIFOSAA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.02 | <0.02 | <0.02 | <0.02 |
| PFAS | | | | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - | - | - | - |
| Sum of PFAS | µg/L | 0.002 | | 6.57 | 0.26 | 7.98 | 6.86 |

Comments

#1 Surface water - Mission Creek water (used to water stock on e.g. properties A, B, C prior to management) (Risk ID 1)

Table 6: Lower Trigger Values Chicken Watering
DITRDCA Norfolk PFAS



| | |
|----------------|------------|
| Location Code | ID013_BORE |
| Field ID | ID013_BORE |
| Date | 17/11/2025 |
| Sample Type | Normal |
| Lab Report No. | ES2537583 |

| | Unit | EQL | Lower Trigger Value - ID013 | |
|---|------|-------|-----------------------------|-------|
| (n:2) Fluorotelomer Sulfonic Acids | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | <0.02 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | 0.9 ^{#1} | <0.01 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | <0.02 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | 1.3 ^{#1} | <0.01 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | <0.02 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | <0.02 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | <0.01 |
| Perfluoroalkyl Sulfonamides | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | | <0.02 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEtFOSAA) | µg/L | 0.002 | | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | <0.05 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | | <0.02 |
| PFAS | | | | |
| Sum of US EPA PFAS (PFOS + PFOA) | µg/L | 0.01 | | - |
| Sum of PFAS | µg/L | 0.002 | | <0.01 |

Comments

#1 Mission Creek water on property ID013 (used to water chickens prior to management) (Risk ID 1)

Table 7: Historical Water Analytical Results
PFAS OMP Norfolk Island



| | | Monitoring Zone | | | | | | | | | | | | | | | | | | |
|---|-------------|-----------------|-------------|------------|------------|--------------|------------|------------|------------|------------|------------|------------|------------|--------------|------------|------------|------------|------------|-----------|-----------|
| | | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public | Public |
| Location Code | Field ID | DEPOT_TANK3 | DEPOT_TANK3 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | DEPOT_TAP1 | ELEC_TAP1 | ELEC_TAP1 | FRE_TAP1 | FRE_TAP1 | FRE_TAP1 | FRE_TAP1 | FRE_TAP1 | FRE_TAP1 | Public |
| Date | Sample Type | 16/05/2023 | 24/06/2024 | 21/01/2020 | 11/03/2021 | 24/05/2022 | 16/05/2023 | 24/06/2024 | 17/11/2025 | 19/05/2023 | 19/11/2025 | 20/01/2020 | 12/03/2021 | 24/05/2022 | 18/05/2023 | 25/06/2024 | 19/11/2025 | 20/01/2020 | ES2317554 | ES2421874 |
| Lab Report No. | Unit | ES2317554 | ES2421874 | ES2002819 | ES2111278 | ES2218760-AC | ES2317554 | ES2421874 | ES2537583 | ES2317554 | ES2537583 | ES2002817 | ES2111256 | ES2218760-AC | ES2317554 | ES2421874 | ES2537583 | ES2002817 | ES2317554 | ES2421874 |
| | EQL | | | | | | | | | | | | | | | | | | | |
| | Unit | | | | | | | | | | | | | | | | | | | |
| | Unit | | | | | | | | | | | | | | | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | | | | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| 6:2 Fluorotelomer sulfonate (6:2 FTS) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | | | | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | | | | | | | | | | | | | | | | | |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorotridecanoic acid (PFTriDA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | | | | | | | | | | | | |
| Perfluorononane sulfonate (PFNS) | µg/L | 0.02 | | | | | | | | | | | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoropropanesulfonic acid (PFPS) | µg/L | 0.02 | | | | | | | | | | | | | | | | | | |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| Perfluoroalkyl Sulfonamides | | | | | | | | | | | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| N-methylperfluorooctane sulfonamidoacetic acid (NmFOSeAA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| N-ethylperfluorooctanesulfonamidoacetic acid (NeFOSeAA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSeA) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSeA) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSe) | µg/L | 0.005 | | | | | | | | | | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSeA) | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |
| PFAS | | | | | | | | | | | | | | | | | | | | |
| Sum of PFAS | µg/L | 0.002 | | | | | | | | | | | | | | | | | | |

Comments

#3 PFAS National Environmental Management Plan (HEPA 2020)

#4 NHMRC (2011)

#5 NHMRC (2019) Guidance on PFAS in Recreational Waters

#6 NHMRC (2011) - Health. Multiplied by a factor of x10

#7 For bioaccumulative contaminants, which include many PFAS, the framework specifies that the 99% species protection DGV should be used in:

- assessing toxicity and bioaccumulation in high conservation value ecosystems
- assessing bioaccumulation in slightly to moderately disturbed ecosystems.

2. Similarly, the 95% species protection DGV has been applied to highly disturbed systems.

#8 Where the criteria refer to the sum of PFOS and PFHxS, this means concentrations of PFOS only, PFHxS only, and the sum of the two (HEPA, 2025).

Table 7: Historical Water Analytical Results
PFAS OMP Norfolk Island



| Monitoring Zone | Watermill Creek | Watermill Creek | Watermill Creek | Watermill Creek | Watermill Creek | Watermill Creek |
|-----------------|------------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Location Code | WC_OMP04_DUCKDAM | WC_OMP05 | WC_OMP05 | WC_OMP05 | WC_OMP05 | WC_OMP05 |
| Field ID | WC_OMP04_DUCKDAM | TC_SW07 | WC_OMP05 | WC_OMP05 | WC_OMP05 | WC_OMP05 |
| Date | 18/11/2025 | 15/03/2021 | 24/05/2022 | 16/05/2023 | 24/06/2024 | 18/11/2025 |
| Sample Type | Normal | Normal | Normal | Normal | Normal | Normal |
| Lab Report No. | ES2537583 | ES2111268 | ES2218760-AC | ES2317554 | ES2421874 | ES2537583 |

| | Unit | EQL | Aquatic ecosystems DGV - high conservation value (99%) & slightly to moderately disturbed (95%) - freshwater | Aquatic ecosystems DGV - highly disturbed (90%) - freshwater | Vegetable Irrigation (Senversa, 2021c) | Stock Watering (Senversa, 2021c) | Drinking Water - Health | Recreational Water - Health | | | | | | |
|--|------|-------|--|--|--|----------------------------------|-------------------------|-----------------------------|--|-------|-------|-------|-------|-------|
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | | | | | | | | - | - | - | - | <0.05 |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | | | | | | | | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorotridecanoic acid (PFTDA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | 19 ^{#3,7} | 220 ^{#3,7} | | | 0.2 ^{#4} | 10 ^{#5} | | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | | | | | | |
| Perfluorononane sulfonate (PFNS) | µg/L | 0.02 | | | | | | | | - | - | - | - | <0.02 |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | 0.00023 ^{#3,7} | 0.13 ^{#3,7} | 1.4 | 0.33 | 0.005 ^{#4} | 2 ^{#5,8} | | 0.04 | <0.01 | 0.04 | <0.01 | 0.02 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | | | 0.8 | 1.2 | 0.03 ^{#4} | 2 ^{#5,8} | | 0.10 | <0.02 | 0.06 | 0.04 | 0.02 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | | | | | 1 ^{#4} | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Perfluoropropanesulfonic acid (PFPrS) | µg/L | 0.02 | | | | | | | | - | - | - | - | <0.02 |
| Sum of PFHxS and PFOS | µg/L | 0.002 | | | | | | | | 0.14 | <0.01 | 0.10 | 0.04 | 0.04 |
| Perfluoroalkyl Sulfonamides | | | | | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMFOFAA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| N-ethylperfluorooctanesulfonamidoacetic acid (NEFOFAA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| N-Ethyl perfluorooctane sulfonamide (EFOFA) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamide (MeFOFA) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | | | | | | | | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Perfluorooctane sulfonamide (FOFA) | µg/L | 0.002 | | | | | | | | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| PFAS | | | | | | | | | | | | | | |
| Sum of PFAS | µg/L | 0.002 | | | | | | | | 0.14 | <0.01 | 0.10 | 0.04 | 0.04 |

Comments
 #3 PFAS National Environmental Management Plan (HEPA 2020)
 #4 NHMRC (2011)
 #5 NHMRC (2019) Guidance on PFAS in Recreational Waters
 #6 NHMRC (2011) - Health. Multiplied by a factor of x10
 #7 For bioaccumulative contaminants, which include many PFAS, the framework specifies that the 99% species protection DGV should be used in:
 ± assessing toxicity and bioaccumulation in high conservation value ecosystems
 ± assessing bioaccumulation in slightly to moderately disturbed ecosystems.
 2. Similarly, the 95% species protection DGV has been applied to highly disturbed systems.
 #8 Where the criteria refer to the sum of PFOS and PFHxS, this means concentrations of PFOS only, PFHxS only, and the sum of the two (HEPA, 2025).

Table 8: Soil and Sediment Analytical Results
DITRDCA Norfolk PFAS



| | | | Location Code | AIRPORT_TRUCKFILL | BALL_SS3 | BALL_SS4 | BALL_SS5 | ID008_SS2 | WC_OMP04_DUCKDAM | | |
|---|-------|--------|---|---|---|--|------------|------------|------------------|---------|---------|
| | | | Field ID | A_TRUCKFILL_SS1 | BALL_SS3 | BALL_SS4 | BALL_SS5 | ID008_SS2 | WC_OMP04_SD | | |
| | | | Date | 20/11/2025 | 21/11/2025 | 21/11/2025 | 21/11/2025 | 20/11/2025 | 19/11/2025 | | |
| | | | Sample Type | Normal | Normal | Normal | Normal | Normal | Normal | | |
| | | | Lab Report No. | ES2537583 | ES2537583 | ES2537583 | ES2537583 | ES2537583 | ES2537583 | | |
| | Unit | EQL | Human Health - NEPM Setting 'A' - Residential | Human Health - NEPM Setting 'C' - Public Open Space | Human Health - NEPM Setting 'D' - Commercial / Industrial | Maintenance of Ecosystems - Commercial / Industrial ^{#10} | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | mg/kg | 0.0005 | | | | <0.0005 | <0.0005 | 0.0032 | <0.0005 | <0.0005 | |
| 6:2 Fluorotelomer Sulfonate (6:2 FTS) | mg/kg | 0.0005 | | | | <0.0005 | <0.0005 | 0.829 | 1.77 | <0.0005 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | mg/kg | 0.0005 | | | | <0.0005 | <0.0005 | 2.28 | 4.07 | <0.0005 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | mg/kg | 0.0005 | | | | <0.0005 | <0.0005 | 0.129 | 0.549 | <0.0005 | |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | mg/kg | 0.0002 | | | | 0.0005 | 0.0004 | 0.0918 | 0.0569 | <0.0002 | |
| Perfluorododecanoic acid (PFDoDA) | mg/kg | 0.0002 | | | | <0.0002 | 0.0004 | 0.0025 | 0.0713 | 0.0005 | |
| Perfluorononanoic acid (PFNA) | mg/kg | 0.0002 | | | | <0.0002 | 0.0008 | 0.0361 | 0.0260 | <0.0002 | |
| Perfluoropentanoic acid (PFPeA) | mg/kg | 0.0002 | | | | 0.0002 | 0.0002 | 0.167 | 0.0541 | <0.0002 | |
| Perfluorotetradecanoic acid (PFTeDA) | mg/kg | 0.0005 | | | | <0.0005 | <0.0005 | 0.0010 | 0.0348 | <0.0005 | |
| Perfluoroheptanoic acid (PFHpA) | mg/kg | 0.0002 | | | | <0.0002 | 0.0004 | 0.0462 | 0.0503 | <0.0002 | |
| Perfluorobutanoic acid (PFBA) | mg/kg | 0.001 | | | | <0.001 | <0.001 | 0.043 | 0.011 | <0.001 | |
| Perfluorodecanoic acid (PFDA) | mg/kg | 0.0002 | | | | <0.0002 | <0.0002 | 0.0182 | 0.0706 | 0.0004 | |
| Perfluorotridecanoic acid (PFTrDA) | mg/kg | 0.0002 | | | | <0.0002 | <0.0002 | 0.0005 | 0.0317 | 0.0003 | |
| Perfluoroundecanoic acid (PFUnDA) | mg/kg | 0.0002 | | | | <0.0002 | 0.0003 | 0.0049 | 0.0934 | 0.0004 | |
| Perfluorooctanoic acid (PFOA) | mg/kg | 0.0002 | 0.06 ^{#1} | 10 ^{#3} | 50 ^{#5} | 0.003 ^{#7} | 0.0012 | 0.0005 | 0.0601 | 0.0680 | 0.0004 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | mg/kg | 0.0002 | 0.003 ^{#2} | 1 ^{#4} | 20 ^{#6} | 0.003 ^{#9} | 0.0841 | 0.0437 | 0.0197 | 0.0084 | 0.0052 |
| Perfluoropentane sulfonic acid (PFPeS) | mg/kg | 0.0002 | | | | | <0.0002 | <0.0002 | <0.0002 | <0.0002 | <0.0002 |
| Perfluorohexane sulfonic acid (PFHxS) | mg/kg | 0.0002 | 0.003 ^{#2} | 1 ^{#4} | 20 ^{#6} | | 0.0009 | 0.0033 | 0.0028 | 0.0005 | <0.0002 |
| Perfluoroheptane sulfonic acid (PFHpS) | mg/kg | 0.0002 | | | | | <0.0002 | 0.0004 | <0.0002 | <0.0002 | <0.0002 |
| Perfluorodecanesulfonic acid (PFDS) | mg/kg | 0.0002 | | | | | 0.0002 | 0.0159 | 0.0105 | 0.0006 | <0.0002 |
| Perfluorobutane sulfonic acid (PFBS) | mg/kg | 0.0002 | | | | | <0.0002 | <0.0002 | <0.0002 | <0.0002 | <0.0002 |
| Sum of PFHxS and PFOS | mg/kg | 0.0002 | 0.003 ^{#2} | 1 ^{#4} | 20 ^{#6} | | 0.0850 | 0.0470 | 0.0225 | 0.0089 | 0.0052 |
| Perfluoroalkyl Sulfonamides | | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | mg/kg | 0.0005 | | | | | <0.0005 | <0.0005 | <0.0005 | <0.0005 | <0.0005 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | mg/kg | 0.0002 | | | | | <0.0002 | <0.0002 | <0.0002 | <0.0002 | <0.0002 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEtFOSAA) | mg/kg | 0.0002 | | | | | <0.0002 | <0.0002 | <0.0002 | <0.0002 | <0.0002 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | mg/kg | 0.0005 | | | | | <0.0005 | <0.0005 | <0.0005 | <0.0005 | <0.0005 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | mg/kg | 0.0005 | | | | | <0.0005 | <0.0005 | <0.0005 | <0.0005 | <0.0005 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | mg/kg | 0.0005 | | | | | <0.0005 | <0.0005 | <0.0005 | <0.0005 | <0.0005 |
| Perfluorooctane sulfonamide (FOSA) | mg/kg | 0.0002 | | | | | <0.0002 | <0.0002 | 0.0015 | <0.0002 | <0.0002 |
| PFAS | | | | | | | | | | | |
| Sum of PFAS | mg/kg | 0.0002 | | | | | 0.0877 | 0.0762 | 3.75 | 6.97 | 0.0072 |

Comments

- #1 PFAS NEMP 3.0: Health, Residential with garden/accessible soil (HIL A)
- #2 PFAS NEMP 3.0: Health, Residential with garden/accessible soil (HIL A). Value is for PFOS+PFHxS
- #3 PFAS NEMP 3.0: Health, Public open space (HIL C)
- #4 PFAS NEMP 3.0: Health, Public open space (HIL C). Value is for PFOS+PFHxS
- #5 PFAS NEMP 3.0: Health, Industrial/commercial (HIL D)
- #6 PFAS NEMP 3.0: Health, Industrial/commercial (HIL D). Value is for PFOS+PFHxS
- #7 PFAS NEMP 3.0: Ecological, indirect exposure. Also consider guideline values for direct exposure (0.005 mg/kg with reptiles, 10 mg/kg without reptiles).
- #8 PFAS NEMP 3.0: Ecological, indirect exposure. Also consider guideline value for direct exposure (1 mg/kg).
- #9 PFAS NEMP 3.0: Ecological, indirect exposure. For intensely developed sites with no secondary consumers and minimal indirect exposure, up to 0.14 mg/kg may be adopted. Also consider guideline value for direct exposure (1 mg/kg).
- #10 PFAS NEMP 3.0: applicable to areas of ecological significance, urban residential / public open space, and commercial / industrial site uses.

| Location Code | ID008_BIOTA1 | ID008_BIOTA2 | ID008_BIOTA3 | ID008_BIOTA4 | WC_OMP04_DUCKDAM |
|----------------------|--------------|--------------|--------------|--------------|------------------|
| Field ID | ID008_BIOTA1 | ID008_BIOTA2 | ID008_BIOTA3 | ID008_BIOTA4 | WC_BIOTA1 |
| Date | 20/11/2025 | 20/11/2025 | 20/11/2025 | 20/11/2025 | 19/11/2025 |
| Sample Type | Normal | Normal | Normal | Normal | Normal |
| Location Description | ID008 | ID008 | ID008 | ID008 | Duck Dam |
| Sample Comments | Zucchini | Tomato | Silverbeet | Parsley | Water Hyacinth |
| Lab Report No. | ES2537238 | ES2537238 | ES2537238 | ES2537238 | ES2537238 |

| | Unit | EQL | Food for Human Consumption - Fruits (all) | Food for Human Consumption - Vegetables (all) | | | | | |
|---|-------|--------|---|---|--------|--------|--------|--------|--------|
| Physical Parameters | | | | | | | | | |
| Weight of Sample Prepared | g | 0.1 | | | 2.1 | 0.9 | 2.0 | 3.6 | 2.9 |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | mg/kg | 0.0001 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| 6:2 Fluorotelomer Sulfonate (6:2 FIS) | mg/kg | 0.0001 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | mg/kg | 0.0002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | mg/kg | 0.0002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | mg/kg | 0.0001 | | | 0.002 | <0.001 | <0.001 | 0.003 | <0.001 |
| Perfluorododecanoic acid (PFDoDA) | mg/kg | 0.0005 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluorononanoic acid (PFNA) | mg/kg | 0.0001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluoropentanoic acid (PFPeA) | mg/kg | 0.0002 | | | <0.002 | <0.002 | <0.002 | 0.021 | <0.002 |
| Perfluorotetradecanoic acid (PFTeDA) | mg/kg | 0.002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluoroheptanoic acid (PFHpA) | mg/kg | 0.0001 | | | 0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorobutanoic acid (PFBA) | mg/kg | 0.0002 | | | <0.005 | <0.005 | <0.005 | 0.012 | <0.005 |
| Perfluorodecanoic acid (PFDA) | mg/kg | 0.0005 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorotridecanoic acid (PFTrDA) | mg/kg | 0.0005 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluoroundecanoic acid (PFUnDA) | mg/kg | 0.0005 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorooctanoic acid (PFOA) | mg/kg | 0.0001 | 0.0051 ^{#1} | 0.0088 ^{#1} | 0.003 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | |
| Perfluorononane sulfonate (PFNS) | mg/kg | 0.001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorooctanesulfonic acid (PFOS) | mg/kg | 0.0001 | 0.001 ^{#2} | 0.0011 ^{#1} | <0.001 | 0.001 | <0.001 | <0.001 | 0.001 |
| Perfluoropentane sulfonic acid (PFPeS) | mg/kg | 0.0001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorohexane sulfonic acid (PFHxS) | mg/kg | 0.0001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluoroheptane sulfonic acid (PFHpS) | mg/kg | 0.0001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluorodecanesulfonic acid (PFDS) | mg/kg | 0.0002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluorobutane sulfonic acid (PFBS) | mg/kg | 0.0001 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Perfluoropropanesulfonic acid (PFPrS) | mg/kg | 0.002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Sum of PFHxS and PFOS | mg/kg | 0.0001 | 0.001 ^{#2} | 0.0011 ^{#1} | <0.001 | 0.001 | <0.001 | <0.001 | 0.001 |
| Perfluoroalkyl Sulfonamides | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | mg/kg | 0.002 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | mg/kg | 0.0002 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEFOSAA) | mg/kg | 0.0002 | | | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | mg/kg | 0.001 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | mg/kg | 0.001 | | | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | mg/kg | 0.001 | | | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 |
| Perfluorooctane sulfonamide (FOSA) | mg/kg | 0.001 | | | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 |
| PFAS | | | | | | | | | |
| Perfluorooctane sulfonic acid (PFOS) - Branched | mg/kg | 0.001 | | | <0.001 | <0.001 | <0.001 | <0.001 | 0.001 |
| Perfluorooctane sulfonic acid (PFOS) - Linear | mg/kg | 0.001 | | | <0.001 | 0.001 | <0.001 | <0.001 | <0.001 |
| Sum of US EPA PFAS (PFOS + PFOA) | mg/kg | 0.0001 | | | - | - | - | - | - |
| Sum of PFAS | mg/kg | 0.0001 | | | 0.006 | 0.001 | <0.001 | 0.036 | 0.001 |

Comments

#1 FSANZ (2017) trigger points for investigation

#2 FSANZ (2017) LOR has been adopted as trigger point - LOR is to be adopted where LOR is higher than the trigger value in accordance with FSANZ

#3 Sum of PFHxS and PFOS calculated using 0.5x LOR for non-detects (calculated by Senversa as not reported by laboratory)



Appendix A: SAQP



PFAS Sampling and Analysis Quality Plan – Year 1 Ongoing Monitoring

Norfolk Island Airport

3 May 2022



Document Information

PFAS Sampling and Analysis Quality Plan – Year 1 Ongoing Monitoring, Norfolk Island Airport

Prepared by:

Senversa Pty Ltd

ABN: 89 132 231 380

Level 6, 15 William Street, Melbourne, VIC 3000

tel: + 61 3 9606 0070

www.senversa.com.au

Prepared for:

Department of Infrastructure, Transport, Regional Development and Communications

GPO Box 594

Canberra ACT 2601

| Revision | Date | Author | Reviewed | Approved | Detail |
|----------|------------|----------------|-----------------------|-----------------------|------------------|
| 0 | 3 May 2022 | Michelle Agnew | Christopher Sandiford | Christopher Sandiford | Draft for review |

Project Manager: Christopher Sandiford

Project Director: Michael Rehfish

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Senversa acknowledges the traditional custodians of the land on which this work was created and pay our respect to Elders past and present.



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Appendix A: Sample Nomenclature



Glossary and Acronyms

| Acronym | Definition | Acronym | Definition |
|---------------|---|--------------|---|
| ADWG | Australian Drinking Water Guidelines | HSEP | Health, Safety and Environment Management Plan |
| AFFF | Aqueous Film Forming Foam | ID | Identification |
| ALS | ALS Environmental Ltd | LOR | Limit of Reporting |
| AS | Australian Standard | LTV | Lower Threshold Value |
| ANZECC | Australian and New Zealand Environment and Conservation Council | NATA | National Association of Testing Authorities |
| ANZG | Australian and New Zealand Governments | NEPC | National Environment Protection Council |
| BoM | Bureau of Meteorology | NEPM | National Environment Protection Measure |
| CoC | Chain of Custody | NHMRC | National Health and Medical Research Council |
| CSM | Conceptual Site Model | NRMMC | Natural Resource Management Ministerial Council |
| DQO | Data Quality Objective | OMP | Ongoing Monitoring Plan |
| DO | Dissolved Oxygen | PFAS | Perfluoroalkyl and Polyfluoroalkyl Substances |
| DoH | Australian Government Department of Health | PFHxS | Perfluorohexane Sulfonic Acid |
| DSI | Detailed Site Investigation | PFOA | Perfluorooctanoic Acid |
| EPA | Environment Protection Authority | PFOS | Perfluorooctane Sulfonate |
| EC | Electrical Conductivity | PS | PFAS Source Zone |
| GME | Groundwater Monitoring Event | PSI | Preliminary Site Investigation |
| HBGV | Health Based Guidance Value | QA | Quality Assurance |
| HEPA | Heads of EPA | QC | Quality Control |
| HHERA | Human Health and Ecological Risk Assessment | RPD | Relative Percentage Difference |
| HHSV | Human Health Screening Values | | |



| Acronym | Definition |
|-------------|------------------------------------|
| SAQP | Sampling and Analysis Quality Plan |
| SPR | Source Pathway Receptor |
| SWL | Standing Water Level |
| SWMS | Safe Work Method Statement |
| TAT | Turnaround Time |

| Acronym | Definition |
|--------------|---|
| TOC | Total Organic Carbon |
| USEPA | United States Environmental Protection Agency |
| UTV | Upper Threshold Value |
| WHO | World Health Organisation |

| Unit of Measurement | Definition |
|---------------------|--------------------------------|
| L | Litres |
| Ha | Hectares |
| m AHD | Metres Australian Height Datum |
| m | Metres |
| mm | Millimetres |
| mV | Millivolts |

| Unit of Measurement | Definition |
|---------------------|---------------------------|
| km | Kilometres |
| m bgl | Metres below ground level |
| mg/L | Milligrams per litre |
| µg/L | Micrograms per litre |
| mg/kg | Milligrams per kilogram |



1.0 Introduction and Objectives

Senversa has been engaged to undertake an investigation of the nature and extent of perfluoroalkyl and polyfluoroalkyl substances (PFAS) at the Norfolk Island Airport (the site) and surrounding land. The site location and layout is shown in **Figure A1**.

In January 2020 Senversa commenced the Preliminary Site Investigation (PSI) which found that legacy aqueous film-forming foam (AFFF) containing PFAS was used on Norfolk Island from the early 1980s until 2015 to suppress liquid fuel fires and for fire training activities. These findings were confirmed in further sampling undertaken as a part of the Detailed Site Investigation (DSI) completed in October 2021, with potentially unacceptable risks identified in the DSI quantitatively assessed within the Human Health and Ecological Risk Assessment (HHERA) also completed in October 2021.

To manage some uses of water, Senversa prepared a PFAS Management Plan detailing the strategy for managing risks associated with PFAS impacts on the airport and across the island. This strategy includes an Ongoing PFAS Monitoring Plan (OMP). To guide the field works proposed to be undertaken during completion of the OMP Year 1 Sampling Event, Senversa has prepared this Sampling and Analysis Quality Plan (SAQP).

1.1 Objectives

1.1.1 OMP Objectives

The overall objective of the OMP is to establish the ongoing monitoring actions which are required to assess:

- Trends in PFAS concentrations in the environment.
- The effectiveness of the selected management options in managing current risks.
- Whether changing conditions exist which may result in changes in the risk profile (and therefore changes to the required management actions).

Information from the monitoring program will be used on an ongoing basis to identify whether the currently selected management action should change. Future changes to the management actions could be:

- **Additional** required actions (for instance where additional water uses are identified, or if PFAS concentrations in the environment increase).
- **Reduced** required actions (for instance where lower PFAS concentrations in the environment mean that previously established management actions are no longer necessary to manage risks).

1.1.2 SAQP Objectives

The objective of this SAQP is to detail the data collection tasks required to complete the proposed Year 1 Monitoring Event including the following:

- Describe the current understanding of the nature and extent of PFAS contamination at the site, based on sampling tasks completed on-site.
- Describe the rationale and data quality objectives for the proposed sampling program.
- Specify the proposed investigation locations and strategy.
- Outline the field methodologies for sample collection.
- Specify key analytical considerations.
- Specify the quality assurance and quality control (QA/QC) program.
- Identify assessment criteria.



2.0 Background

2.1 Key Site Information

The following summary of general information for the site and surrounds is considered relevant to the development of this SAQP.

Table 2-1 Key Site Information Summary

| Site Element | Relevant Information from Previous Investigations |
|------------------------------------|---|
| Location and Size | Norfolk Island is situated in the Pacific Ocean, approximately 1,676 kilometres (km) from Sydney. The site is located in the south-western portion of Norfolk Island. The site occupies approximately 120 hectares (ha). See Figure 1 for an overview of the site location and layout. |
| Land Uses | <u>On-site</u> The site is the Norfolk Island International Airport which comprises two runways and associated terminal buildings and carparks. The first runway of the airport was constructed on 25 December 1942 with the assistance of the United States Air Force to assist with war efforts. The airport contains 120 ha of land with 95 ha used for aviation purposes. <u>Off-site</u> The site is surrounded by rural properties and vegetated land to the north, south and west with the township of Burnt Pine to the north-east. |
| Geology and Soil Conditions | The site area is relatively flat with an elevation of 113 metres (m) above sea level. Soils are predominately derived from weathered Tertiary aged basaltic lava and tuff across the centre of the island, with Quaternary Aged alluvium and calcarenite present around much of the perimeter of the site extending inward from the coast for between 100 m and 500 m. |
| Hydrogeology | An upper aquifer is located across Norfolk Island in the base of porous alluvium and weathered basaltic rock. The groundwater moves towards sea level through a complex network of fractures and other interconnecting features in the volcanic bedrock (R.S Abell & A.C. Falkland, 1991). Groundwater is generally good quality and is suitable for domestic use. Groundwater type is classified as sodium chloride type with deuterium/oxygen correlation indicating direct groundwater infiltration. More than 450 bores (R.S Abell & A.C. Falkland, 1991) are known to be on the island. |

2.2 Regulatory Framework

For the purposes of this investigation the following federal guidance has been adopted:

- National Environment Protection (Assessment of Site Contamination) Amendment Measure (NEPM), National Environmental Protection Council (NEPC) (2013).
- PFAS National Environmental Management Plan (NEMP) 2.0, Heads of EPAs (HEPA) Australia and New Zealand (2020).
- Health Based Guidance Values for PFAS For Use in Site Investigations in Australia. Australian Government Department of Health (DoH) (2017).
- Australian and New Zealand Guidelines for Fresh and Marine Water Quality, Australian and New Zealand Governments (ANZG) (2018).



Senversa will adopt a QA/QC approach that is based on guidance from the following sources:

- Australian Standard (AS) 4482.1-2005 Guide to the investigation and sampling of sites with potentially contaminated soil, Part 1: Non-volatile and semi-volatile compounds.
- Schedule B (3) Guideline on Laboratory Analysis of Potentially Contaminated Soils, NEPM.
- United States Environmental Protection Agency (USEPA)- Guidance on Systematic Planning Using the Data Quality Objectives Process EPA QA/G-4.
- USEPA - Guidance on Environmental Data Verification and Data Validation EPA QA/G-8.

DRAFT



3.0 Identified PFAS Impacts

3.1 Norfolk Island Airport Description

Norfolk Island airport is an international airport with access to the airside portion of the airport strictly managed. Other uses of the site include the following:

- Fire Station.
- Bureau of Meteorology (BoM) weather station.
- Council offices.
- Freight forwarding office.
- Former drill ground.
- Waste depot.
- Wastewater treatment plant.

Low level concentrations of PFAS are present across all areas the airport, however concentrations of PFOS and perfluorohexane sulfonate (PFOS+PFHxS) appear to be highest at Source Area 4 (Current Drill Ground). This is where Legacy AFFF was used most recently for training in 2015 as shown on **Figure 3-1** below.



Figure 3-1: Surficial Soil Concentrations of PFOS+PFHxS



3.2 PFAS Impacted Surface Water Catchments

Surface water catchments outside of the airport with PFAS present that require management are Mission Creek and to a lesser degree in Watmill Creek Catchment.

3.2.1 Mission Creek Surface Water Catchment

Surface water samples from the Mission Creek catchment showed the highest concentrations at locations closest to PFAS source zones (PS) PS01 and PS02 at the airport (World War II Dam and MC_SW21). The pathway of PFAS from PS01 and PS02 into Mission Creek is considered to be both groundwater from source zones and surface water run off over PFAS-impacted soils on the airport through drainage lines, which is supported by the concentration in sediment sample MC_SD20.

PFAS concentrations consistently decreased further at each downstream location within Mission Creek (i.e., concentrations decreased with distance away from airport), with the exception of MC_SW25, which reported low levels of PFAS. The decrease in concentrations is shown in **Figure 3-2** below.

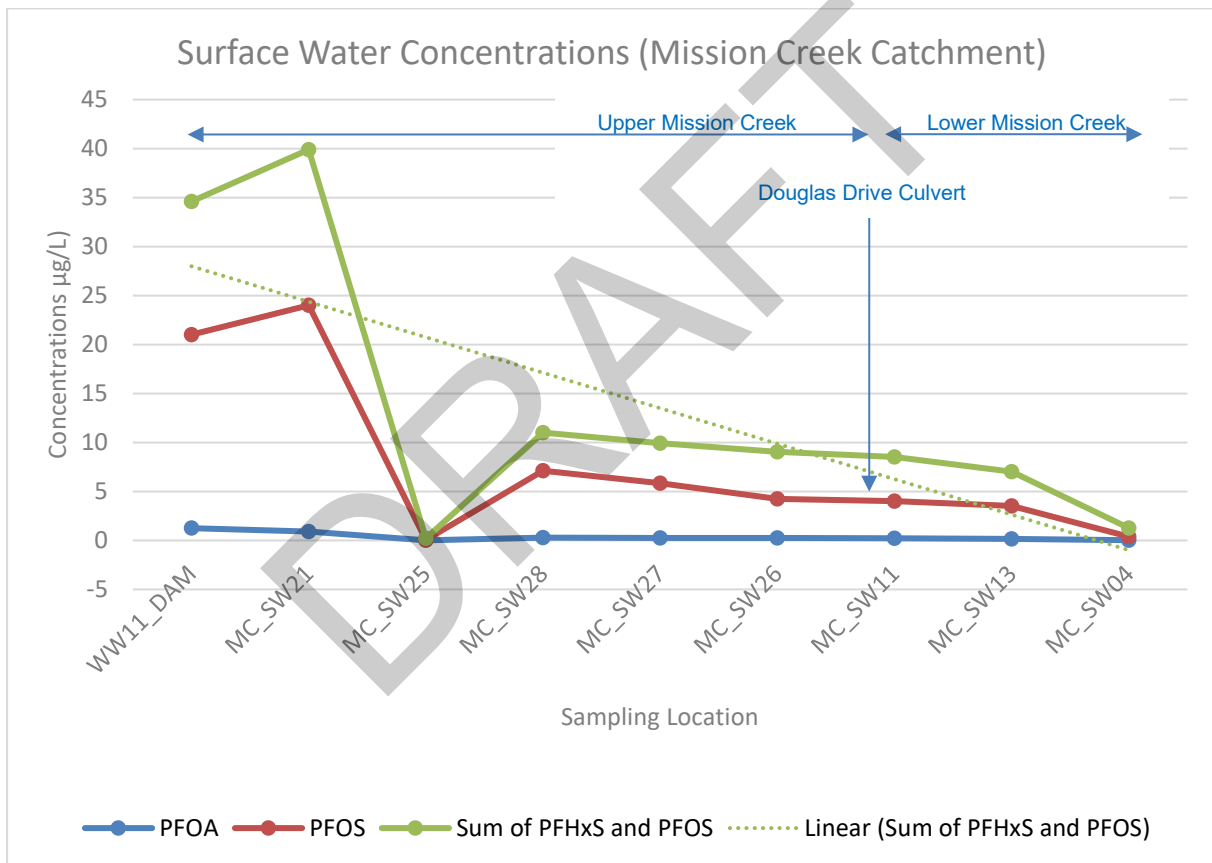


Figure 3-2: Surface Water PFAS Concentrations in Mission Creek

One surface water sample (MC_SW24) was collected from the upper Mission Creek catchment west of the waste depot (source zone PS03) on a separate Mission Creek tributary branch showed significantly lower PFAS concentrations than the tributary downgradient of PS01 and PS02.

A significant drop in PFAS concentrations was reported in MC_SW25, which is just after the confluence of two tributaries in the upper Mission Creek. There was limited evidence of surface water being further impacted down-gradient of PS04 with Mission Creek, adjacent to where Mission Creek sample MC_SW25 was collected.



It is noted MC_SW25 would be expected to receive run off from PS04 but may not receive flows from both tributaries of Mission Creek after the confluence at the exact sampling point. This is due to the creek bed being large and wide (creek bed and low-lying areas covered in substantial reed beds with moisture noted across most of the low-lying area). Additionally, the surface water sample (MC_SW25) may represent the water coming from upstream, rather than from PS04.

This indicates the highest PFAS impacts are likely to be from the northern tributary and hence from airport sources in the northern portion of the airport (PS01 and PS02).

3.2.2 Watermill Creek Catchment

Within the Watermill / Town Creek catchment, the highest PFAS concentration in surface water (TC_SW06 – PFOS+PFHxS: 1.14 micro grams per litre [$\mu\text{g/L}$]) was identified downstream of the Maintenance Depot (PS05). PFAS concentrations consistently decreased further at each downstream location before being below detection limits at the point of discharge into Emily Bay. The decrease in concentrations is shown in **Figure 3-2** below.

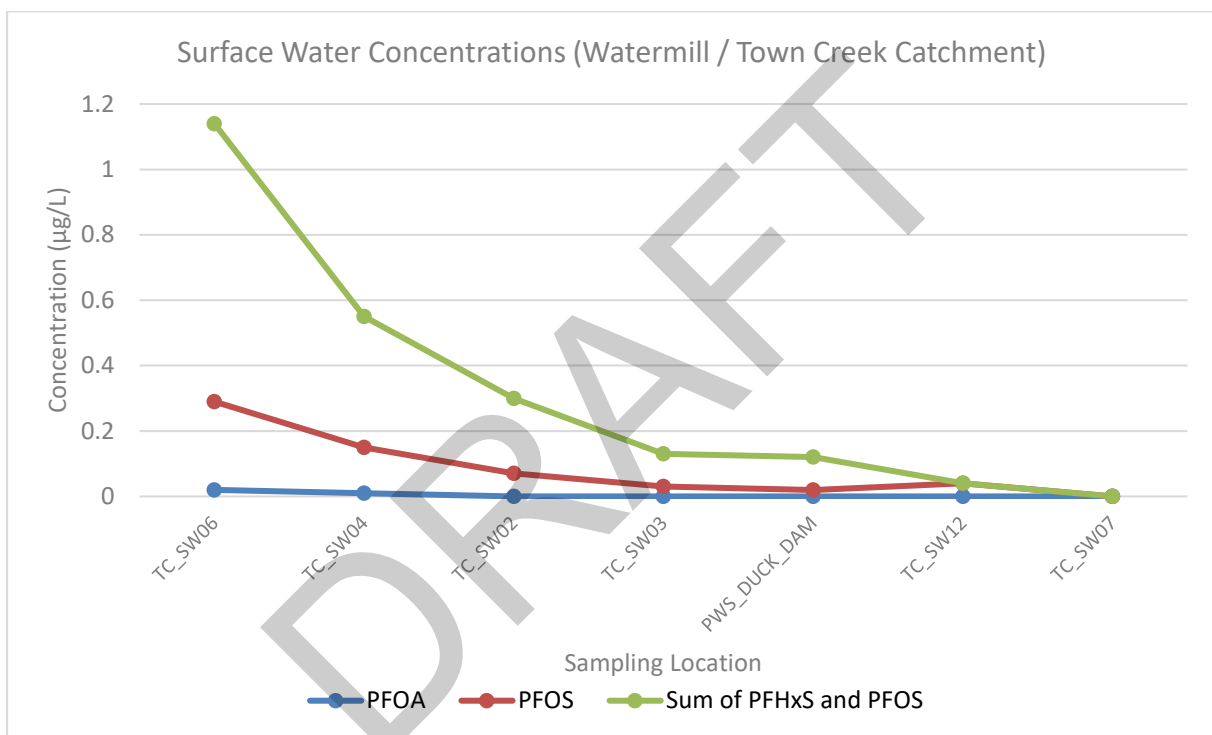


Figure 3-3: Surface Water PFAS Concentrations in Watermill / Town Creek Catchment

Two surface water samples (TC_SW05 and TC_SW13) were collected in the upper reaches of Watermill / Town Creek Catchment but were from different tributaries. TC_SW05 was below adopted criteria (95% levels for this catchment) and was collected on a different tributary to TC_SW06; location is in close proximity to the airport boundary, however it is not downgradient from any identified source zones. TC_SW13 was also collected from a separate upper reach away from the airport (circa 900 m from airport boundary) but is downgradient of source zone PS14 (perfumery). Concentrations of PFAS in TC_SW13 were above criteria, indicating PS14 is a potential source of PFAS



4.0 Investigation Strategy

4.1 Monitoring of surface water and sediment in creeks

4.1.1 Scope and rationale

More data is required to understand the range in concentrations in different creeks over time. Additionally, management actions on the airport may result in a decreasing trend in concentrations in creeks over time. In order to assess trends in PFAS concentrations in creeks on island, ongoing monitoring will be completed in a number of creeks, with the focus on Mission Creek and Watermill Creek which both receive run-off from the airport.

- The highest concentrations have been identified in Mission Creek and multiple sample locations are selected along Mission Creek in order to provide an ongoing understanding of the level and extent of PFAS impacts, and also to help assess which on-airport sources are contributing to the PFAS in Mission Creek. In the lower catchment of Mission Creek (i.e., further downstream) only sediment samples have been collected to-date as water was not present during sampling.
- Concentrations in Watermill Creek are much lower than in Mission Creek and decrease along the length of Watermill Creek (with the highest concentrations measured in the upper part of the catchment). Downstream of Watermill Dam (Duck Dam), concentrations are below the health-based guidance value (HBGV) for drinking water. Ongoing monitoring is required to confirm the extent of impacts within Watermill Creek over time.
- Samples will also be collected from Cascade Creek and Headstone Creek. Low concentrations (below the HBGV for drinking water) have been measured in these creeks to-date. Ongoing monitoring is required to assess changing conditions.

Where practicable, monitoring should be scheduled for times when water is likely to be present (i.e., after periods of rainfall and not in drought conditions). On the first round of monitoring, paired surface water / sediment samples should be collected as only one round of monitoring data is currently available in most locations. On subsequent monitoring rounds, surface water will be collected from all locations if present. If surface water is absent, a sediment sample will be collected instead.



4.1.2 Sampling locations and frequency

The sample locations are depicted on **Figure A1** and summarised below.

Table 4-1: OMP Creek sampling locations

| Location | Number of locations | Sample IDs | Notes | Frequency of sampling; sampling media |
|-----------------|---------------------|-------------------------------|---|---|
| Mission Creek | 12 | WWII_DAM, MC_OMP01 – MC_OMP11 | MC_OMP08 represents creek water within a paddock accessible to cattle prior to management (see also Section 4.3 for discussion of monitoring of managed stock water). MC_OMP10 targets Mission Pool where water has not been present on previous sampling rounds; if water is present anywhere within Mission Pool, it should be sampled. | Annual First round: surface water and sediment Subsequent rounds: water only, or sediment if water is absent |
| Watermill Creek | 5 | WC_OMP01 – WC_OMP05 | | |
| Cascade Creek | 1 | COCKPIT_SW01 | | |
| Headstone Creek | 1 | PWS_HEAD_DAM | | |

4.2 Monitoring of water utilised for irrigation

4.2.1 Scope and rationale

There are two properties in the Mission Creek catchment which use water from Mission Creek for produce irrigation. The risks associated with this have been assessed to be low and acceptable based on current data. Ongoing monitoring is required to assess if the risk profile might change.

4.2.2 Sampling locations and frequency

The sample locations are depicted on **Figure A1** and summarised below:

Table 4-2: Mission Creek irrigation water sampling locations

| Location | Number of locations | Sample IDs | Frequency of sampling; sampling media |
|---------------|---------------------|--------------------------|---|
| Mission Creek | 2 | ID013_SW01 ID016_BORE | Annual - Point of use water sampling |



4.3 Monitoring of water for which stock watering use is currently managed

4.3.1 Scope and rationale

There are three properties in the Mission Creek catchment where cattle previously had access to water from Mission Creek for stock watering. There was one property where water from Mission Creek was used for chicken drinking water. The PFAS Management Plan recommends that these uses are managed going forward. Monitoring of these water sources is required to assess trends in concentrations. If concentrations decrease, management may be no longer required.

4.3.2 Sampling locations and frequency

The sample locations are depicted on **Figure A1** and summarised below:

Table 4-3: Managed stock water sampling locations (Mission Creek)

| Location | Previous use (prior to management) | Number of locations | Sample IDs | Notes | Frequency of sampling; sampling media |
|------------------------|------------------------------------|---------------------|--|---|--|
| Mission Creek | Cattle stock watering | 4 | ID014_BORE | HHERA Property A | Annual Point of use water sampling |
| | | | ID015_BORE | HHERA Property B | |
| | | | MC_OMP08 | MC_OMP08 represents creek water within a paddock accessible to cattle prior to management (HHERA Property C). See also Section 4.1 for discussion of creek monitoring. | |
| | | | MC_OMP10 | MC_OMP10 represents Mission Pool, which was accessible to cattle prior to management although water has not been identified in the monitoring undertaken by Senversa. See also Section 4.1 for discussion of creek monitoring. | |
| Chicken drinking water | 1 | ID013_SW01 | This water is also used for irrigation (see Section 4.2) | | |



4.4 Monitoring of Airport Bore and facility water supplies

4.4.1 Scope and rationale

There are a number of water supplies in public facilities which were previously used for drinking water / domestic use, but where PFAS impacts above the HBGV for drinking water are currently present. These uses are currently managed (e.g., through the provision of alternate water supplies).

Monitoring of these water supplies is required to assess trends in concentrations. If concentrations decrease, management of water use may be no longer required. It is noted that new reticulated supplies are planned for some facilities at the airport including the new fire station.

4.4.2 Sampling locations and frequency

Table 4-4: Managed public water supply sampling locations

| Location | Number of locations | Sample IDs | Notes | Frequency of sampling; sampling media |
|-----------------------------|---------------------|---|--|---------------------------------------|
| On-airport | 5 | AIRPORT_BORE | | Annual |
| | | FRE_TAP1 | Fire station kitchen; new rainwater tanks recently installed | Point of use water sampling |
| | | FRE_TAP2 | Fire hydrant (used for fire testing). New water supply is being put in place. Testing to be superseded with new supply for fire testing after switchover | |
| | | A_TAP1 | Airport terminal female toilets | |
| | | A_TAP4 | Mech/maintenance shed adjacent airport terminal and gate 1 | |
| Off-airport: Works depot | 4 | DEPOT_TAP, DEPOT_TANK1, DEPOT_TANK2, DEPOT_TANK3 | | |

4.5 Summary of samples to be collected

All non-private sample locations to be targeted in the ongoing monitoring program are depicted on **Figure A1**.

Table 4-5: OMP sampling location summary

| Sample purpose | Location | Number of locations | Sample IDs | Frequency of sampling; sampling media |
|----------------|-----------------|---------------------|-----------------------------------|--|
| Creek sampling | Mission Creek | 12 | WWII_DAM, MC_OMP01 to MC_OMP11 | Annual First round: surface water and sediment |
| | Watermill Creek | 5 | WC_OMP01 to WC_OMP05 | |
| | Cascade Creek | 1 | COCKPIT_SW01 | |



| Sample purpose | Location | Number of locations | Sample IDs | Frequency of sampling; sampling media |
|------------------------|-----------------|---------------------|---|---|
| | Headstone Creek | 1 | PWS_HEAD_DAM | Subsequent rounds: water only, or sediment if water is absent |
| Irrigation water | Mission Creek | 2 | ID013_SW01 ID016_BORE | Annual Point of use water sampling |
| Managed stock water | Mission Creek | 2* | ID014_BORE ID015_BORE (MC_OMP08*) (MC_OMP10*) (ID013_SW01*) | |
| Managed water supplies | On-airport | 5 | AIRPORT_BORE FRE_TAP1 FRE_TAP2 A_TAP1 A_TAP4 | |
| | Off-airport | 4 | DEPOT_TAP DEPOT_TANK1 DEPOT_TANK2 DEPOT_TANK3 | |
| Total | | 32 | | |

Notes: * There are 5 samples relevant for managed stock watering, however only two unique samples (ID014_BORE and ID015_BORE, used for cattle watering) not also collected for another purpose. MC_OMP08 and MC_OMP10 (creek locations with possible cattle access in the absence of management) are included in the total sample numbers for creek sampling and ID013_SW01 (used for chicken watering) is included in the total sample numbers for irrigation water sampling

4.6 DQOs

The data quality objective (DQO) process is a systematic planning approach outlined in the NEPM (2013) that is used to define the purpose of the investigation to be undertaken and the type, quantity and quality of data needed to inform decisions relating to the assessment of site contamination. Proposed DQOs for the ongoing monitoring are outlined in the table below.

Table 4-6: DQO Summary

DQO Seven-step Process

1. State the problem.

Elevated concentrations of PFAS have been reported in the Airport Bore (groundwater at and adjacent to the airport) and in water supplies at a number of public facilities which were historically supplied with water from the airport bore. The risks associated with these concentrations are currently managed (i.e., groundwater is not currently used for drinking).

In addition, PFAS has been identified in creeks which collect run-off from the airport, with the highest concentrations identified in Mission Creek. These elevated concentrations are not considered to pose a significant risk to human health from recreational direct contact, however they are contributing to a potentially elevated risk for ecosystem receptors.

In addition, potentially elevated exposures have not been excluded for cattle product or chicken egg consumption where livestock have access to Mission Creek water for drinking. Management of livestock access to water from Mission Creek catchment (surface water and/or groundwater) is therefore currently required. Risks have been assessed to be low and acceptable for livestock drinking water from other creeks.



DQO Seven-step Process

Risks from produce consumption are assessed to be low and acceptable based on the current concentrations measured at properties within the Mission Creek catchment where water is used for produce irrigation. Risks are also assessed to be low and acceptable where water from other creeks is used for produce irrigation.

Further, the concentration trends in surface water, sediment and groundwater are not well understood based on the available monitoring data.

2. Identify the decision/goal of the study.

The goal is to monitor the nature and extent of PFAS impacts and identify trends and changes to PFAS impacts in the environment on and off-site that may alter the understanding or assessment of identified risks into the future.

3. Identify the information inputs.

The primary inputs are considered to be PFAS concentrations in groundwater, surface water and sediment.

4. Define the boundaries of the study.

Ongoing monitoring will be undertaken at a selected number of surface water locations and point of use water supplies at and surrounding the site.

5. Develop the analytical approach/decision rules.

The data will be used in the to assess whether site-derived PFAS has changed in nature and extent which may alter the understanding or assessment of identified risks into the future to human or ecological receptors.

The useability of the data will be assessed in terms of accuracy and reliability in forming conclusions on the concentrations within the samples collected, based on guidance from the relevant sources listed above. The data quality objectives, measures and acceptance criteria to be adopted for monitoring should be outlined in the SAQP to be developed for each monitoring round.

It is required that, as a minimum, the following type and frequency of quality control samples be collected.

Field duplicates (intra laboratory and inter laboratory) samples at a rate of at least 1 in 10 separately groundwater and surface water.

Rinsate blanks where equipment decontamination will be necessary (e.g., groundwater sampling) at a rate of one per day per set of equipment.

As part of the reporting, the results of the monitoring should be used to assess trends using an appropriate statistical approach such as Mann-Kendall methods, or similar, to identify increases, declines or stabilisation of concentrations across monitoring rounds to a specified statistical confidence limit based on the amount of data collected over time.

Some examples of the decisions to be made from investigation results include:

If detections of PFAS are reported in field blanks or rinsate blanks, then consider if there is a potential for cross contamination between sample locations and what impact this has on conclusions of trends.

If reported PFAS concentrations in relevant sample locations increase above the defined upper threshold values (UTVs) defined in the OMP (and/or an increasing trend is identified), then consider further risk assessment to assess whether additional management measures are required.

If reported PFAS concentrations in relevant sample locations decrease below the lower threshold values (LTVs) defined in the OMP (and/or a decreasing trend is identified), then consider further risk assessment to assess whether management measures can be reduced.

6. Specify performance or acceptance criteria.

Adopted screening criteria, LTVs and UTVs defined in the OMP will be used to provide a screening level of results obtained during sampling and assess if risk revision is required.

A data validation checklist with specific acceptance criteria and discussion of results must be documented and reviewed as part of the SAQP development.

At the end of the initial monitoring period, reporting should assess trends in concentrations. This should include development and use of a statistical based decision criteria to assess the significance of trends. Where significant trends are identified, the requirement for further monitoring, assessment and/or management (in the case of an increasing trend) or cessation of monitoring (in the case of a decreasing trend) will be assessed.

7. Develop the plan for obtaining data.

The overarching scope and methodology is provided in this OMP. Prior to each sampling event, a SAQP should be developed which assesses the appropriateness of sample locations, sampling methodologies and risk screening/assessment criteria. The SAQP is to outline the optimum manner to collect the data required to meet the objectives for the assessment and which will meet the project DQOs.

Permission to access sampling locations on public and private properties is to be confirmed prior to sampling.



4.7 Health, Safety and Environment Management

A Health, Safety and Environment Management Plan (HSEP) will be prepared for the investigation to outline how safety and the environment will be managed during field investigations. This will include site specific risk assessment, safe work method statements (SWMS) and waste management plan.

- All Senversa staff involved in the site works will be inducted to the HSEP.
- Senversa personnel will have sufficient information, instruction, training and competency to safely undertake work at the site. Minimum training requirements for personnel will be listed in the HSEP and should be reviewed for all field work.
- Senversa will complete necessary inductions (to be confirmed on arrival on Island) and comply with Norfolk Island Airport site rules and regulations whilst on the site.

4.8 Sampling and Investigation Methodology

The following section describes the methodology to be adopted by field personnel in the conduct of the surface water, sediment, groundwater, soil and biota sampling.

4.8.1 Specific Sampling Requirements

The table below summarises the specific methodology and investigation techniques to be adopted for the various proposed sampling tasks.

Table 4-7: Summary of Specific Sampling Requirements

| Sample Type | Detail |
|---------------|---|
| Surface Water | <ul style="list-style-type: none"> • Surface water samples will be collected either directly into the sampling containers or using a hand-held sampling device (e.g., Swing Sampler) with subsequent decanting into the laboratory sampling containers. • Surface water samples will be collected prior to sediment to minimise disturbance and avoid excess sediment load in the water sample. • Direct surface water sampling methods that are used will depend on location access. Sampling of deeper drains may have health and safety risks associated with access, and an appropriate sampling method for that location will be reviewed and applied. • Water quality parameters (pH, redox, dissolved oxygen, electrical conductivity, and temperature) will be recorded at each sample location using a calibrated water quality meter. <p>Sample locations will be selected on island prior to works based on location and safe access requirements.</p> |
| Sediment | <p>Sediment samples will be collected using a gloved hand with the aid of a small hand trowel/shovel and transferred into sample jars using disposable nitrile gloves.</p> <ul style="list-style-type: none"> • Sediment samples will be collected from the base of the waterbody (i.e., 0 - 0.05 m below the top of the sediment layer) beneath any surface water (if present). • Sampling of potentially deeper waterbodies may have health and safety risks associated with access, and an appropriate sampling method for that location will likely utilise hand tools such as shovels or trowels. • Access to the exact location indicated for sampling may be dependent on ground cover, the angle of any banks, etc. Therefore, the location where samples are collected may vary. • Where grass, reeds or vegetation growth covers the sediment, this material will be moved aside using hand tools, with the sediment sample collected from directly underneath the vegetation layer. • Any tools used (trowel/shovel) will be decontaminated prior to use. |



| Sample Type | Detail |
|---|--|
| Groundwater (point of use water) Sampling | <ul style="list-style-type: none"> The outlet to be sampled is to be determined as the first extraction discharge point within the water supply infrastructure (i.e., closed tap to the extraction well discharge). The outlet / tap will be turned on to flush it of water for approximately 30 seconds by using a smooth flowing water stream at moderate pressure. Where a line to be sampled supplied both hot and cold water, only the cold water will be sampled. If a tap is not available, a disposable bailer will be placed inside the bore to collect the sample. Samples will be placed directly into laboratory supplied bottles. Water quality parameters (pH, redox, dissolved oxygen, electrical conductivity, and temperature) will be recorded at each sample location using a calibrated water quality meter. |

4.8.2 General Sampling Requirements

The following table details general sampling techniques associated with the site works.

Table 4-8: General Sampling Requirements

| Activity | Description and Further Information |
|---|--|
| Field Parameter Measurement | <p>Field water quality parameters will be measured using a water quality meter prior to sampling for all surface water and bore water sampling. The parameters include pH, electrical conductivity, dissolved oxygen, oxidation reduction potential (redox) and temperature.</p> <p>For soil and sediment sampling, field observations will be noted of characteristics such as colour, particle size, odour, discoloration, presence of unusual materials such as waste, etc.</p> |
| Photographs | A photograph of the sampling location will be taken at each sampling location for record. |
| Location Survey | All sample locations will be logged using the ArcGIS "Collector" application to enable the location of each sample to be uploaded each evening once connected to the internet. |
| Sample Handling and Preservation | <ul style="list-style-type: none"> Samples will be placed into laboratory-supplied jars and bottles containing appropriate preservatives for the selected analytes to be tested. Samples will be collected and stored on bagged ice prior to and during transit to the laboratory to minimise sample degradation. Sample bottles will be filled to the top with no head space and splashing during filling should be prevented. All samples collected will be recorded on field logs sheets. Chain of Custody (CoC) forms will be completed for transport. Quality control samples will be collected during the sampling program as per Section 4.10.2. |
| Waste Disposal | Water purged from taps or bores is expected to be minimal in volume and will be disposed at the ground surface. |
| Equipment Calibration | Equipment requiring calibration (water quality meter) for environmental assessment purposes will be calibrated by the supplier or by Senversa staff prior to use. Relevant calibration certificates will be provided in the report. |



| Activity | Description and Further Information |
|---|---|
| Avoidance of Cross Contamination | <p>Sampling procedures used to prevent cross contamination will consider the guidance provided in Appendix 1 of the Interim Guideline of the Assessment and Management of PFAS (WA DER, 2016¹) during site works and involve:</p> <p>Samples will be placed into laboratory-supplied jars / bottles appropriate for PFAS sampling (i.e., without Teflon liners).</p> <p>Decontamination of re-usable sampling equipment will be completed between sampling locations, using a potable water wash, and rinse with potable water.</p> <p>Use of dedicated disposable latex free gloves that will be replaced between each sample collection and location.</p> <p>Quality control samples to assess cross contamination will be collected during the sampling program as per Section 4.10.2.</p> |

4.9 Sample Nomenclature

Sample nomenclature will be based on sample matrix type (surface water, sediment, groundwater). The proposed sample nomenclature to be used is presented in **Appendix A**.

4.10 Laboratory Analysis

All groundwater, surface water, sediment, soil and biota samples will be submitted to chemical laboratories (ALS Environmental Pty Ltd as primary laboratory and Eurofins Environmental Pty Ltd as secondary laboratory) that are NATA accredited for the methods used.

The following PFAS analyses (extended suite of 28 individual PFAS) will be completed on samples collected and scheduled for analysis:

- Perfluorobutane sulfonic acid (PFBS)
- Perfluoropentane sulfonic acid (PFPeS)
- Perfluorohexane sulfonic acid (PFHxS)
- Perfluoroheptane sulfonic acid (PFHpS)
- Perfluorooctane sulfonic acid (PFOS)
- Perfluorodecane sulfonic acid (PFDS)
- Perfluorobutanoic acid (PFBA)
- Perfluoropentanoic acid (PFPeA)
- Perfluorohexanoic acid (PFHxA)
- Perfluoroheptanoic acid (PFHpA)
- Perfluorooctanoic acid (PFOA)
- Perfluorononanoic acid (PFNA)
- Perfluorodecanoic acid (PFDA)
- Perfluoroundecanoic acid (PFUnDA)
- Perfluorododecanoic acid (PFDoDA)
- Perfluorotridecanoic acid (PFTrDA)
- Perfluorotetradecanoic acid (PFTeDA)
- Perfluorooctane sulfonamide (FOSA)
- N-Methyl perfluorooctane sulphonamide (MeFOSA)
- N-Ethyl perfluorooctane sulfonamide (EtFOSA)
- N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE)
- N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE)
- N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA)
- N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA)
- 4:2 Fluorotelomer sulfonic acid (4:2 FTS)
- 6:2 Fluorotelomer sulfonic acid (6:2 FTS)
- 8:2 Fluorotelomer sulfonic acid (8:2 FTS)
- 10:2 Fluorotelomer sulfonic acid (10:2 FTS)

¹ WA DER 2016. *Interim Guidelines on the Assessment and Management of Perfluoroalkyl and Polyfluoroalkyl Substances – Contaminated Sites Guidelines*. Government of Western Australia, Department of Environment Regulations. February 2016.



Sediment samples will additionally be analysed for pH and total organic carbon (TOC).

The following laboratory limits of reporting, where achievable, will be requested from the laboratories:

- Sediment – <0.005 milligrams per kilogram (mg/kg)
- Water – <0.01 µg/L

All PFAS samples collected will be analysed on a standard 5-7day turn-around time (TAT). However, the time required for transport of samples from site to Sydney laboratories will be in addition to any laboratory guaranteed TAT. Senversa notes that the standard quarantine turn-around time at Sydney Airport is 7 days.

4.10.1 Physical Parameters

In addition to the collection of samples for laboratory analysis, the parameters outlined in the table below will also be recorded.

Table 4-9: Physical Parameters to be Assessed for Different Media

| Sampling Media | Parameters |
|----------------------------------|---|
| Sediment | Logged to AS1726:2017 ² ; visual and olfactory observations. |
| Groundwater | Physio-chemical parameters as per EPA Publication 669 ³ , obtained during sampling; visual and olfactory observations. |
| Surface Water / Tank & Tap Water | Physio-chemical parameters, obtained during purging and sampling; visual and olfactory observations. |

4.10.2 Quality Assurance Procedures

The data QA/QC procedures to be adopted must provide a consistent approach to evaluation of whether the DQOs required by the project have been achieved. The process focuses on assessment of the useability of the data in terms of accuracy and reliability in forming conclusions on the condition of the element of the environment being investigated.

Table 4-10: Data Quality Objectives for QA/QC Elements

| QA/QC Element | Data Quality Objectives |
|--------------------------------|--|
| Analytical Laboratories | All methods to be used will be NATA accredited. Changing these arrangements must be justified with detailed assessment and comparison of laboratory methods and analytical reference standards used. |
| Turnaround Times | A standard laboratory analysis TAT of 5-7 days will be requested for all samples submitted for analysis. |

² AS1726:2017. *Geotechnical Site Investigations*. 5 February 2017.

³ EPA Victoria 2022. *Groundwater Sampling Guidelines*. Publication 669.1. February 2022.



QA/QC Element**Data Quality Objectives**

Analytical QA/QC Guidance

The QA/QC approach must be based on guidance from the following sources:

- AS4482.1-2005 Guide to the investigation and sampling of sites with potentially contaminated soil, Part 1: Non-volatile and semi-volatile compounds.
- NEPC 2013, Schedule B (3) Guideline on Laboratory Analysis of Potentially Contaminated Soils.
- USEPA - Guidance on Systematic Planning Using the Data Quality Objectives Process EPA QA/G-4.
- USEPA - Guidance on Environmental Data Verification and Data Validation EPA QA/G-8.

QA/QC Procedures

The QA/QC procedures applied will include the use of equipment decontamination, Chain of Custody documentation, laboratory data verification and the use of quality control samples in accordance with Section 8.2 of AS4482.1-2005.

All rinsate blanks and laboratory method blanks have an acceptance limit of concentrations below the laboratory limit of reporting. Detection of an analyte in a rinsate sample must trigger an assessment of the decontamination process followed by an assessment if the analyte reported is a contaminant of interest or if it impacts the validity of the assessment data.

The % relative percentage difference (RPD) for field and laboratory duplicates must meet the NEPM (NEPC, 2013) guidelines.

A data quality assurance review, which includes a data validation checklist with specific acceptance criteria for each batch of samples. Data quality will be checked against the data validation checklist as results become available throughout the investigation program to establish if further checking of precision or accuracy is required as the investigation progresses.



5.0 Reporting Requirements

On completion of the OMP Year 1 field program, Senversa will prepare an interpretive report on the nature and extent of PFAS. The report will include the following:

- An executive summary.
- A summary of the project objectives and scope of works consistent with those outlined in this SAQP.
- A summary of the environmental setting of the site, including the site-specific topography, geology and hydrogeology.
- A summary of the surface water, sediment and groundwater sampling methodology used.
- Analytical results, including quality assurance assessment.
- Qualitative risk assessment using published and site-specific data.
- Trend analysis and assessment of results against the upper and lower trigger values as defined in the OMP.
- Updated conceptual site model including sources, pathways and receptor linkages and identified data gaps.
- Figures including site and sample location plans and PFAS criteria exceedances.
- Tables and appendices of supporting documentation from field investigations.
- Conclusions on risks to sensitive receptors and assessment against defined trigger values and decision tree as outlined in the OMP.



6.0 References

- ANZG 2018. *Australian and New Zealand Guidelines for Fresh and Marine Water Quality*. Australian and New Zealand Governments and Australian state and territory governments, Canberra ACT, Australia. Available at www.waterquality.gov.au/anz-guidelines.
- AS1726:2017. *Geotechnical Site Investigations*. 5 February 2017.
- AS4482.1:2005. *Guide to the Investigation and Sampling of Sites with Potentially Contaminated Soil, Part 1: Non-Volatile and Semi-Volatile Compounds*. 2 November 2005.
- Department of Health 2017. *Health Based Guidance Values for PFAS For Use in Site Investigations in Australia*. Australian Government Department of Health.
- HEPA 2020. *PFAS National Environmental Management Plan - Version 2.0*. January 2020.
- NEPC, 2013. *National Environment Protection (Assessment of Site Contamination) Amendment Measure 2013 (No. 1)*, Canberra: National Environment Protection Council.
- NHMRC & NRMCC, 2011. *Australian Drinking Water Guidelines*, National Water Quality Management Strategy Document 6: National Health and Medical Research Council & Natural Resource Management Ministerial Council.
- R.S Abell & A.C. Falkland 1991. *The hydrogeology of Norfolk Island, South Pacific Ocean*.
- Senversa, 2020. *Preliminary Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport*. 13 March 2020.
- Senversa 2021b. *Detailed Site Investigation into Per- and Polyfluoroalkyl Substances (PFAS), Norfolk Island Airport, Revision 5*. 12 November 2021.
- Senversa 2021c. *Human Health and Ecological Risk Assessment (PFAS), Norfolk Island Airport, Revision 3*. 12 November 2021.
- Senversa 2021d. *Ongoing Monitoring Plan, Norfolk Island Airport, Revision 0*. 24 November 2021.
- Senversa 2021e. *PFAS Management Plan, Norfolk Island Airport, Revision 1*. 10 December 2021.
- USEPA 2000. *Guidance on Systematic Planning Using the Data Quality Objectives Process, EPA QA/G-4*: United States Environmental Protection Agency.
- USEPA 2002. *Guidance on Environmental Data Verification and Data Validation*, Washington D.C: United States Environmental Protection Agency.
- EPA Victoria 2022. *Groundwater Sampling Guidelines*. Publication 669.1. February 2022.
- WA DER 2016. *Interim Guidelines on the Assessment and Management of Perfluoroalkyl and Polyfluoroalkyl Substances – Contaminated Sites Guidelines*. Government of Western Australia, Department of Environment Regulations. February 2016.



Figures

Figure 1: PFAS Management Areas

DRAFT



| Legend: | |
|---|---|
| Surface Water / Sediment (Cascade and Headstone Creeks) | Approximate Airport Boundary |
| Surface Water / Sediment (Mission Creek) | Approximate Extent of Upper Watermill Catchment |
| Surface Water / Sediment (Watermill Creek) | Approximate Extent of Mission Creek Catchment |
| Tap | Alternative Water Supply |
| Stock Water (Beef Cattle) | Creek |
| Fruit and Vegetable Irrigation Water | Potential Sources |
| Pump | Group 1 Potential PFAS Primary Source Areas |
| Stock Water (Chickens) | Off-site |

| Details: | |
|-----------------------------------|------------------|
| Data Sources: | Created: T. Sohi |
| Aerial imagery: Esri maps | PM: C. Sandiford |
| Vector Datasets: Open Street Maps | Approved: x |
| | Date: 10/12/2021 |
| | Revision: 0 |

| | | | |
|--|-----|-------------|----------|
| 0 | 710 | 1,420 | 2,130 |
| Metres | | | |
| Datum/Projection: GDA 1994 MGA Zone 55 | | Scale: (A3) | 1:25,000 |

| | |
|-------------------|--|
| Figure No: | A1 |
| Title: | PFAS Management Areas |
| Project: | PFAS Management Plan |
| Location: | Norfolk Island International Airport Department for Infrastructure, Transport, Regional Development and Communications |
| Client: | |

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Maxar

Last Edited by: tara.sohi

Preliminary PFAS Investigation - Senversa Sample Nomenclature

| Sample Type | Sample Nomenclature | Detail |
|--|---|---|
| Soil – Surface | SS01, SS02 etc. | For surface soil samples collected within the top 10 cm of the surface. |
| Soil bores | SB01_0.01, SB01_0.05 etc. | For all target sample locations where soil bores are advanced using a hand auger. |
| Surface Water Samples | SW01, SW02 etc. | A two-letter identifier will be added to the beginning of the sample ID to identify catchment area, e.g. MC for Mission Creek (MC_SW01). |
| Sediment Samples | SD01, SD02 etc. | To be paired with surface water locations (i.e. MC_SD01 to be paired with MC_SW01) or a unique ID to be assigned in the event surface water is not sampled. |
| Water Supply Samples (Public) | PWS_01, PWS_02 etc. | Publicly accessible bores will contain an individual Bore ID starting with PWS (Public Water Supply). |
| Water Supply Samples (Private Property) | ID001_BORE_01, ID001_BORE_02, ID001_TAP_01 etc. | De-identified IDs to be assigned to each sampled property or location. Sample type to be identified i.e. bore/ tap/ tank etc. |
| Biota Samples | ID001_FRUIT_01 | De-identified IDs to be assigned to each sampled property as per water supply samples above. Biota type to be identified i.e. fruit/ egg/ grass etc. |
| Quality Samples | QC101, QC102 etc. | To be used for blind (intra-laboratory) duplicates. QA/QC register will be used during field works to record and track the quality samples collected. |
| | QC201, QC202 etc. | To be used for split (inter-laboratory) duplicates. QC sets will be paired (i.e. QC101 and QC201) at each location. |
| | QC301, QC302 etc. | To be used for rinsate blanks. |
| | QC401, QC402 etc. | To be used for trip and/or field blanks. |

Always include a “0” before single digit numbers; this is important for ESDAT data management.

Senversa Pty Ltd

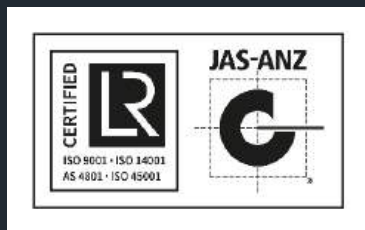
ABN 89 132 231 380

www.senversa.com.au

enquiries@senversa.com.au

LinkedIn: Senversa

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Appendix B: Data Validation



Appendix B: Quality Assurance / Quality Control

The data quality assurance and control (QA/QC) procedures adopted by Senversa provide a consistent approach to evaluation of whether the data quality objectives (DQO's) required by the project have been achieved. The process focuses on assessment of the useability of the data in terms of accuracy and reliability in forming conclusions on the condition of the element of the environment being investigated. The approach is generally based on guidance from the following sources:

- National Environment Protection Council (NEPC), National Environment Protection (Assessment of Site Contamination) Amendment Measure No. 1 2013 (NEPM), Schedule B2: Guideline on Site Characterisation.
- NEPC – National Environment Protection (Assessment of Site Contamination) Amendment Measure No. 1 2013 (NEPM), Schedule B3: Guideline on Laboratory Analysis of Potentially Contaminated Soils.
- Heads of Environmental Protection Authorities (HEPA), PFAS National Environmental Management Plan (PFAS NEMP) 3.0, 2025.
- United States Environmental Protection Agency (USEPA) – Guidance on Systematic Planning Using the Data Quality Objectives Process (EPA QA/G-4).
- USEPA – Guidance on Environmental Data Verification and Data Validation (EPA QA/G-8).

Quality Assurance Procedure

The following data quality objectives, measures and acceptance criteria were adopted to verify compliance with the planned QA procedures:

| Quality Assurance Process | Data Quality Element | Objectives and Measure | Acceptance Criteria |
|---|---|---|--|
| Standard Procedures | Comparability, Reproducibility, Representativeness. | Standard field sampling procedures and forms used. | No deviation from standard procedure and forms used. |
| Equipment Calibration | Accuracy. | All equipment calibrated in accordance with manufacturers specifications. | All equipment calibrated in accordance with manufacturers specifications. |
| Testing Method Accreditation | Accuracy and Comparability. | NATA accredited methods used for all analyses determined. | Primary and secondary laboratories to use NATA accredited methods for all analytes determined |
| Quality Control Sampling Frequency | Precision and Repeatability. | Field QC sampling frequency in accordance with AS4482.1-2005 and the PFAS NEMP 2.0. | Field Duplicates – ≥ 1 in 10 primary samples. Secondary Duplicates – ≥ 1 in 10 primary samples. Rinsate Blanks – ≥ 1 per day, per matrix per equipment. |



| Quality Assurance Process | Data Quality Element | Objectives and Measure | Acceptance Criteria |
|--|--|--|--|
| | Accuracy, Precision and Comparability. | Laboratory QC analysis frequency in accordance with NEPC (2013), Schedule B3. | Laboratory Duplicates – at least 1 in 10 analyses or one per process batch Method Blanks – at least 1 per process batch. Surrogate Recoveries – all samples spiked where appropriate (e.g. chromatographic analysis of organics). Laboratory Control Samples – at least 1 per process batch. Matrix Spikes – at least 1 per matrix type per process batch. |
| Sample Preservation, Handling and Holding Times | Accuracy. | Samples appropriately preserved upon collection, stored and transported, and analysed within holding times. | Sample containers, holding times and preservation in accordance laboratory specific method requirements. |
| Data Management | Accuracy. | No errors in data transcription. | Entry of field data verified by peer. |
| Data Useability | Completeness. | Limits of reporting less than adopted beneficial use investigation levels. Sample volumes and analytical methods selected to enable required limits of reporting to be achieved. | Limits of reporting less than investigation levels. |

Quality Control Sampling and Analysis

The following data quality objectives, measures and acceptance criteria were adopted to evaluate the validity of the analytical data produced.

| Quality Control Process | Data Quality Element | Objectives and Measure | Acceptance Criteria |
|--|------------------------------------|---|---|
| Field Duplicate Sampling and Analysis | Precision and Field Repeatability. | Field duplicate samples used to assess the variability in analyte concentration between samples collected from the sample location and the reproducibility of the laboratory analysis. Where required, resubmission of previously analysed samples for chemicals within their holding times may be undertaken to further assess level of precision. | Analysed for same chemicals as primary sample RPD $< 30\%$ of mean concentration where both concentrations $> 20 \times$ limit of reporting. RPD $< 50\%$ of mean concentration where higher concentration $10 - 20 \times$ limit of reporting. RPD - No limit where both concentrations $< 10 \times$ limit of reporting. |

¹ Relative Percent Difference (%): Calculated as: $(\text{Result No.1} - \text{Result No. 2}/\text{Mean Result}) \times 100$



| Quality Control Process | Data Quality Element | Objectives and Measure | Acceptance Criteria |
|---|------------------------------------|---|---|
| Secondary Duplicate Sampling and Analysis | Accuracy. | Results are accurate and free from laboratory error. Secondary duplicate samples sent to a secondary laboratory to assess the accuracy of the analyte concentrations reported by the primary laboratory. | <p>Analysed for same chemicals as primary sample.</p> <p>RPD <30% of mean concentration where both concentrations >20 x limit of reporting.</p> <p>RPD <50% of mean concentration where higher concentration 10 – 20 x limit of reporting</p> <p>RPD - No limit where both concentrations < 10 x limit of reporting.</p> |
| Field Rinsate Blank Preparation and Analysis | Accuracy and Representativeness. | <p>Cross contamination of samples does not occur between sampling locations due to carry-over from sampling equipment.</p> <p>Rinsate blank samples prepared for each sampling procedure. Where possible the rinsate blanks are prepared immediately after sampling locations known to contain concentrations of the chemicals of concern above the limit of quantification and / or before sampling locations where the chemicals being targeted in the laboratory analysis are to be compared to investigation levels near the limit of quantification of the chemical.</p> | Analyte concentrations below limits of reporting. |
| Trip Blank Sampling and Analysis | Accuracy and Representativeness. | <p>Cross contamination between samples does not occur in transit or as an artefact of the sample handling procedure.</p> <p>Trip blank samples prepared by the laboratory which accompany the empty sampling containers from the laboratory to the sampling site, and return with the samples to the laboratory to assess whether cross contamination occurs between samples or as an artefact of the sampling procedure.</p> | Analyte concentrations below limits of reporting. |
| Laboratory QC Analysis | Laboratory Precision and Accuracy. | <p>Laboratory duplicates.</p> <p>Laboratory control spike.</p> <p>Surrogate recovery.</p> <p>Matrix spike recovery</p> | <p>As specified by the laboratory.</p> <p>Dynamic recovery limits as specified by the laboratory.</p> <p>Dynamic recovery limits as specified by the laboratory.</p> <p>Recovery 70% – 130% or dynamic recovery limits specified by laboratory. However note that recovery of phenols is generally significantly lower and a recovery in the range 20% to 130% is considered acceptable by most laboratories.</p> |



Data Verification and Validation

The data validation process involved the checking of analytical procedure compliance with acceptance criteria and an assessment of the accuracy and precision of analytical data from the range of quality control indicators generated from both the sampling and analytical programmes.

The checks undertaken are summarised in the attached data validation checklist **Table B1**. Field replicate analytical results relevant to the project are summarised in **Table B2**.

Instances where the data quality acceptance criteria were not achieved are discussed in the table below:

| Item | Comment |
|--|---|
| Field and Secondary Duplicate Sampling and Analysis | <p><u>Water</u></p> <p>All water sample primary and duplicate sample pairs were within acceptable RPD limits, with the exception of:</p> <ul style="list-style-type: none"> Intra-laboratory sample QC104 (primary sample WC_OMP03) RPD outliers reported for PFOS (35%) and PFHxS (39%). Inter-laboratory sample QC202 (primary sample MC_OMP01) RPD outlier reported for PFOS (39%). <p>Primary and duplicate PFAS concentrations were within the same order of magnitude and the variations did not alter criteria exceedances. The non-conformity is considered likely due to low analyte concentrations. Therefore, non-compliance is not considered to impact on the quality of data.</p> <p><u>Biota</u></p> <p>No duplicate samples were collected for biota analysis due to the preliminary nature of this aspect of the investigation. Biota results should be considered to have a lower level of certainty.</p> |
| Laboratory Matrix Spike Analysis | <p><u>Batch ES2537583 soil and water analysis:</u></p> <ul style="list-style-type: none"> PFOS, PFHxS, PFDS MS recovery not determined, background level greater than or equal to 4x spike level. <p><u>Batch ES2537238 biota analysis:</u></p> <ul style="list-style-type: none"> PFPeA MS recovery not determined, background level greater than or equal to 4x spike level. MeFOSA recovery greater than upper data quality objective (123%). 10:2 FTS recovery less than lower data quality objective (91%). <p>MeFOSA and 10:2 FTS are not a critical CoPC and this low magnitude non-conformance is not considered to impact interpretation of the biota results.</p> |
| Laboratory Control Sample Analysis | <p><u>Batch ES2537238 biota analysis:</u></p> <ul style="list-style-type: none"> MeFOSA recovery greater than upper control limit (107 - 118%). <p>MeFOSA is not a critical CoPC and this minor non-conformance is not considered to impact interpretation of the biota results.</p> |
| Data Representativeness | <p><u>Sample Receipt Temperature</u></p> <p>Samples were received by the primary laboratory at 21.4-22.3°C, which is above the recommended temperature range ($\leq 6^\circ\text{C}$). Given the persistent nature of PFAS, the elevated temperature is not considered to have affected the quality of the data presented.</p> |

Data Suitability

While a small number of QC results were outside specified acceptance criteria, these were not considered to significantly impact on the quality or representativeness of the data, and the majority of results indicated that the precision and accuracy of the data were within acceptable limits. The results are therefore considered to be representative of chemical concentrations in the environmental media sampled at the time of sampling, and to be suitable to be used for their intended purpose in forming conclusions relating to the contamination status of water, soil, sediment and biota at the site.

| | | | | | |
|---|--|----------------------------------|-----------------------|---|---|
| Project Name: Ongoing Monitoring - Year 4 | | Project Number: C17776 | | | |
| Sampling Information | Sample Media: Soil, Sediment, Water, Biota | Sample Type | No. | Frequency | DQI Compliant? |
| | Date Sampled: 17-20/11/2025 | Primary: | 54 | - | |
| | Days of Sampling: 4 | Intra-laboratory duplicate (FD): | 6 | 1 per 9 primary samples | Yes |
| | Sampling Personnel: MA, HD | Inter-laboratory duplicate (FT): | 6 | 1 per 9 primary samples | Yes |
| | Primary Laboratory: ALS | Trip Blank (TB): | 0 | 1 per - day / batch | Yes |
| | Secondary Laboratory: Envirolab | Rinsate Blank (RB): | 1 | 1 per 4 day / batch | - |
| | No. Batches: 3 | Trip Spike (TS): | 0 | 1 per - day / batch | - |
| | Batch IDs: ES2537583, ES2537238, 396838 | Other: | NA | - | - |
| Precision | Intra-laboratory Duplicate (FD) analyses | | | | |
| | Analyte Group | Primary ID | Duplicate ID | DQI Compliant? | Comments |
| | PFAS Sediment | WC_OMP04_DUC | QC100 | Yes | |
| | PFAS Water | WWII_DAM | QC101 | Yes | |
| | PFAS Water | MC_OMP01 | QC102 | Yes | |
| | PFAS Water | MC_OMP06 | QC103 | Yes | |
| | PFAS Water | MC_OMP03 | QC104 | No | RPD exceedance for PFOS (35%) and PFHxS (39%). |
| | PFAS Water | MC_OMP02 | QC105 | Yes | |
| | Inter-laboratory Duplicate (FT) analyses | | | | |
| | Analyte Group | Primary ID | Duplicate ID | DQI Compliant? | Comments |
| | PFAS Sediment | WC_OMP04_DUC | QC200 | Yes | |
| | PFAS Water | WWII_DAM | QC201 | Yes | |
| | PFAS Water | MC_OMP01 | QC202 | No | RPD exceedance for PFOS (39%). |
| | PFAS Water | MC_OMP06 | QC203 | Yes | |
| | PFAS Water | MC_OMP03 | QC204 | Yes | |
| PFAS Water | MC_OMP02 | QC205 | Yes | | |
| Laboratory Duplicate (LD) analyses | | | | | |
| Batch No. | Matrix | Analyte Group | DQI Compliant? | Comments | |
| ES2537583 | Soil, water | PFAS | Yes | Frequencies met NEPM B3 and ALS QC standards. | |
| ES2537238 | Biota | PFAS | Yes | | |
| 396838 | Soil, water | PFAS | Yes | | |
| Accuracy | Laboratory Control Sample (LCS) analyses | | | | |
| | Batch No.(s) | Matrix | Analyte Group | DQI Compliant? | Comments |
| | ES2537583 | Soil, water | PFAS | Yes | |
| | ES2537238 | Biota | PFAS | No | MeFOSA recovery greater than upper control limit (107 - 118%). |
| | 396838 | Soil, water | PFAS | Yes | |
| | Surrogate Compound or Reference analyses | | | | |
| | Batch No.(s) | Matrix | Analyte Group | DQI Compliant? | Comments |
| | ES2537583 | Soil, water | PFAS | Yes | |
| | ES2537238 | Biota | PFAS | Yes | |
| | 396838 | Soil, water | PFAS | Yes | |
| | Spike Samples | | | | |
| | Type | Analyte Group | Batch No. | DQI Compliant? | Comments |
| | Laboratory matrix spike (MS) | PFAS | ES2537583 | No | PFOS, PFHxS, PFDS MS recovery not determined, background level greater than or equal to 4x spike level. |
| | Laboratory matrix spike (MS) | PFAS | ES2537238 | No | PFPeA MS recovery not determined, background level greater than or equal to 4x spike level. MeFOSA recovery greater than upper data quality objective (123%). 10:2 FTS recovery less than lower data quality objective (91%). |
| | Trip Spike (TS) | PFAS | | Yes | |
| Blank Samples | | | | | |
| Type | Analyte Group | DQI Compliant? | Comments | | |
| Laboratory Blank (LB): | PFAS | Yes | | | |
| Rinsate Blank (RB): | PFAS | Yes | | | |
| Field equipment calibration | | | | | |
| Equipment | Calibrated? | Record? | Equipment | Calibrated? | Record? |
| WQM | Yes | Yes | | | |
| Representativeness | DQI Compliant? | | | | |
| | Comments | | | | |
| | Appropriate & standard sampling methods used for media/CoPC? | Yes | | | |
| | Appropriate decontamination procedures carried out? | Yes | | | |
| | Samples collected in appropriate containers / preservatives? | Yes | | | |
| | Samples received at appropriate temperature / or with ice present? | No | | | |
| Samples extracted / analysed within holding times? | Yes | | | | |
| Samples analysed using appropriate NATA accredited methods? | Yes | | | | |
| Comparability | DQI Compliant? | | | | |
| | Comments | | | | |
| | Consistent sampling methods used? | Yes | | | |
| | Sampler(s) appropriately trained? | Yes | | | |
| Consistent site conditions and field scientist(s)? | Substantial | | | | |
| Consistent analytical methods used? | Yes | | | | |
| Completeness | DQI Compliant? | | | | |
| | Comments | | | | |
| | Field records / logs complete and retained? | Yes | | | |
| | Frequency of QC samples adequate per sampling plan? | Yes | | | |
| | Requested analyses completed per sampling plan? | Yes | | | |
| Appropriate PQLs? (relative to adopted criteria; available) | Yes | | | | |
| Chain-of-custody forms complete and correct? | Yes | | | | |
| QC check of data tables (against field records / laboratory reports)? | Yes | | | | |
| Data Use Suitability | | | | | |
| Yes / No | | | | | |
| Data from critical samples considered of suitable quality? | Yes | | | | |
| Data considered suitable for the objective of the assessment? | Yes | | | | |
| Overall Comments: While a small number of QC results were outside specified acceptance criteria, these were not considered to significantly impact on the quality or representativeness of the data, and majority of results indicated that the precision and accuracy of the data was within acceptable limits. The results are therefore considered to be representative of chemical concentrations in the environmental media sampled at the time of sampling, and to be suitable to be used for their intended purpose in forming conclusions relating to the contamination status of soil, sediment, water and biota at the site. | | | | | |
| Performed By: | MA | Checked By: | KR | | |
| Date: | 19/01/2026 | Date: | 20/01/2026 | | |

Table B2: Water RPD Results
Norfolk PFAS OMP



| | | Location Code | MC_OMP06 | MC_OMP06 | | MC_OMP06 | MC_OMP06 | | WWII_DAM | WWII_DAM | | WWII_DAM | WW11_DAM | |
|---|------|----------------|------------|------------|-----|------------|------------|-----|------------|------------|-----|------------|------------|-----|
| | | Field ID | MC_OMP06 | QC103 | | MC_OMP06 | QC203 | | WW11_DAM | QC101 | | WW11_DAM | QC201 | |
| | | Date | 20/11/2025 | 20/11/2025 | | 20/11/2025 | 20/11/2025 | | 20/11/2025 | 20/11/2025 | | 20/11/2025 | 20/11/2025 | |
| | | Sample Type | Normal | Field_D | | Normal | Interlab_D | | Normal | Field_D | | Normal | Interlab_D | |
| | | Lab Report No. | ES2537583 | ES2537583 | RPD | ES2537583 | 396838 | RPD | ES2537583 | ES2537583 | RPD | ES2537583 | 396838 | RPD |
| | Unit | EQL | | | | | | | | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| 6:2 Fluorotelomer Sulfonate (6:2 FIS) | µg/L | 0.01 | <0.05 | <0.05 | 0 | <0.05 | <0.01 | 0 | <0.05 | <0.05 | 0 | <0.05 | <0.01 | 0 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.02 | <0.05 | <0.05 | 0 | <0.05 | <0.02 | 0 | <0.05 | <0.05 | 0 | <0.05 | <0.02 | 0 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| Perfluoroalkane Carboxylic Acids | | | | | | | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.02 | 0.55 | 0.59 | 7 | 0.55 | - | - | 0.49 | 0.53 | 8 | 0.49 | - | - |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluorononanoic acid (PFNA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.02 | 0.16 | 0.17 | 6 | 0.16 | - | - | 0.12 | 0.14 | 15 | 0.12 | - | - |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.02 | 0.10 | 0.10 | 0 | 0.10 | - | - | 0.11 | 0.10 | 10 | 0.11 | - | - |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.1 | 0.1 | <0.1 | 0 | 0.1 | - | - | <0.1 | 0.1 | 0 | <0.1 | - | - |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.01 | 0.22 | 0.21 | 5 | 0.22 | 0.19 | 15 | 0.30 | 0.29 | 3 | 0.30 | 0.26 | 14 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.01 | 5.09 | 4.51 | 12 | 5.09 | 4.5 | 12 | 13.5 | 12.8 | 5 | 13.5 | 12 | 12 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.02 | 0.41 | 0.42 | 2 | 0.41 | - | - | 0.47 | 0.40 | 16 | 0.47 | - | - |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.01 | 3.37 | 4.04 | 18 | 3.37 | 3.3 | 2 | 3.87 | 4.20 | 8 | 3.87 | 3.9 | 1 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.02 | 0.18 | 0.22 | 20 | 0.18 | - | - | 0.27 | 0.28 | 4 | 0.27 | - | - |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.01 | 0.39 | 0.49 | 23 | 0.39 | 0.40 | 3 | 0.39 | 0.45 | 14 | 0.39 | 0.38 | 3 |
| Perfluoroalkyl Sulfonamides | | | | | | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEIFOSAA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |
| N-Ethyl perfluorooctane sulfonamide (EtFOA) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| N-Methyl perfluorooctane sulfonamide (MeFOA) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.05 | <0.05 | <0.05 | 0 | <0.05 | - | - | <0.05 | <0.05 | 0 | <0.05 | - | - |
| Perfluorooctane sulfonamide (FOA) | µg/L | 0.02 | <0.02 | <0.02 | 0 | <0.02 | - | - | <0.02 | <0.02 | 0 | <0.02 | - | - |

*RPDs have only been considered where a concentration is greater than 1 times the EQL.
 **Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each E)
 ***Interlab Duplicates are matched on a per compound basis as methods vary between labo

Table B3: Soil RPD Results
Norfolk PFAS OMP



| | | Location Code | WC_OMP04_DUCKDAM | WC_OMP04_DUCKDAM | | WC_OMP04_DUCKDAM | WC_OMP04_DUCKDAM | |
|---|-------|----------------|------------------|------------------|-----|------------------|------------------|-----|
| | | Field ID | WC_OMP04_SD | QC100 | | WC_OMP04_SD | QC200 | |
| | | Date | 19/11/2025 | 19/11/2025 | | 19/11/2025 | 19/11/2025 | |
| | | Sample Type | Normal | Field_D | | Normal | Interlab_D | |
| | | Lab Report No. | ES2537583 | ES2537583 | RPD | ES2537583 | 396838 | RPD |
| | Unit | EQL | | | | | | |
| (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| 6:2 Fluorotelomer Sulfonate (6:2 FIS) | mg/kg | 0.0001 | <0.0005 | <0.0005 | 0 | <0.0005 | <0.0002 | 0 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | mg/kg | 0.0002 | <0.0005 | <0.0005 | 0 | <0.0005 | <0.0004 | 0 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| Perfluoroalkane Carboxylic Acids | | | | | | | | |
| Perfluorohexanoic acid (PFHxA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorododecanoic acid (PFDoDA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorononanoic acid (PFNA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluoropentanoic acid (PFPeA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorotetradecanoic acid (PFTeDA) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| Perfluoroheptanoic acid (PFHpA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorobutanoic acid (PFBA) | mg/kg | 0.001 | <0.001 | <0.001 | 0 | <0.001 | - | - |
| Perfluorodecanoic acid (PFDA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorotridecanoic acid (PFTrDA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluoroundecanoic acid (PFUnDA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorooctanoic acid (PFOA) | mg/kg | 0.0001 | <0.0002 | <0.0002 | 0 | <0.0002 | <0.0002 | 0 |
| Perfluoroalkane Sulfonic Acids | | | | | | | | |
| Perfluorooctanesulfonic acid (PFOS) | mg/kg | 0.0001 | 0.0100 | 0.0083 | 19 | 0.0100 | 0.0086 | 15 |
| Perfluoropentane sulfonic acid (PFPeS) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorohexane sulfonic acid (PFHxS) | mg/kg | 0.0001 | 0.0008 | 0.0009 | 12 | 0.0008 | 0.0008 | 0 |
| Perfluoroheptane sulfonic acid (PFHpS) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorodecanesulfonic acid (PFDS) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| Perfluorobutane sulfonic acid (PFBS) | mg/kg | 0.0001 | <0.0002 | <0.0002 | 0 | <0.0002 | <0.0002 | 0 |
| Perfluoroalkyl Sulfonamides | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEIFOSAA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | mg/kg | 0.0005 | <0.0005 | <0.0005 | 0 | <0.0005 | - | - |
| Perfluorooctane sulfonamide (FOSA) | mg/kg | 0.0002 | <0.0002 | <0.0002 | 0 | <0.0002 | - | - |

*RPDs have only been considered where a concentration is greater than 1 times the EQL.

**Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 1000 (1 - 10 x EQL); 50 (10 - 20 x EQL); 30 (> 20 x EQL))

***Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

Table B4: Blanks Analytical Results
NORFOLK PFAS OMP



| | | Field ID | QC500 |
|---|------|----------------|------------|
| | | Date | 21/11/2025 |
| | | Sample Type | Rinsate |
| | | Lab Report No. | ES2537583 |
| | Unit | EQL | |
| (n:2) Fluorotelomer Sulfonic Acids | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | µg/L | 0.005 | <0.005 |
| 6:2 Fluorotelomer Sulfonate (6:2 FIS) | µg/L | 0.005 | <0.005 |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | µg/L | 0.005 | <0.005 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | µg/L | 0.005 | <0.005 |
| Perfluoroalkane Carboxylic Acids | | | |
| Perfluorohexanoic acid (PFHxA) | µg/L | 0.002 | <0.002 |
| Perfluorododecanoic acid (PFDoDA) | µg/L | 0.002 | <0.002 |
| Perfluorononanoic acid (PFNA) | µg/L | 0.002 | <0.002 |
| Perfluoropentanoic acid (PFPeA) | µg/L | 0.002 | <0.002 |
| Perfluorotetradecanoic acid (PFTeDA) | µg/L | 0.005 | <0.005 |
| Perfluoro-n-hexadecanoic acid (PFHxDA) | µg/L | 0.005 | <0.005 |
| Perfluoroheptanoic acid (PFHpA) | µg/L | 0.002 | <0.002 |
| Perfluorobutanoic acid (PFBA) | µg/L | 0.01 | <0.01 |
| Perfluorodecanoic acid (PFDA) | µg/L | 0.002 | <0.002 |
| Perfluorotridecanoic acid (PFTrDA) | µg/L | 0.002 | <0.002 |
| Perfluoroundecanoic acid (PFUnDA) | µg/L | 0.002 | <0.002 |
| Perfluorooctanoic acid (PFOA) | µg/L | 0.002 | <0.002 |
| Perfluoroalkane Sulfonic Acids | | | |
| Perfluorooctanesulfonic acid (PFOS) | µg/L | 0.002 | <0.002 |
| Perfluoropentane sulfonic acid (PFPeS) | µg/L | 0.002 | <0.002 |
| Perfluorohexane sulfonic acid (PFHxS) | µg/L | 0.002 | <0.002 |
| Perfluoroheptane sulfonic acid (PFHpS) | µg/L | 0.002 | <0.002 |
| Perfluorodecanesulfonic acid (PFDS) | µg/L | 0.002 | <0.002 |
| Perfluorobutane sulfonic acid (PFBS) | µg/L | 0.002 | <0.002 |
| Perfluoroalkyl Sulfonamides | | | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | µg/L | 0.005 | <0.005 |
| N-methylperfluorooctane sulfonamidoacetic acid (NMeFOSAA) | µg/L | 0.002 | <0.002 |
| N-ethyl-perfluorooctanesulfonamidoacetic acid (NEFOSAA) | µg/L | 0.002 | <0.002 |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | µg/L | 0.005 | <0.005 |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | µg/L | 0.005 | <0.005 |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | µg/L | 0.005 | <0.005 |
| Perfluorooctane sulfonamide (FOSA) | µg/L | 0.002 | <0.002 |



CERTIFICATE OF CALIBRATION

Air-Met Scientific Pty Ltd
 PO Box 133
 Nunawading VIC 3131
 Australia
 Phone: 1800 000 744

This document certifies that the instrument detailed has been calibrated to the parameters listed below

Certificate Number : 18L101818-R-8-20251114
Calibration Date: 14/11/2025
Next Calibration Due: 14/02/2026

Customer: Air-Met Scientific (National Rental)
Address: 65 Moray Street , , SOUTH MELBOURNE , 3205
Equipment No: EP020314
Unit Under Test: YSI ProPlus Water Quality Meter
Serial No: 18L101818
Service Order No: CAL09965

| Test Results | | | |
|--------------|---------------------------|------|----------|
| Item | Test | Pass | Comments |
| BATTERY | Capacity checked | Pass | |
| DISPLAY | Display/Backlight checked | Pass | |
| PCB | Inspected | Pass | |
| CASE | Inspected & Cleaned | Pass | |
| KEYPAD | Function checked | Pass | |
| SENSOR(S) | Cleaned | Pass | |

Reference Equipment:

| Equipment ID | Description | Expiry Date | Reference Certificate Number |
|--------------|-------------|-------------|------------------------------|
| MECOND | 452241 | | |
| MEPH7 | 441789 | | |
| MEPH4 | 443801 | | |
| MEORP | | | |
| MEDO | 445389 | | |

Results:

| Sensor | Units | Applied Value | Results | | Pass/ Fail | Comments |
|--------------|-------|---------------|----------|---------|------------|----------|
| | | | As Found | As Left | | |
| TEMPERATURE | °C | 17.8 | 17.8 | 17.8 | Pass | |
| CONDUCTIVITY | µS/cm | 2760 | 2677 | 2760 | Pass | |
| PH4 | pH | 4 | 4.33 | 4 | Pass | |
| PH7 | pH | 7 | 7.31 | 7 | Pass | |



| | | | | | | |
|------------------|----|--------|-------|--------|------|--|
| ORP | mV | 237.74 | 226.6 | 237.74 | Pass | |
| DISSOLVED OXYGEN | % | 100 | 114.6 | 100 | Pass | |

Note:

Calibrated By: REBECCA.MASSOUD

Signed: REBECCA.MASSOUD





Appendix C: Field Observations



Appendix D: Laboratory Certificates

CERTIFICATE OF ANALYSIS 396838

Client Details

| | |
|------------------|---------------------------------------|
| Client | Senversa Pty Ltd |
| Attention | Michelle Agnew |
| Address | 6/15 William St, Melbourne, VIC, 3000 |

Sample Details

| | |
|---|----------------------------------|
| Your Reference | <u>C17776 Norfolk DSI</u> |
| Number of Samples | 1 Sediment, 5 Water |
| Date samples received | 28/11/2025 |
| Date completed instructions received | 01/12/2025 |

Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client unless as indicated below in the method summaries. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

Please refer to the last page of this report for any comments relating to the results.

Report Details

Date results requested by 05/12/2025

Date of Issue 05/12/2025

NATA Accreditation Number 2901. This document shall not be reproduced except in full.

Accredited for compliance with ISO/IEC 17025 - Testing. **Tests not covered by NATA are denoted with ***

Results Approved By

Josh Williams, Senior R&D Chemist
 Phalak Inthakesone, Organics Development Manager, Sydney
 Timothy Toll, Senior Chemist

Authorised By

Nancy Zhang, Laboratory Manager

| PFAS in Soils Short | | |
|--|-------|------------|
| Our Reference | | 396838-1 |
| Your Reference | UNITS | QC200 |
| Date Sampled | | 19/11/2025 |
| Type of sample | | Sediment |
| Date prepared | - | 04/12/2025 |
| Date analysed | - | 05/12/2025 |
| Perfluorobutanesulfonic acid | µg/kg | <0.2 |
| Perfluorohexanesulfonic acid - PFHxS | µg/kg | 0.8 |
| Perfluorooctanesulfonic acid PFOS | µg/kg | 8.6 |
| Perfluorooctanoic acid PFOA | µg/kg | <0.2 |
| 6:2 FTS | µg/kg | <0.2 |
| 8:2 FTS | µg/kg | <0.4 |
| Surrogate ¹³ C ₈ PFOS | % | 93 |
| Surrogate ¹³ C ₂ PFOA | % | 100 |
| Extracted ISTD ¹³ C ₃ PFBS | % | 84 |
| Extracted ISTD ¹⁸ O ₂ PFHxS | % | 89 |
| Extracted ISTD ¹³ C ₄ PFOS | % | 89 |
| Extracted ISTD ¹³ C ₄ PFOA | % | 93 |
| Extracted ISTD ¹³ C ₂ 6:2FTS | % | 106 |
| Extracted ISTD ¹³ C ₂ 8:2FTS | % | 114 |
| Total Positive PFHxS & PFOS | µg/kg | 9.3 |
| Total Positive PFOS & PFOA | µg/kg | 8.6 |
| Total Positive PFAS | µg/kg | 9.3 |

| Moisture | | |
|----------------|-------|------------|
| Our Reference | | 396838-1 |
| Your Reference | UNITS | QC200 |
| Date Sampled | | 19/11/2025 |
| Type of sample | | Sediment |
| Date prepared | - | 02/12/2025 |
| Date analysed | - | 03/12/2025 |
| Moisture | % | 51 |

| PFAS in Waters Short | | | | | | |
|--|-------|------------|------------|------------|------------|------------|
| Our Reference | | 396838-2 | 396838-3 | 396838-4 | 396838-5 | 396838-6 |
| Your Reference | UNITS | QC201 | QC202 | QC203 | QC204 | QC205 |
| Date Sampled | | 20/11/2025 | 20/11/2025 | 20/11/2025 | 20/11/2025 | 20/11/2025 |
| Type of sample | | Water | Water | Water | Water | Water |
| Date prepared | - | 04/12/2025 | 04/12/2025 | 04/12/2025 | 04/12/2025 | 04/12/2025 |
| Date analysed | - | 05/12/2025 | 05/12/2025 | 05/12/2025 | 05/12/2025 | 05/12/2025 |
| Perfluorobutanesulfonic acid | µg/L | 0.38 | 0.65 | 0.40 | 0.06 | 0.04 |
| Perfluorohexanesulfonic acid - PFHxS | µg/L | 3.9 | 6.4 | 3.3 | 0.43 | 0.23 |
| Perfluorooctanesulfonic acid PFOS | µg/L | 12 | 11 | 4.5 | 0.33 | 0.05 |
| Perfluorooctanoic acid PFOA | µg/L | 0.26 | 0.41 | 0.19 | 0.02 | 0.01 |
| 6:2 FTS | µg/L | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| 8:2 FTS | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 |
| Surrogate ¹³ C ₈ PFOS | % | 96 | 97 | 94 | 94 | 94 |
| Surrogate ¹³ C ₂ PFOA | % | 92 | 98 | 94 | 98 | 93 |
| Extracted ISTD ¹⁸ O ₂ PFHxS | % | 107 | 103 | 101 | 102 | 105 |
| Extracted ISTD ¹³ C ₄ PFOS | % | 98 | 100 | 100 | 102 | 102 |
| Extracted ISTD ¹³ C ₄ PFOA | % | 109 | 109 | 106 | 107 | 108 |
| Extracted ISTD ¹³ C ₂ 6:2FTS | % | 121 | 117 | 124 | 115 | 117 |
| Extracted ISTD ¹³ C ₂ 8:2FTS | % | 120 | 127 | 122 | 124 | 124 |
| Total Positive PFHxS & PFOS | µg/L | 16 | 18 | 7.8 | 0.76 | 0.27 |
| Total Positive PFOA & PFOS | µg/L | 12 | 12 | 4.6 | 0.35 | 0.06 |
| Total Positive PFAS | µg/L | 16 | 19 | 8.4 | 0.84 | 0.33 |

| Method ID | Methodology Summary |
|--|---|
| <p>Inorg-008</p> <p>Org-029</p> | <p>Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.</p> <p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p> |

Client Reference: C17776 Norfolk DSI

| QUALITY CONTROL: PFAS in Soils Short | | | | | Duplicate | | | Spike Recovery % | | |
|--|-------|-----|---------|------------|-----------|------|------|------------------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-3 | [NT] |
| Date prepared | - | | | 04/12/2025 | [NT] | [NT] | [NT] | [NT] | 04/12/2025 | [NT] |
| Date analysed | - | | | 05/12/2025 | [NT] | [NT] | [NT] | [NT] | 05/12/2025 | [NT] |
| Perfluorobutanesulfonic acid | µg/kg | 0.1 | Org-029 | <0.1 | [NT] | [NT] | [NT] | [NT] | 106 | [NT] |
| Perfluorohexanesulfonic acid - PFHxS | µg/kg | 0.1 | Org-029 | <0.1 | [NT] | [NT] | [NT] | [NT] | 98 | [NT] |
| Perfluorooctanesulfonic acid PFOS | µg/kg | 0.1 | Org-029 | <0.1 | [NT] | [NT] | [NT] | [NT] | 100 | [NT] |
| Perfluorooctanoic acid PFOA | µg/kg | 0.1 | Org-029 | <0.1 | [NT] | [NT] | [NT] | [NT] | 99 | [NT] |
| 6:2 FTS | µg/kg | 0.1 | Org-029 | <0.1 | [NT] | [NT] | [NT] | [NT] | 102 | [NT] |
| 8:2 FTS | µg/kg | 0.2 | Org-029 | <0.2 | [NT] | [NT] | [NT] | [NT] | 118 | [NT] |
| Surrogate ¹³ C ₈ PFOS | % | | Org-029 | 95 | [NT] | [NT] | [NT] | [NT] | 98 | [NT] |
| Surrogate ¹³ C ₂ PFOA | % | | Org-029 | 98 | [NT] | [NT] | [NT] | [NT] | 97 | [NT] |
| Extracted ISTD ¹³ C ₃ PFBS | % | | Org-029 | 104 | [NT] | [NT] | [NT] | [NT] | 102 | [NT] |
| Extracted ISTD ¹⁸ O ₂ PFHxS | % | | Org-029 | 107 | [NT] | [NT] | [NT] | [NT] | 107 | [NT] |
| Extracted ISTD ¹³ C ₄ PFOS | % | | Org-029 | 108 | [NT] | [NT] | [NT] | [NT] | 107 | [NT] |
| Extracted ISTD ¹³ C ₄ PFOA | % | | Org-029 | 112 | [NT] | [NT] | [NT] | [NT] | 110 | [NT] |
| Extracted ISTD ¹³ C ₂ 6:2FTS | % | | Org-029 | 111 | [NT] | [NT] | [NT] | [NT] | 110 | [NT] |
| Extracted ISTD ¹³ C ₂ 8:2FTS | % | | Org-029 | 107 | [NT] | [NT] | [NT] | [NT] | 110 | [NT] |

Client Reference: C17776 Norfolk DSI

| QUALITY CONTROL: PFAS in Waters Short | | | | | Duplicate | | | Spike Recovery % | | |
|--|-------|------|---------|------------|-----------|------|------|------------------|------------|------|
| Test Description | Units | PQL | Method | Blank | # | Base | Dup. | RPD | LCS-W1 | [NT] |
| Date prepared | - | | | 04/12/2025 | [NT] | [NT] | [NT] | [NT] | 04/12/2025 | [NT] |
| Date analysed | - | | | 05/12/2025 | [NT] | [NT] | [NT] | [NT] | 05/12/2025 | [NT] |
| Perfluorobutanesulfonic acid | µg/L | 0.01 | Org-029 | <0.01 | [NT] | [NT] | [NT] | [NT] | 100 | [NT] |
| Perfluorohexanesulfonic acid - PFHxS | µg/L | 0.01 | Org-029 | <0.01 | [NT] | [NT] | [NT] | [NT] | 99 | [NT] |
| Perfluorooctanesulfonic acid PFOS | µg/L | 0.01 | Org-029 | <0.01 | [NT] | [NT] | [NT] | [NT] | 100 | [NT] |
| Perfluorooctanoic acid PFOA | µg/L | 0.01 | Org-029 | <0.01 | [NT] | [NT] | [NT] | [NT] | 106 | [NT] |
| 6:2 FTS | µg/L | 0.01 | Org-029 | <0.01 | [NT] | [NT] | [NT] | [NT] | 100 | [NT] |
| 8:2 FTS | µg/L | 0.02 | Org-029 | <0.02 | [NT] | [NT] | [NT] | [NT] | 105 | [NT] |
| Surrogate ¹³ C ₈ PFOS | % | | Org-029 | 93 | [NT] | [NT] | [NT] | [NT] | 103 | [NT] |
| Surrogate ¹³ C ₂ PFOA | % | | Org-029 | 96 | [NT] | [NT] | [NT] | [NT] | 99 | [NT] |
| Extracted ISTD ¹⁸ O ₂ PFHxS | % | | Org-029 | 108 | [NT] | [NT] | [NT] | [NT] | 108 | [NT] |
| Extracted ISTD ¹³ C ₄ PFOS | % | | Org-029 | 112 | [NT] | [NT] | [NT] | [NT] | 105 | [NT] |
| Extracted ISTD ¹³ C ₄ PFOA | % | | Org-029 | 116 | [NT] | [NT] | [NT] | [NT] | 111 | [NT] |
| Extracted ISTD ¹³ C ₂ 6:2FTS | % | | Org-029 | 129 | [NT] | [NT] | [NT] | [NT] | 132 | [NT] |
| Extracted ISTD ¹³ C ₂ 8:2FTS | % | | Org-029 | 129 | [NT] | [NT] | [NT] | [NT] | 136 | [NT] |

Result Definitions

| | |
|-------------|---|
| NT | Not tested |
| NA | Test not required |
| INS | Insufficient sample for this test |
| PQL | Practical Quantitation Limit |
| < | Less than |
| > | Greater than |
| RPD | Relative Percent Difference |
| LCS | Laboratory Control Sample |
| NS | Not specified |
| NEPM | National Environmental Protection Measure |
| NR | Not Reported |

Quality Control Definitions

| | |
|--|--|
| Blank | This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples. |
| Duplicate | This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable. |
| Matrix Spike | A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist. |
| LCS (Laboratory Control Sample) | This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample. |
| Surrogate Spike | Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples. |

Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Air volumes are typically provided by customers (often as flow rate(s) and sampling time(s) and/or simply volumes) sampled or exposure times (determines 'volume' passive badges are exposed to)). Hence in such circumstances the volume measurement is inevitably not covered by Envirolab's NATA accreditation. An exception may occur where Envirolab Newcastle does the sampling where accreditation exists for certain types of sampling and hence volume determination(s). Note air volumes are often used to determine concentrations for dust and/or analyses on filters, sorbents and in impingers. For canister sampling, the air volume is covered by Envirolab's NATA accreditation.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates can be downloaded from the [Envirolab Resources website](#) or obtained directly by contacting the laboratory.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

For Dust Deposit Gauge (DDG) analysis the sampling, sampling period and funnel exposure area do not fall under Envirolab's NATA accreditation (unless the Newcastle laboratory where responsible for the sampling), hence the annotation on the DDG units of reporting.

Urine Analysis - The BEI values listed are taken from the 2022 edition of "TLVs and BEIs Threshold Limits" by ACGIH.

Report Comments


PFAS Soil: PQLs raised due to high moisture content.

Chain of Custody Documentation

Senversa Pty Ltd
www.senversa.com.au
ABN 89 132 231 380

Laboratory: ALS NSW
Address: 277-289 Woodpark Road, Smithfield
Contact: Sample Receipt
Phone:

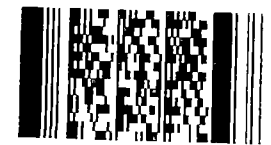
Job Number: C17776 Purchase Order: *senversa national quote*
Project Name: Norfolk DSI Quote No: *1*
Sampled By: *Michelle Agnew, Hollie Dunstan* Turn Around Time: *STD*
Project Manager: *Michelle Agnew* Page: *1 of 45*
Email Report To: *michelle.agnew@senversa.com.au* Phone/Mobile: *044 8910424*

| Sample Information | | Container Information | | Analysis Required | | | | | | | | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. | | | |
|--------------------|-------------------|-----------------------|----------|-------------------|-------------|---------------|---|---|---|---|---|---|---|---|---|
| Lab ID | Sample ID | Matrix* | Date | Time | Type / Code | Total Bottles | | | | | | | | | |
| 1 | WU11-DAM | WATER | 20/11/25 | | | | X | X | X | X | X | X | X | X |  HOLD |
| 2 | MC_OMP01 | | | | | | X | X | X | X | X | X | X | X | |
| 3 | MC_OMP02 | | | | | | X | X | X | X | X | X | X | X | |
| 4 | MC_OMP03 | | | | | | X | X | X | X | X | X | X | X | |
| 5 | MC_OMP04a | | | | | | X | X | X | X | X | X | X | X | |
| 6 | MC_OMP05a | | | | | | X | X | X | X | X | X | X | X | |
| 7 | MC_OMP06 | | | | | | X | X | X | X | X | X | X | X | |
| 8 | MC_OMP07a | | | | | | X | X | X | X | X | X | X | X | |
| 9 | MC_OMP08 | | | | | | X | X | X | X | X | X | X | X | |
| 9 | MC_OMP09 | | 17/11/25 | | | | X | X | X | X | X | X | X | X | |
| 10 | MC_OMP10 | | 19/11/25 | | | | X | X | X | X | X | X | X | X | |
| 11 | MC_OMP11 | | 17/11/25 | | | | X | X | X | X | X | X | X | X | |
| 12 | WC_OMP01 | | " | | | | X | X | X | X | X | X | X | X | |
| 13 | WC_OMP02 | | " | | | | X | X | X | X | X | X | X | X | |
| 14 | WC_OMP03 | | 18/11/25 | | | | X | X | X | X | X | X | X | X | |
| 15 | WC_OMP04 DUCK DAM | | " | | | | X | X | X | X | X | X | X | X | |
| 16 | WC_OMP05 | | " | | | | X | X | X | X | X | X | X | X | |
| 17 | COCKPIT-SWC1 | | 18/11/25 | | | | X | X | X | X | X | X | X | X | |
| 18 | FRE-TAP1 | | 19/11/25 | | | | X | X | X | X | X | X | X | X | |
| 19 | FRE-TAP2 | | 19/11/25 | | | | X | X | X | X | X | X | X | X | |
| Total | 2019 | | | | | | | | | | | | | | |

CHIROLAB
Chatswood NSW 2067
Ph: (02) 9978 8200

Job No: *396830*
Date Received: *21/11/25*
Time Received: *17:10*
Received By: *[Signature]*
Temp: *6*
Cooling: *icepack*
Security: *handwritten*

Environmental Division
Sydney
Work Order Reference
ES2537583



Telephone : + 61-2-8784 8555

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples:
Sampler Name: *Michelle Agnew* Signature: *[Signature]* Date: *21/11/25*

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|-----------------|-------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic;
V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic;
F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____

Senversa Pty Ltd
www.senversa.com.au
ABN 89 132 231 380

Laboratory: ALS NSW
Address: 277-289 Woodpark Road, Smithfield
Contact: Sample Receipt
Phone:

Analysis Required

Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc.

Job Number: C17776 Purchase Order:
Project Name: Norfolk DSI Quote No:
Sampled By: Turn Around Time:
Project Manager: Page: 2 of 5
Email Report To: michelle.agnew@senversa.com.au Phone/Mobile:

Table with columns: Lab ID, Sample ID, Matrix, Date, Time, Type / Code, Total Bottles. Rows include DEPOT_TAPI, DEPOT_TANK1, DEPOT_TANK2, DEPOT_TANK3, A_TAPI, AIRPORT_BORE, PWS_HEAD_DAM, PWS_HEAD_TOILET, PWS_BUMBORA_TOILET, IDO13_BORE, IDO14_BORE, IDO15_BORE, IDO16_BORE, PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP5, PWS_HOSP_TAP6, PWS_HOSP_TAP7, PWS_HOSP_TAP8, A_TAPI2.

Table for Analysis Required with columns for various analytes. Includes handwritten notes: 'PEAS standard', 'PEAS', 'Email Analysis Request', 'HOLD', and '396F38 28/11/25 AB'.

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: Sampler Name: michelle agnew Signature: [Signature] Date: 21/11/25

Table for Relinquished By, Method of Shipment, and Received by. Includes fields for Name/Signature, Date, and Time.

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO3) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic; V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: [Signature] Checked by: [Signature]

Chain of Custody Documentation

Laboratory: ALS NSW
Address: 277-289 Woodpark Road, Smithfield
Contact: Sample Receipt
Phone:

Job Number: C17776
Purchase Order:
Project Name: Norfolk DSI
Quote No:
Sampled By:
Turn Around Time:
Project Manager:
Page: 3 of 5
Email Report To: michelle.agnew@senversa.com.au
Phone/Mobile:

| Sample Information | | | Container Information | | Analysis Required | | | | | | | | | | | | | | | |
|--------------------|------------------------|----------|-----------------------|------|-------------------|---------------|--|--|--|--|--|--|--|--|--|--|--|--|--|--|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | | | | | | | | | | | | | | |
| ✓ 40 | A-TAP316A | Water | 17/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 41 | A-TAP5 | | | | | | | | | | | | | | | | | | | |
| ✓ 42 | A-TAP4 | | 19/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 43 | A-TAP5 | | " | | | | | | | | | | | | | | | | | |
| ✓ 44 | WCO1 | | 18/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 45 | WCO2 | | 18/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 46 | WC_OMP04_SD | SED. | 19/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 47 | FRE-TAP6 | Water | " | | | | | | | | | | | | | | | | | |
| ✓ 48 | AN-ELEC-TAP1 | | " | | | | | | | | | | | | | | | | | |
| ✓ 49 | WASTE-TAP1 | | 20/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 50 | COUNCIL-TAP1 | | | | | | | | | | | | | | | | | | | |
| ✓ 51 | ID008-SS1 | SOIL | | | | | | | | | | | | | | | | | | |
| ✓ 52 | ID008-SS2 | | | | | | | | | | | | | | | | | | | |
| ✓ 53 | ID008-DI00A | | | | | | | | | | | | | | | | | | | |
| ✓ 54 | WASTE-TAP1 | | | | | | | | | | | | | | | | | | | |
| ✓ 55 | WASTE-TAP1 | | | | | | | | | | | | | | | | | | | |
| ✓ 56 | A-TRUCKFILL-SS1 | | | | | | | | | | | | | | | | | | | |
| ✓ 57 | BALL-SS3 | SOIL | 21/11/25 | | | | | | | | | | | | | | | | | |
| ✓ 58 | BALL-SS4 | | " | | | | | | | | | | | | | | | | | |
| ✓ 59 | BALL-SS5 | | " | | | | | | | | | | | | | | | | | |
| ✓ 59 | BALL-SD1 | sed | " | | | | | | | | | | | | | | | | | |
| Total | | 15+ | | | | | | | | | | | | | | | | | | |

TAPS standard
TAPS
Email Analysis Request
HOLD
Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc.

396838
28/11/25
MB

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples:
Sampler Name: michelle Agnew
Signature: [Signature]
Date: 21/11/25

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|-----------------|--------------|--|
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: | Date: | |
| Of: | Time: | Date/Time: | Of: | Time: | |
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: | Date: | |
| Of: | Time: | Date/Time: | Of: | Time: | |
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: | Date: | |
| Of: | Time: | Date/Time: | Of: | Time: | |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic;
V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic;
F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____

Senversa Pty Ltd
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ABN 89 132 231 380

Laboratory:
Address:
Contact:
Phone:

Job Number: C17776
Project Name: Norfolk DSI
Sampled By:
Project Manager:
Email Report To: michelle.agnew@senversa.com.au
Purchase Order:
Quote No:
Turn Around Time:
Page: 4 of 5
Phone/Mobile:

| Sample Information | | | | Container Information | | Analysis Required | | | | | | | | | | | | | | | |
|--------------------|------------------|----------|----------|-----------------------|-------------|-------------------|--|--|--|--|--|--|--|--|--|--|--|--|--|--|--|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | | | | | | | | | | | | | | | |
| 60 | QC100 | SED | 19/11/25 | | | | | | | | | | | | | | | | | | |
| 61 | QC101 | WATER | 20/11/25 | | | | | | | | | | | | | | | | | | |
| 62 | QC102 | | | | | | | | | | | | | | | | | | | | |
| 63 | QC103 | | | | | | | | | | | | | | | | | | | | |
| 64 | QC104 | | | | | | | | | | | | | | | | | | | | |
| 65 | QC105 | | | | | | | | | | | | | | | | | | | | |
| | QC106 | | | | | | | | | | | | | | | | | | | | |
| | QC200 | SED | 19/11/25 | | | | | | | | | | | | | | | | | | |
| | QC201 | WATER | 20/11/25 | | | | | | | | | | | | | | | | | | |
| | QC202 | | | | | | | | | | | | | | | | | | | | |
| | QC203 | | | | | | | | | | | | | | | | | | | | |
| | QC204 | | | | | | | | | | | | | | | | | | | | |
| | QC205 | | | | | | | | | | | | | | | | | | | | |
| | QC206 | | | | | | | | | | | | | | | | | | | | |
| 66 | QC500 | WATER | 20/11/25 | | | | | | | | | | | | | | | | | | |
| 67 | QC300 | " | 20/11/25 | | | | | | | | | | | | | | | | | | |
| | WATER | | | | | | | | | | | | | | | | | | | | |
| 68 | BALL SWI | Water | 21/11/25 | | | | | | | | | | | | | | | | | | |
| Total 14 | | | | | | | | | | | | | | | | | | | | | |

09-03-21

Analysis Required: PFAS Standard, PFAS, Email Analysis Request, SEND TO ENVIRONMENTAL HOLD

Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc.

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples:
Sampler Name: michelle Agnew
Signature: [Signature]
Date: 21/11/25

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|-----------------|-------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic; V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____



Senversa Pty Ltd
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ABN 89 132 231 380

Chain of Custody Documentation

Laboratory: **ALS NSW**
Address:
Contact:
Phone:

| Analysis Required | | | | | | | | | | | |
|-------------------|--|--|--|--|--|--|--|--|--|------|---|
| | | | | | | | | | | HOLD | Comments: e.g. Highly contaminated sample; hazardous materials present; trace LORs etc. |
| | | | | | | | | | | | |
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Job Number: C17776 Purchase Order:
 Project Name: Norfolk DSI Quote No: **Senversa National**
 Sampled By: **MA, TD** Turn Around Time: **std**
 Project Manager: **Michelle Agnew** Page: **5 of 5**
 Email Report To: **michelle.agnew@senversa.com.au** Phone/Mobile: **0448910424**

| Sample Information | | | | Container Information | | |
|--------------------|-----------------|----------|----------|-----------------------|-------------|---------------|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles |
| 69 | 0WC-OMP04-SD | SOIL | 19/11/25 | | | |
| 70 | ID008-SS1 | | 20/11/25 | | | |
| 71 | ID008-SS1 | | 20/11/25 | | | |
| 72 | QC100 | | 19/11/25 | | | |
| 73 | QC200 | | " | | | |
| 74 | A-TRUCKFILL-SS1 | | 20/11/25 | | | |
| 75 | A-TRUCKFILL-SS2 | | " | | | |
| 76 | BALL-SS4 | | 21/11/25 | | | |
| 77 | BALL-SS5 | | " | | | |
| 78 | BALL-SS3 | ↓ | " | | | |
| 79 | BALL-SD1 | | " | | | |
| Total | | | | | | |

396838
28/11/25
MA

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: Sampler Name: **Michelle Agnew** Signature: **[Signature]** Date: **21/11/25**

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|---------------------------|-----------------------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: MA | Date: 24/11/25 |
| Of: | Time: | Date/Time: | | Of: MA | Time: 08:45 |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic; V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____

Jack Clifton

From: Andrew Wotherspoon
Sent: Tuesday, 25 November 2025 9:23 AM
To: Jack Clifton
Cc: Samples Sydney
Subject: RE: [EXTERNAL] - Samples sent from Norfolk Island
Attachments: Import Permit - Soil and Water Samples 2024-09-24_10-58-45-AM (1).pdf

Apologies Jack, following on from this email, Michelle has mentioned the following for samples being sent to Envirolab using the attached permit -

- Attached is the Envirolab import permit. Please send the following samples to Envirolab:
- QC200, QC201, QC202, QC203, QC304, QC205

From my understanding, Envirolab are the same biosecurity as us so hoping there's no issues with this but let me know and I can certainly assist.

Kind Regards,



Andrew Wotherspoon
Client Services Officer, Environmental
Sydney, NSW

O: +61 2 8784 8555

D: +61 2 8784 8514

andrew.wotherspoon@alsglobal.com

277-289 Woodpark Road

Smithfield NSW 2164 AUSTRALIA

alsglobal.com



[EnviroMail 93 Australia](#) [Re-release - Laboratory quality assurance/quality control](#)

[EnviroMail 148 Australia](#) [- Interpreting TOP Assay](#)

[EnviroMail 149 Australia](#) [- Net Zero commitments and roadmap](#)

[EnviroMail 150 Australia](#) [- Spotlight on emerging contaminants - Triclosan](#)

[EnviroMail 151 Australia](#) [- Expanding the scope of PFAS analysis in soils and waters](#)

396838

28/11/25

AB



396838

28/11/25

CUSTOMS DECLARATION

SENDER'S NAME: Senversa Pty Ltd _____

ADDRESS: Level 6, 15 William Street, Melbourne VIC 3000

RECEIVER'S NAME: AUSTRALIAN LABORATORY SERVICES

ADDRESS: ALS Environmental
277-289 Woodpark Road
SMITHFIELD NSW 2164

RECEIVER'S CONTACT NAME: Scott James

RECEIVER'S CONTACT PHONE NO.: +61-2-8784-8555

FULL DESCRIPTION OF GOODS: Environmental Samples for Analysis

PURPOSE FOR SENDING: Analytical Testing (Environmental)

VALUE FOR CUSTOMS PURPOSES ONLY: \$39 AUD

* NUMBER OF PACKAGES: 2 _____

* TOTAL WEIGHT: _____

* CONSIGNMENT NOTE NUMBER: 081-5514-5506 . _____

Away Bill

COURIER COMPANY: Burnt Pine Travel _____

BIOSECURITY ENTRY NUMBER: _____

I declare the above information to be true and correct to the best of my knowledge.

Signed: Michelle Agnew

Dated: 21/11/2025

CUSTOMS DECLARATION

SENDER'S NAME: Senversa Pty Ltd

ADDRESS: Level 6, 15 William Street, Melbourne VIC 3000

RECEIVER'S NAME: AUSTRALIAN LABORATORY SERVICES

ADDRESS: ALS Environmental
277-289 Woodpark Road
SMITHFIELD NSW 2164

RECEIVER'S CONTACT NAME: Scott James

RECEIVER'S CONTACT PHONE NO.: 61-2-8784-8555

FULL DESCRIPTION OF GOODS: Environmental Samples for Analysis
Water: approx. 80 20ml plastic jars.
Soil: 11 plastic jars

PURPOSE FOR SENDING: Analytical Testing (Environmental)

VALUE FOR CUSTOMS PURPOSES ONLY: \$39 AUD

* NUMBER OF PACKAGES: 2

* TOTAL WEIGHT: _____

* CONSIGNMENT NOTE NUMBER: 081-5514-5506

Always Bill

COURIER COMPANY: Burnt Pine Travel

BIOSECURITY ENTRY NUMBER: _____

I declare the above information to be true and correct to the best of my knowledge.

Signed: Michelle Agnew

Dated: 21/11/2025

SUPPLIER'S DECLARATION:
Subject to Permit 0009235853, section 1.1 condition f), I declare the goods meet the following conditions:
1. have not been sourced or sampled from waste collection or waste treatment facilities (human and/or animal), intensive animal production facilities, farm sites and/or aquaculture facilities.
2. are free from visible seeds, human, animal and plant debris and other non-commodity biosecurity risk material.

signed: michelle agnew. michelle Agnew 21/11/2025.



Permit to import conditionally non-prohibited goods
 This permit is issued under *Biosecurity Act 2015* Section 179 (1)

Permit: 0009235853

**Valid for: multiple consignments
 between 23 August 2024 and 23 August 2028**

This permit is issued to: **Australian Laboratory Services Pty Ltd.**
 277/289 Woodpark Road
 Smithfield NSW 2164
 AUSTRALIA

Attention: Mr Scott James

This permit is issued for the import of Biological products (Non-standard goods).

| | |
|--------------------|-------------------|
| Exporter details: | Various exporters |
| Country of export: | Various countries |

This permit includes the following good(s). Refer to the indicated page for details of the permit conditions:

| | | | |
|---------------------------------------|--|---|---------|
| 1. Soil and water samples | Country of origin: Various countries | Permit Conditions: Environmental samples for laboratory analysis, culture and isolation not permitted | Page 4 |
| 2. Animal fluids and tissues | Country of export: Various countries Country of origin: Various countries | | |
| 3. Laboratory material | Country of origin: Various countries | Permit Conditions: Laboratory materials for in vitro use only | Page 11 |
| 4. Microorganisms (including viruses) | Country of export: Various countries Country of origin: Various countries | Permit Conditions: Standard laboratory microorganisms and infectious agents (and derivatives) | Page 14 |
| 5. Soil and water samples | | | |

This permit is granted subject to the requirement that fees determined under section 592(1) are paid.

James Teale
 Subdelegate of the Director of Biosecurity

Date: 23 August 2024



Permit to import conditionally non-prohibited goods

This permit is issued under *Biosecurity Act 2015* Section 179 (1)

Permit: 0009431147

**Valid for: multiple consignments
 between 17 October 2024 and 17 October 2029**

This permit is issued to: Envirolab Services Pty Ltd
 Ground Floor Organic Laboratory Area
 12 Ashley Street
 Chatswood NSW 2067
 AUSTRALIA

Attention: Mr Matthew Mansfield

This permit is issued for the import of Biological products (Standard goods).

| | |
|-------------------|-------------------|
| Exporter details: | Various exporters |
|-------------------|-------------------|

This permit includes the following good(s). Refer to the indicated page for details of the permit conditions:

| | |
|---------------------------|--|
| 1. Soil and water samples | |
| End use: | In-vitro |
| Country of export: | Various countries |
| Country of origin: | Various countries |
| Permit Conditions: | Environmental samples for laboratory analysis, culture and isolation not permitted |
| | Page 3 |

NOTE: Where a good has more than one set of permit conditions please read each set to determine which set of permit conditions applies to a specific consignment.

----- End of commodity list -----

This permit is granted subject to the requirement that fees determined under section 592(1) are paid.

| | |
|--|-------------------------|
| Sarah Jeffress Subdelegate of the Director of Biosecurity | Date: 24 September 2024 |
|--|-------------------------|

Important information about this permit and the import of goods

Note: This permit covers Department of Agriculture, Fisheries and Forestry import conditions. It is the permit holder's responsibility to ensure all legal requirements relating to the goods described in this permit are met. While the permit holder should rely on their own inquiries, the following information is provided to assist the permit holder in meeting legal obligations in relation to the importation of the goods described in this permit.

Information about this permit

Authority to import

The permit holder is authorised to import the goods described in this permit subject to the listed conditions specified in this permit.

Compliance with permit conditions and assessment and management of biosecurity risk

All imports are subject to biosecurity control and may be subject to biosecurity inspection on arrival to determine compliance with the listed permit conditions and to assess the level of biosecurity risk associated with the goods. Imports that do not comply with the import conditions specified in the permit may present an unacceptable level of biosecurity risk and may be subject to biosecurity measures that may include treatment, export or destruction at the permit holder's expense or forfeited to the Commonwealth.

Additionally, non-compliance with import permit conditions may constitute an offence or contravention of a civil penalty provision under section 187 of the *Biosecurity Act 2015*.

Change of import conditions

The Director of Biosecurity may, in accordance with section 180 of the *Biosecurity Act 2015* vary or revoke the conditions on a permit or impose further conditions.

General information about importing goods

Notification of import

Notification of the import must be provided to the Department of Agriculture, Fisheries and Forestry for all imported goods other than goods imported as accompanied baggage or goods imported via the mail and not prescribed under *the Customs Act 1901*, or where other exceptions specified in the *Biosecurity Regulation 2016* apply. Notification must be provided in accordance with section 120 of the *Biosecurity Act 2015* and Part 1 of Chapter 2 of the *Biosecurity Regulation 2016*. Please refer to '[Sending your goods to Australia](#)' on the Department of Agriculture, Fisheries and Forestry website.

Provision of required documentation

It is recommended that all required documentation accompanies each consignment. Required documentation must be presented to the Department of Agriculture, Fisheries and Forestry for assessment. Airfreight or mail shipments should have all required documentation securely attached to the outside of the package, and clearly marked "Attention Department of Agriculture, Fisheries and Forestry". Documentation may include the permit (or permit number), government certification and invoice.

If the product description on the permit varies from the identifying documentation provided, the goods will not be released from biosecurity control unless evidence is provided to the biosecurity officer that the permit covers the goods in the consignment.

Any documentation provided must comply with the Department of Agriculture, Fisheries and Forestry's [minimum documentation requirements policy](#).

Non-commodity cargo clearance

In addition to the conditions for the goods being imported, non-commodity biosecurity risks are assessed including container cleanliness, packaging and destination concerns, and may be subject to inspection and treatment on arrival. Please refer to the [Non-Commodity Cargo Clearance BICON case](#) for further information.

Fees

Fees are payable to the Department of Agriculture, Fisheries and Forestry for certain services (see the *Biosecurity Charges Imposition (General) Regulation 2016*, Part 2 of Chapter 9 of the *Biosecurity Regulation 2016* and Part 3 of Chapter 11 of the *Biosecurity Act 2015*). Detail on how the department applies fees and levies may be found in the [Charging guidelines](#).

Compliance with other regulatory provisions

Goods imported into Australia may be subject to regulatory requirements under other legislation. It is the permit holder's responsibility to identify and ensure they have complied with all requirements of any other regulatory agency or advisory body prior to and after importation.

Permit conditions

It is the importer's responsibility to ensure that the following permit conditions are met in relation to each consignment. Where more than one set of permit conditions is shown for a good please read each set of conditions to determine which applies to a specific consignment.

1. Environmental samples for laboratory analysis, culture and isolation not permitted

This section contains permit conditions for the following commodity (or commodities):

- | |
|---------------------------|
| 1. Soil and water samples |
|---------------------------|

1.1. Biosecurity Pathway

Import conditions prior to arrival in Australian territory

- a. The goods must be:
1. Soil samples (including, but not limited to, subsoil, aquatic or marine soil, sediments, silt)
 2. Water samples
 3. Dust samples
 4. Mining slurry samples
 5. Inorganic material contaminated with soil and/or water.



The goods may also be imported as cores, swabs, and/or filters used on the listed sample types where all conditions can be met.

- b. The samples must not be sourced from waste collection or waste treatment facilities (human and/or animal), intensive animal production facilities, farm sites, horticultural sites and/or aquaculture facilities.
- c. The imported goods must be free from visible seed, human, animal and plant debris and other biosecurity risk material.
- d. Liquid or water samples must be imported in a volume less than or equal to 1 L or 1 kg per individually packaged unit.
- e. Solid samples must be imported in units less than or equal to 10 L or 10 kg per individually packaged unit.
- f. The goods must meet the following import conditions.
To demonstrate compliance with this requirement you must present the following on a Manufacturer's declaration or Supplier's declaration:
A statement that the goods:
1. have not been sourced or sampled from waste collection or waste treatment facilities (human and/or animal), intensive animal production facilities, farm sites and/or aquaculture facilities
 2. are free from visible seed, human, animal and plant debris and other non-commodity biosecurity risk material.

Import conditions after arrival in Australian territory

- g. The goods are for use at approved arrangement sites class 5. The level of containment must be BC level 1 or higher.
- h. The goods must only be directed to AA sites with current approval from the Department of Agriculture, Fisheries and Forestry as a class 5 approved arrangement site at the time of importation and until such time that all imported material and its derivatives are removed for disposal or export.
- i. The goods and their derivatives shall not be removed from these sites, except for treatment, disposal or export, without the prior approval of the Director of Biosecurity.
- j. **Post entry/end use conditions**
1. The goods must not be exposed to or used in animals, plants, microorganisms, cell cultures or the environment, and must not be used in or on humans.
 2. The goods must not be used for culture or isolation of microorganisms and infectious agents, or for the synthesis of replication-competent microorganisms or infectious agents.
 3. Microorganisms or infectious agents must not be cultured or isolated from the materials imported under the permit conditions in this permit case.
 4. The goods must be labelled with the end use conditions on the smallest individually packaged unit.
- OR
- The smallest individually packaged unit must be accompanied by documentation stating the end use conditions. This documentation must be provided to the end user of the goods.



Where applicable, the importer or end user must comply with:

1. International (e.g. International Air Transport Association) and domestic requirements concerning the safe handling, transport and labelling of biological material
2. AS/NZS 2243 Safety in Laboratories standards
3. Office of the Gene Technology Regulator (OGTR) requirements
4. The Security Sensitive Biological Agents (SSBA) regulatory scheme.



Records of treatment, disposal and release of all imported items must be retained by the AA site for Department of Agriculture, Fisheries and Forestry audit purposes.

- k. The goods may be released from biosecurity containment for further in vitro analysis if the samples are subjected to any of the sample preparation or analysis techniques listed below:
1. One or a combination of the following:
 - 1.1. Solid phase extraction (SPE), liquid-liquid extraction (LLE) or solid-liquid extraction (SLE) using organic solvents such as hexane, isooctane, carbon tetrachloride, chloroform, tetrahydrofuran, dichloromethane (methylene dichloride), acetone, isopropanol, methanol, ethyl acetate, and acetonitrile (methyl cyanide, cyanomethane).
 - 1.2. Atomic absorption spectrometry
 - 1.3. The use of a flame atomiser
 - 1.4. Induction coupled plasma analysis
 - 1.5. Gas chromatography
 - 1.6. Liquid chromatography

- 1.7. Mass spectrometry
 - 1.8. Optical emission spectrometry
 - 1.9. Thermal, electron or chemical ionisation
 - 1.10. Thermoluminescence dating in which the sample material is progressively heated from 110°C to at least 350°C (excludes low temperature thermoluminescence)
 - 1.11. High temperature combustion (>600°C)
 2. Sample preparation which includes autoclaving at a minimum of 121°C and 15psi for 30 minutes.
 3. Complete acid digestion using one or a combination of the following: concentrated hydrochloric (HCl 32-37%/~12M), nitric (HNO₃ 65-70%/~16M), perchloric (HClO₄ ~70%/11M), sulphuric (H₂SO₄ 95- 98%/~18M) and hydrofluoric (HF 40-48%/~27M) acid in:
 - 3.1. A microwave digestion system at $\geq 150^{\circ}\text{C}$ and $\geq 15\text{psi}$ for at least 20 minutes; or
 - 3.2. A heating block at a minimum of 100°C for at least 30 minutes.
 4. Acidification at less than or equal to pH 2.0 throughout the final sample for at least 30 minutes.
 5. Alkalisiation at greater than or equal to pH 10 throughout the final sample for at least 30 minutes.
- l. Genetic material extracted from imported samples may be released from biosecurity containment provided all of the following conditions are met:
1. The genetic material must be extracted using a standard laboratory procedure that lyses cells, and degrades lipids, proteins and other molecules, and results in a purified DNA and/or RNA product that is unable to replicate.
 2. Individual sample sizes must be 2 mL or 2 g or less and in sealed containers.
 3. Genetic material removed from AA site containment must be used for *in vitro* analytical procedures only (e.g. PCR, sequencing, Northern/Southern blotting).
 4. Genetic material must not be used for the synthesis of replication-competent microorganisms, infectious agents, or homologues.
 5. Genetic material and derivatives must not be directly or indirectly exposed to animals.
 6. Any sample remnants, sample containers and disposable equipment that have contacted the samples must be subjected to a Department of Agriculture, Fisheries and Forestry approved treatment method prior to disposal OR the genetic material, its containers and disposable equipment that has contacted the genetic material must be treated as “potentially contaminated wastes” as described in section 12 of AS/NZS 2243.3 Safety in Laboratories Part 3: Microbiological safety and containment.
- m. The goods may be treated using one of the following treatment methods. After treatment, the goods may then be released from biosecurity control.
- Soil samples (and other non-liquid goods)
1. dry heat treatment at 160°C for 2 hours (sample must not exceed 500g in weight) (applied in the current AA or AA class 12.3 or 4.1), or
 2. heat treatment in an autoclave at 121°C, 103kPa (15 psi) for 30 minutes (applied in the current AA or AA class 8.3), or
 3. heat treatment in an autoclave at 134°C, 103kPa (15 psi) for 4 minutes (applied in the current AA or AA class 8.3), or
 4. ionising radiation to a level that achieves a minimum absorbed dose of 50kGy (applied in AA class 4.2).
- Water samples (and other liquid goods)

1. heat treatment in an autoclave at 121°C, 103kPa (15 psi) for 30 minutes (applied in current AA or AA class 8.3), or
 2. heat treatment in an autoclave at 134°C, 103kPa (15 psi) for 4 minutes (applied in current AA or AA class 8.3), or
 3. heat treatment at a minimum core temperature of 100°C for at least 30 minutes (applied in the current AA or AA class 12.3 or 4.1), or
 4. ionising radiation to a level that achieves a minimum absorbed dose of 50kGy (applied in AA class 4.2).
 5. addition of sodium hypochlorite to water to a final chlorine concentration of 2,500 ppm, stirring the contents and allowing a standing time of 2 hours before disposal into the sewerage system.
 6. addition of calcium hypochlorite to water to a final chlorine concentration of 2,500 ppm, stirring the contents and allowing a standing time of 2 hours before disposal into the sewerage system.
- n. On completion of work all imported materials and the direct or indirect derivatives thereof must be disposed of by treatment methods (as listed) or other methods approved in writing by the Director of Biosecurity.

Additional information

- o. **Commercial administrative conditions**
Documents must be provided with each consignment which:
1. identify the consignment (if non-personal) e.g. entry number
 2. identify all goods being imported as part of this consignment e.g. invoice or waybill or importer's manifest
 3. describe the goods being imported (where not clear).
e.g. 1: Product XRab = Purified protein derived from rabbits
e.g. 2: Product AX = Synthetic antibiotic
e.g. 3: Comte = Cheese.
- p. Under the Biosecurity Charges Imposition (General) Regulation 2016 and Chapter 9, Part 2 of the Biosecurity Regulation 2016, fees are payable to the Department of Agriculture, Fisheries and Forestry for all services. Detail on how the department applies fees and levies may be found in the Charging guidelines.
- q. In addition to the conditions for the goods being imported, non-commodity concerns must be assessed including container cleanliness, packaging and destination concerns, and may be subject to inspection and treatment on arrival. Please refer to the Non-Commodity Cargo Clearance BICON case for further information.

----- **End of permit conditions** -----



DATA QUALITY ASSESSMENT SUMMARY

Report Details

| | |
|-----------------------------------|--------------------|
| Envirolab Report Reference | 396838 |
| Client ID | Senversa Pty Ltd |
| Project Reference | C17776 Norfolk DSI |
| Date Issued | 05/12/2025 |

QC DATA

All laboratory QC data was within the Envirolab Group's specifications.

HOLDING TIME COMPLIANCE EVALUATION

All preservation / holding times (based on AS/ASPHA/ISO/NEPM/USEPA reference documents and standards) are compliant.

Certain analyses have had their recommended technical holding times elongated by filtering and/or freezing on receipt at the laboratory (e.g. BOD, chlorophyll/Pheophytin, nutrients and acid sulphate soil tests).

COMPLIANCE TO QC FREQUENCY (NEPM)

Internal laboratory QC rate complies with NEPM requirements (LCS/MB/MS 1 in 20, Duplicates 1 in 10 samples). Note, samples are batched together with other sample consignments in order to assign QC sample frequency.

QC Evaluation

| | |
|---|---|
| Duplicate(s) was performed as per NEPM frequency | ✓ |
| Laboratory Control Sample(s) were analysed with the samples received | ✓ |
| A Method Blank was performed with the samples received | ✓ |
| Matrix spike(s) was performed as per NEPM frequency (Not Applicable for Air samples) | ✓ |

Refer to Certificate of Analysis for all Quality Control data.

SAMPLE RECEIPT ADVICE

Client Details

| | |
|------------------|------------------|
| Client | Senversa Pty Ltd |
| Attention | Michelle Agnew |

Sample Login Details

| | |
|---|--------------------|
| Your reference | C17776 Norfolk DSI |
| Envirolab Reference | 396838 |
| Date Sample Received | 28/11/2025 |
| Date Instructions Received | 01/12/2025 |
| Date Results Expected to be Reported | 05/12/2025 |

Sample Condition

| | |
|---|---------------------|
| Samples received in appropriate condition for analysis | Yes |
| No. of Samples Provided | 1 Sediment, 5 Water |
| Turnaround Time Requested | Standard |
| Temperature on Receipt (°C) | 6 |
| Cooling Method | Ice |
| Sampling Date Provided | YES |

Comments

Nil

Please direct any queries to:

Aileen Hie

Phone: 02 9910 6200
Fax: 02 9910 6201
Email: ahie@envirolab.com.au

Jacinta Hurst

Phone: 02 9910 6200
Fax: 02 9910 6201
Email: jhurst@envirolab.com.au

Analysis Underway, details on the following page:



| Sample ID | PFAS in Soils Short | PFAS in Waters Short |
|-----------|---------------------|----------------------|
| | QC200 | ✓ |
| QC201 | | ✓ |
| QC202 | | ✓ |
| QC203 | | ✓ |
| QC204 | | ✓ |
| QC205 | | ✓ |

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.

We acknowledge receipt of your samples and Purchase Order (PO) (if provided). If a PO includes your terms & conditions, Envirolab hereby expressly rejects and will not be bound by any external or third-party terms and conditions, including those referenced or attached to a PO.

All services to be performed by Envirolab will be governed exclusively by Envirolab's General Terms and Conditions attached to this acknowledgement ([Envirolab Terms](#)) via hyperlink or found on our websites.

If you do not object in writing within two (2) business days of the date of this acknowledgement, you will be deemed to have accepted the Envirolab Terms. In addition, your provision of further instructions, additional samples, payment of any invoice, or acceptance of services or results from Envirolab will constitute acceptance of the Envirolab Terms. For clarity, Envirolab's commencement or continuation of work following receipt of the PO is performed solely under the Envirolab Terms and does not constitute acceptance of any external terms. All rights are expressly reserved.



QUALITY CONTROL REPORT

Work Order : **ES2537238**

Page : 1 of 9

Amendment : **1**

Client : **SENVERSA PTY LTD**

Laboratory : Environmental Division Sydney

Contact : MICHELLE AGNEW

Contact : Andrew Wotherspoon

Address : Level 24, 1 Market St,
SYDNEY NSW 2000

Address : 277-289 Woodpark Road Smithfield NSW Australia 2164

Telephone : ----

Telephone : +61-2-8784 8555

Project : C17776 Norfolk DSI

Date Samples Received : 24-Nov-2025

Order number : ----

Date Analysis Commenced : 04-Dec-2025

C-O-C number : ----

Issue Date : 24-Dec-2025

Sampler : Michelle Agnew Hollie Dunsten

Site : ----

Quote number : EN/000

No. of samples received : 6

No. of samples analysed : 5



Accreditation No. 825
Accredited for compliance with
ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories

Position

Accreditation Category

Alex Rossi

Organic Chemist

Sydney Organics, Smithfield, NSW



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key : Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

= Indicates failed QC

* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

Sub-Matrix: BIOTA

| | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|--------------|--|------------|-----------------------------------|-------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7054345) | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | 1 | 3 | 59.0 | No Limit |
| | | EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7099301) | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | 1 | 1 | 0.0 | No Limit |
| | | EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7054345) | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |



Sub-Matrix: **BIOTA**

Laboratory Duplicate (DUP) Report

| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
|--|--------------|---|------------|--------|-------|-----------------|------------------|---------|--------------------|
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7054345) - continued | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | <5 | 0.0 | No Limit | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7099301) | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 1 (2)* | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | <5 | 0.0 | No Limit |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7054345) | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 1 (5)* | µg/kg | <5 | <5 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 2 (5)* | µg/kg | <5 | <5 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7099301) | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 1 (5)* | µg/kg | <5 | <5 | 0.0 | No Limit |



Sub-Matrix: BIOTA

| | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|--------------|---|-------------|-----------------------------------|-------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7099301) - continued | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | <1 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 2 (5)* | µg/kg | <5 | <5 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7054345) | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7099301) | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | <2 | 0.0 | No Limit |
| EP231P: PFAS Sums (QC Lot: 7054345) | | | | | | | | | |
| ES2537238-001 | WC_BIOTA1 | EP231X: Sum of PFAS | ---- | 1 | µg/kg | 1 | 3 | 100 | No Limit |
| EP231P: PFAS Sums (QC Lot: 7099301) | | | | | | | | | |
| ES2537238-004 | ID008_BIOTA2 | EP231X: Sum of PFAS | ---- | 1 | µg/kg | 1 | 1 | 0.0 | No Limit |



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

Sub-Matrix: BIOTA

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | |
|---|------------|-----|-------|-----------------------------|---------------------------------------|---------------------------|------------------------------|------|
| | | | | Result | Spike Concentration | Spike Recovery (%) LCS | Acceptable Limits (%) Low | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7054345) | | | | | | | | |
| EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | 5 µg/kg | 76.9 | 70.0 | 130 |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | 5 µg/kg | 89.4 | 72.0 | 128 |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | 5 µg/kg | 95.6 | 73.0 | 123 |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | 5 µg/kg | 95.6 | 67.0 | 130 |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | 5 µg/kg | 82.2 | 70.0 | 132 |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | <1 | 5 µg/kg | 94.3 | 68.0 | 136 |
| EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | 5 µg/kg | 97.1 | 70.0 | 130 |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 1 | µg/kg | <1 | 5 µg/kg | 109 | 59.0 | 134 |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7099301) | | | | | | | | |
| EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | 5 µg/kg | 93.3 | 70.0 | 130 |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | 5 µg/kg | 83.8 | 72.0 | 128 |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | 5 µg/kg | 100 | 73.0 | 123 |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | 5 µg/kg | 88.0 | 67.0 | 130 |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | 5 µg/kg | 103 | 70.0 | 132 |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | <1 | 5 µg/kg | 108 | 68.0 | 136 |
| EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | 5 µg/kg | 106 | 70.0 | 130 |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 1 | µg/kg | <1 | 5 µg/kg | 113 | 59.0 | 134 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7054345) | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | 25 µg/kg | 118 | 71.0 | 135 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 1 | µg/kg | <1 | 5 µg/kg | 121 | 69.0 | 132 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | <1 | 5 µg/kg | 100 | 70.0 | 132 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | <1 | 5 µg/kg | 104 | 71.0 | 131 |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | <1 | 5 µg/kg | 106 | 69.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | 5 µg/kg | 104 | 72.0 | 129 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | 5 µg/kg | 102 | 69.0 | 133 |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | 5 µg/kg | 97.2 | 64.0 | 136 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 1 | µg/kg | <1 | 5 µg/kg | 88.4 | 69.0 | 135 |
| EP231X: Perfluorotridecanoic acid (PFTTrDA) | 72629-94-8 | 1 | µg/kg | <1 | 5 µg/kg | 99.8 | 66.0 | 139 |
| EP231X: Perfluorotetradecanoic acid (PFTTeDA) | 376-06-7 | 2 | µg/kg | <2 | 12.5 µg/kg | 105 | 69.0 | 133 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7099301) | | | | | | | | |



Sub-Matrix: BIOTA

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | |
|---|------------|-----|-------|--------------------------|---------------------------------------|--------------------|-----------------------|-----|
| | | | | Result | Spike Concentration | Spike Recovery (%) | Acceptable Limits (%) | |
| | | | | | LCS | Low | High | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7099301) - continued | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | 25 µg/kg | 125 | 71.0 | 135 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 1 | µg/kg | <1 | 5 µg/kg | 111 | 69.0 | 132 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | <1 | 5 µg/kg | 102 | 70.0 | 132 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | <1 | 5 µg/kg | 103 | 71.0 | 131 |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | <1 | 5 µg/kg | 104 | 69.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | 5 µg/kg | 97.8 | 72.0 | 129 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | 5 µg/kg | 105 | 69.0 | 133 |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | 5 µg/kg | 107 | 64.0 | 136 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 1 | µg/kg | <1 | 5 µg/kg | 105 | 69.0 | 135 |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 1 | µg/kg | <1 | 5 µg/kg | 86.8 | 66.0 | 139 |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 2 | µg/kg | <2 | 12.5 µg/kg | 107 | 69.0 | 133 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7054345) | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 1 | µg/kg | <1 | 5 µg/kg | 106 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 2 | µg/kg | <2 | 12.5 µg/kg | # 107 | 88.1 | 105 |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | 12.5 µg/kg | 114 | 81.6 | 144 |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | 12.5 µg/kg | 113 | 84.7 | 135 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | 12.5 µg/kg | 117 | 20.5 | 150 |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | 5 µg/kg | 102 | 63.0 | 144 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | 5 µg/kg | 103 | 61.0 | 139 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7099301) | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 1 | µg/kg | <1 | 5 µg/kg | 104 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 2 | µg/kg | <2 | 12.5 µg/kg | # 118 | 88.1 | 105 |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | 12.5 µg/kg | 114 | 81.6 | 144 |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | 12.5 µg/kg | 117 | 84.7 | 135 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | 12.5 µg/kg | 117 | 20.5 | 150 |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | 5 µg/kg | 108 | 63.0 | 144 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | 5 µg/kg | 105 | 61.0 | 139 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7054345) | | | | | | | | |



Sub-Matrix: BIOTA

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | | |
|--|-------------|-----|-------|--------------------------|---------------------------------------|--------------------|-----------------------|------|--|
| | | | | Result | Spike | Spike Recovery (%) | Acceptable Limits (%) | | |
| | | | | | Concentration | LCS | Low | High | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7054345) - continued | | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | 5 µg/kg | 117 | 62.0 | 145 | |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | 5 µg/kg | 110 | 64.0 | 140 | |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | 5 µg/kg | 115 | 65.0 | 137 | |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | 5 µg/kg | 97.4 | 93.4 | 130 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7099301) | | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | 5 µg/kg | 101 | 62.0 | 145 | |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | 5 µg/kg | 98.0 | 64.0 | 140 | |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | 5 µg/kg | 115 | 65.0 | 137 | |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | 5 µg/kg | 104 | 93.4 | 130 | |
| EP231P: PFAS Sums (QCLot: 7054345) | | | | | | | | | |
| EP231X: Sum of PFAS | ---- | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums (QCLot: 7099301) | | | | | | | | | |
| EP231X: Sum of PFAS | ---- | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- | |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

Sub-Matrix: BIOTA

| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Matrix Spike (MS) Report | | | |
|---|--------------|--|------------|--------------------------|--------------------|-----------------------|------|
| | | | | Spike | Spike Recovery (%) | Acceptable Limits (%) | |
| | | | | Concentration | MS | Low | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7054345) | | | | | | | |
| ES2537238-003 | ID008_BIOTA1 | EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 5 µg/kg | 77.8 | 70.0 | 130 |
| | | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 5 µg/kg | 97.6 | 72.0 | 128 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 5 µg/kg | 122 | 73.0 | 123 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 5 µg/kg | 104 | 67.0 | 130 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 5 µg/kg | 102 | 70.0 | 132 |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 5 µg/kg | 112 | 68.0 | 136 |
| | | EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 5 µg/kg | 109 | 70.0 | 130 |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 5 µg/kg | 103 | 59.0 | 134 |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7099301) | | | | | | | |
| ES2537238-006 | ID008_BIOTA4 | EP231X: Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 5 µg/kg | 84.4 | 70.0 | 130 |
| | | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 5 µg/kg | 78.8 | 72.0 | 128 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 5 µg/kg | 103 | 73.0 | 123 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 5 µg/kg | 82.4 | 67.0 | 130 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 5 µg/kg | 113 | 70.0 | 132 |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 5 µg/kg | 99.5 | 68.0 | 136 |



Sub-Matrix: BIOTA

| | | | | Matrix Spike (MS) Report | | | |
|---|--------------|---|------------|--------------------------|------------------|-----------------------|------|
| | | | | Spike | SpikeRecovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7099301) - continued | | | | | | | |
| ES2537238-006 | ID008_BIOTA4 | EP231X: Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 5 µg/kg | 107 | 70.0 | 130 |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 5 µg/kg | 107 | 59.0 | 134 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7054345) | | | | | | | |
| ES2537238-003 | ID008_BIOTA1 | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 25 µg/kg | 116 | 71.0 | 135 |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 5 µg/kg | 100 | 69.0 | 132 |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 5 µg/kg | 105 | 70.0 | 132 |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 5 µg/kg | 109 | 71.0 | 131 |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 5 µg/kg | 112 | 69.0 | 133 |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 5 µg/kg | 112 | 72.0 | 129 |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 5 µg/kg | 119 | 69.0 | 133 |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 5 µg/kg | 115 | 64.0 | 136 |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 5 µg/kg | 102 | 69.0 | 135 |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 5 µg/kg | 116 | 66.0 | 139 |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 12.5 µg/kg | 112 | 69.0 | 133 | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7099301) | | | | | | | |
| ES2537238-006 | ID008_BIOTA4 | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 25 µg/kg | 130 | 71.0 | 135 |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 5 µg/kg | # Not Determined | 69.0 | 132 |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 5 µg/kg | 111 | 70.0 | 132 |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 5 µg/kg | 95.2 | 71.0 | 131 |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 5 µg/kg | 110 | 69.0 | 133 |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 5 µg/kg | 96.0 | 72.0 | 129 |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 5 µg/kg | 111 | 69.0 | 133 |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 5 µg/kg | 105 | 64.0 | 136 |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 5 µg/kg | 102 | 69.0 | 135 |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 5 µg/kg | 76.0 | 66.0 | 139 |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 12.5 µg/kg | 101 | 69.0 | 133 |
| | | EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7054345) | | | | | |
| ES2537238-003 | ID008_BIOTA1 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 5 µg/kg | 121 | 67.0 | 137 |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 12.5 µg/kg | 95.2 | 88.1 | 105 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 12.5 µg/kg | 113 | 81.6 | 144 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 12.5 µg/kg | 103 | 84.7 | 135 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 12.5 µg/kg | 94.0 | 20.5 | 150 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 5 µg/kg | 109 | 63.0 | 144 |



Sub-Matrix: BIOTA

| | | | | Matrix Spike (MS) Report | | | |
|---|--------------|---|-------------|--------------------------|------------------|-----------------------|------|
| | | | | Spike | SpikeRecovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7054345) - continued | | | | | | | |
| ES2537238-003 | ID008_BIOTA1 | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 5 µg/kg | 109 | 61.0 | 139 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7099301) | | | | | | | |
| ES2537238-006 | ID008_BIOTA4 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 5 µg/kg | 113 | 67.0 | 137 |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 12.5 µg/kg | # 123 | 88.1 | 105 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 12.5 µg/kg | 106 | 81.6 | 144 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 12.5 µg/kg | 120 | 84.7 | 135 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 12.5 µg/kg | 106 | 20.5 | 150 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 5 µg/kg | 106 | 63.0 | 144 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 5 µg/kg | 92.5 | 61.0 | 139 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7054345) | | | | | | | |
| ES2537238-003 | ID008_BIOTA1 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 5 µg/kg | 121 | 62.0 | 145 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 5 µg/kg | 118 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 5 µg/kg | 112 | 65.0 | 137 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 5 µg/kg | # 91.3 | 93.4 | 130 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7099301) | | | | | | | |
| ES2537238-006 | ID008_BIOTA4 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 5 µg/kg | 107 | 62.0 | 145 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 5 µg/kg | 104 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 5 µg/kg | 118 | 65.0 | 137 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 5 µg/kg | 94.1 | 93.4 | 130 |



QA/QC Compliance Assessment to assist with Quality Review

| | | | |
|--------------|---------------------------------|-------------------------|---------------------------------|
| Work Order | : ES2537238 | Page | : 1 of 5 |
| Amendment | : 1 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 |
| Site | : ---- | Issue Date | : 24-Dec-2025 |
| Sampler | : Michelle Agnew Hollie Dunsten | No. of samples received | : 6 |
| Order number | : ---- | No. of samples analysed | : 5 |

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers : Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- **NO** Method Blank value outliers occur.
- **NO** Duplicate outliers occur.
- Laboratory Control outliers exist - please see following pages for full details.
- Matrix Spike outliers exist - please see following pages for full details.
- For all regular sample matrices, where applicable to the methodology, **NO** surrogate recovery outliers occur.

Outliers : Analysis Holding Time Compliance

- **NO** Analysis Holding Time Outliers exist.

Outliers : Frequency of Quality Control Samples

- **NO** Quality Control Sample Frequency Outliers exist.



Outliers : Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: BIOTA

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|--|----------------------|------------------|---|-------------|----------------|-----------|---|
| Laboratory Control Spike (LCS) Recoveries | | | | | | | |
| EP231C: Perfluoroalkyl Sulfonamides | QC-7054345-002 | ---- | N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 107 % | 88.1-105% | Recovery greater than upper control limit |
| EP231C: Perfluoroalkyl Sulfonamides | QC-7099301-002 | ---- | N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 118 % | 88.1-105% | Recovery greater than upper control limit |
| Matrix Spike (MS) Recoveries | | | | | | | |
| EP231B: Perfluoroalkyl Carboxylic Acids | ES2537238--006 | ID008_BIOTA4 | Perfluoropentanoic acid (PFPeA) | 2706-90-3 | Not Determined | ---- | MS recovery not determined, background level greater than or equal to 4x spike level. |
| EP231C: Perfluoroalkyl Sulfonamides | ES2537238--006 | ID008_BIOTA4 | N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 123 % | 88.1-105% | Recovery greater than upper data quality objective |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | ES2537238--003 | ID008_BIOTA1 | 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 91.3 % | 93.4-130% | Recovery less than lower data quality objective |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for VOC in soils vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: BIOTA

Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|---|--------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| Biota Sample Pre-Preparation | | | | | | | |
| HDPE Soil Jar (Biota-PP) WC_BIOTA1 | 19-Nov-2025 | 04-Dec-2025 | ---- | ---- | ---- | ---- | ---- |
| HDPE Soil Jar (Biota-PP) ID008_BIOTA1, | ID008_BIOTA3 | 20-Nov-2025 | 04-Dec-2025 | ---- | ---- | ---- | ---- |
| HDPE Soil Jar (Biota-PP) ID008_BIOTA2, | ID008_BIOTA4 | 20-Nov-2025 | 22-Dec-2025 | ---- | ---- | ---- | ---- |



Matrix: **BIOTA** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|---|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | |
| HDPE Soil Jar (EP231-PFOS-SP) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231-PFOS-SP) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 10-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231-PFOS-SP) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 23-Dec-2025 | 01-Feb-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 24-Dec-2025 | 01-Feb-2026 | ✓ |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | |
| HDPE Soil Jar (EP231X) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 24-Dec-2025 | 01-Feb-2026 | ✓ |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | |
| HDPE Soil Jar (EP231X) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 24-Dec-2025 | 01-Feb-2026 | ✓ |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | |
| HDPE Soil Jar (EP231X) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 24-Dec-2025 | 01-Feb-2026 | ✓ |
| EP231P: PFAS Sums | | | | | | | |
| HDPE Soil Jar (EP231X) WC_BIOTA1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA1, ID008_BIOTA3 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 11-Dec-2025 | 14-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_BIOTA2, ID008_BIOTA4 | 20-Nov-2025 | 23-Dec-2025 | 19-May-2026 | ✓ | 24-Dec-2025 | 01-Feb-2026 | ✓ |



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: **BIOTA** Evaluation: ✖ = Quality Control frequency not within specification ; ✔ = Quality Control frequency within specification.

| Quality Control Sample Type | Method | Count | | Rate (%) | | | Quality Control Specification |
|--|--------|-------|---------|----------|----------|------------|--------------------------------|
| | | QC | Reaular | Actual | Expected | Evaluation | |
| Analytical Methods | | | | | | | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 5 | 40.00 | 10.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 5 | 40.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 5 | 40.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 5 | 40.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|--|---------------|--------|--|
| PFOS - Linear/Branched Speciation | EP231-PFOS-SP | BIOTA | In-house: Linear PFOS is determined by quantitation of the separate linear peak using linear PFOS. Branched PFOS is determined as the difference between total PFOS (determined using a mixed linear/branched standard) and linear PFOS. |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | BIOTA | In-house: A sample extract is analysed by LC-Electrospray-MS-MS, Negative Mode using MRM using internal standard quantitation. Isotopically labelled analogues of target analytes used as internal standards and surrogates are added to a portion of biota which is then solvent extracted. A portion of extract is exchanged into the analytical solvent mixture, combined with an equal volume reagent water and filtered for analysis. Method procedures and data quality objectives conform to US DoD QSM 5.3, table B-15 requirements. |
| Preparation Methods | Method | Matrix | Method Descriptions |
| Prep-Preparation for Biota Analysis | * Biota-PP | BIOTA | A sample is prepared from whole or particular tissues/organs, identified, homogenised and the total weight of prepared sample recorded. |
| Sample Preparation for PFAS in Biota | EP231-PR | BIOTA | In-house: Isotopically labelled analogues of target analytes used as internal standards and surrogates are added to a portion of homogenised biota which is then extracted with MTBE and an ion pairing reagent. A portion of extract is exchanged into the analytical solvent mixture, combined with an equal volume reagent water and filtered for analysis. Method procedures conform to US DoD QSM 5.3, table B-15 requirements. |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537238-AA | Page | : 1 of 6 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 16:15 |
| Order number | : ---- | Date Analysis Commenced | : 04-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 11:59 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- Amendment (22/12/2025): This report has been amended following the request to add additional analysis EP231X PFAS Full Suite to samples 004 and 006 as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to organise spill reports as requested by Michelle Agnew.
- EP231X (Biota): ALS NATA accreditation for PFAS in Biota covers all Perfluoroalkyl Sulfonic Acids, Perfluoroalkyl Carboxylic Acids and (n:2) Fluorotelomer Sulfonic Acids in fish (whole and muscle), plants and vegetable matrices, with the exception PFBA (fish only), EtFOSA, MeFOSE, EtFOSE, MeFOSAA, EtFOSAA.
- EP231: Stable isotope enriched internal standards are added to samples prior to extraction. Target compounds have a direct analogous internal standard with the exception of PFPeS, PFHpA, PFDS, PFTrDA and 10:2 FTS. These compounds use an internal standard that is chemically related and has a retention time close to that of the target compound. The DQO for internal standard response is 50-150% of that established at initial calibration. PFOS is quantified using a certified, traceable standard consisting of linear and branched PFOS isomers. These practices are in line with recommendations in the National Environmental Management Plan for PFAS (Australian HEPA) and also conform to QSM 5.3 (US DoD) requirements.



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | Sample ID | WC_BIOTA1 | ---- | ---- | ---- | ---- |
|---|------------|-----|-------------------|---------------|-------|-------|-------|-------|
| Sampling date / time | | | 19-Nov-2025 00:00 | ---- | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537238-001 | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- |
| Biota Sample Pre-Preparation | | | | | | | | |
| ∅ Sample Description | ---- | - | -- | plant | ---- | ---- | ---- | ---- |
| ∅ Weight of Sample Prepared | ---- | 0.1 | g | 2.9 | ---- | ---- | ---- | ---- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | |
| Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) - Linear | 2795-39-3 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) - Branched | ---- | 1 | µg/kg | 1 | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | 1 | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- |
| Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | | Sample ID | WC_BIOTA1 | ---- | ---- | ---- | ---- |
|--|-------------|-----|-------|-------------------|-----------|-------|-------|-------|-------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | ---- | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537238-001 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- | --- |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 5 | µg/kg | <5 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 5 | µg/kg | <5 | ---- | ---- | ---- | ---- | ---- |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- | ---- |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | ---- | ---- | ---- | ---- | ---- |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | ---- | ---- | ---- | ---- | ---- |
| EP231P: PFAS Sums | | | | | | | | | |
| ^ Sum of PFAS | ---- | 1 | µg/kg | 1 | ---- | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | | Sample ID | WC_BIOTA1 | ---- | ---- | ---- | ---- |
|--------------------------------------|--------------------|-----|-------|-------------------|-----------|-------|-------|-------|------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537238-001 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231P: PFAS Sums - Continued | | | | | | | | | |
| [^] Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 1 | µg/kg | 1 | --- | --- | --- | --- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 1 | % | 91.2 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 1 | % | 84.5 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: BIOTA | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 50 | 130 |
| 13C8-PFOA | ---- | 50 | 130 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537238-AB | Page | : 1 of 6 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 16:15 |
| Order number | : ---- | Date Analysis Commenced | : 04-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 11:59 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 4 | | |
| No. of samples analysed | : 4 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- Amendment (22/12/2025): This report has been amended following the request to add additional analysis EP231X PFAS Full Suite to samples 004 and 006 as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to organise spill reports as requested by Michelle Agnew.
- EP231X (Biota): ALS NATA accreditation for PFAS in Biota covers all Perfluoroalkyl Sulfonic Acids, Perfluoroalkyl Carboxylic Acids and (n:2) Fluorotelomer Sulfonic Acids in fish (whole and muscle), plants and vegetable matrices, with the exception PFBA (fish only), EtFOSA, MeFOSE, EtFOSE, MeFOSAA, EtFOSAA.
- EP231: Stable isotope enriched internal standards are added to samples prior to extraction. Target compounds have a direct analogous internal standard with the exception of PFPeS, PFHpA, PFDS, PFTrDA and 10:2 FTS. These compounds use an internal standard that is chemically related and has a retention time close to that of the target compound. The DQO for internal standard response is 50-150% of that established at initial calibration. PFOS is quantified using a certified, traceable standard consisting of linear and branched PFOS isomers. These practices are in line with recommendations in the National Environmental Management Plan for PFAS (Australian HEPA) and also conform to QSM 5.3 (US DoD) requirements.



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | | Sample ID | ID008_BIOTA1 | ID008_BIOTA2 | ID008_BIOTA3 | ID008_BIOTA4 | ---- |
|---|------------|-----|-------|-------------------|-------------------|-------------------|-------------------|--------------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537238-003 | ES2537238-004 | ES2537238-005 | ES2537238-006 | ----- | |
| | | | | Result | Result | Result | Result | ---- | |
| Biota Sample Pre-Preparation | | | | | | | | | |
| ∅ Sample Description | ---- | - | -- | plant | plant | plant | plant | ---- | |
| ∅ Weight of Sample Prepared | ---- | 0.1 | g | 2.1 | 0.9 | 2.0 | 3.6 | ---- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluoropropane sulfonic acid (PFPrS) | 423-41-6 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) - Linear | 2795-39-3 | 1 | µg/kg | <1 | 1 | <1 | <1 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) - Branched | ---- | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 1 | µg/kg | <1 | 1 | <1 | <1 | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| Perfluorononane sulfonic acid (PFNS) | 68259-12-1 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 5 | µg/kg | <5 | <5 | <5 | 12 | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 2 | µg/kg | <2 | <2 | <2 | 21 | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 1 | µg/kg | 2 | <1 | <1 | 3 | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 1 | µg/kg | 1 | <1 | <1 | <1 | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 1 | µg/kg | 3 | <1 | <1 | <1 | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | | Sample ID | ID008_BIOTA1 | ID008_BIOTA2 | ID008_BIOTA3 | ID008_BIOTA4 | ---- |
|--|-------------|-----|-------|---------------|-------------------|-------------------|-------------------|-------------------|------|
| Sampling date / time | | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | ---- |
| Compound | CAS Number | LOR | Unit | ES2537238-003 | ES2537238-004 | ES2537238-005 | ES2537238-006 | ----- | |
| | | | | Result | Result | Result | Result | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 5 | µg/kg | <5 | <5 | <5 | <5 | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 5 | µg/kg | <5 | <5 | <5 | <5 | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 1 | µg/kg | <1 | <1 | <1 | <1 | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 2 | µg/kg | <2 | <2 | <2 | <2 | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| ^ Sum of PFAS | ---- | 1 | µg/kg | 6 | 1 | <1 | 36 | ---- | |



Analytical Results

| Sub-Matrix: BIOTA (Matrix: BIOTA) | | | | Sample ID | ID008_BIOTA1 | ID008_BIOTA2 | ID008_BIOTA3 | ID008_BIOTA4 | ---- |
|--------------------------------------|--------------------|-----|-------|---------------|-------------------|-------------------|-------------------|-------------------|------|
| Sampling date / time | | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | ---- |
| Compound | CAS Number | LOR | Unit | ES2537238-003 | ES2537238-004 | ES2537238-005 | ES2537238-006 | ----- | |
| | | | | Result | Result | Result | Result | ---- | |
| EP231P: PFAS Sums - Continued | | | | | | | | | |
| ^ Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 1 | µg/kg | <1 | 1 | <1 | <1 | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 1 | % | 93.6 | 99.4 | 102 | 106 | ---- | |
| 13C8-PFOA | ---- | 1 | % | 88.3 | 91.5 | 94.6 | 98.0 | ---- | |



Surrogate Control Limits

| Sub-Matrix: BIOTA | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 50 | 130 |
| 13C8-PFOA | ---- | 50 | 130 |

| Location Name | Date | type code hidden | PFAS Standard LOR | PFAS: low level | HOLD | Send to Envirolab | Comments |
|--------------------|------------|------------------|-------------------|-----------------|------|-------------------|---------------|
| A_TAP1 | 17/11/2025 | SW | | X | | | |
| A_TAP12 | 17/11/2025 | SW | | | X | | |
| A_TAP15 | 17/11/2025 | SW | | X | | | |
| A_TAP16A | 17/11/2025 | SW | | | X | | |
| A_TAP4 | 19/11/2025 | SW | | X | | | |
| A_TAP5 | 19/11/2025 | SW | | X | | | |
| A_TRUCKFILL_SS1 | 20/11/2025 | SOIL | X | | | | |
| A_TRUCKFILL_SS2 | 20/11/2025 | SOIL | | | X | | |
| AIRPORT_BORE | 17/11/2025 | SW | X | | | | Elevated PFAS |
| BALL_SED01 | 21/11/2025 | SED | | | X | | |
| BALL_SW01 | 21/11/2025 | SW | | | X | | |
| BALL_SS1 | 21/11/2025 | SOIL | X | | | | |
| BALL_SS2 | 21/11/2025 | SOIL | X | | | | |
| BALL_SS3 | 21/11/2025 | SOIL | X | | | | |
| Cockpit_SW01 | 18/11/2025 | SW | X | | | | |
| COUNCIL_TAP | 20/11/2025 | SW | | X | | | UPDATED COC |
| DEPOT_TANK1 | 17/11/2025 | SW | X | | | | |
| DEPOT_TANK2 | 17/11/2025 | SW | X | | | | |
| DEPOT_TANK3 | 17/11/2025 | SW | | | X | | |
| DEPOT_TAP | 17/11/2025 | SW | | X | | | |
| ELEC_TAP1 | 19/11/2025 | SW | | X | | | |
| FRE_TAP1 | 19/11/2025 | SW | | X | | | |
| FRE_TAP2 | 19/11/2025 | SW | X | | | | |
| FRE_TAP6 | 19/11/2025 | SW | | X | | | |
| ID008_SS1 | 20/11/2025 | SOIL | | | X | | |
| ID008_SS2 | 20/11/2025 | SOIL | X | | | | |
| ID013_BORE | 17/11/2025 | GW | X | | | | |
| ID014_BORE | 17/11/2025 | SW | X | | | | |
| ID016_BORE | 17/11/2025 | GW | X | | | | |
| ID019_BORE | 20/11/2025 | SW | X | | | | |
| MC_OMP01 | 20/11/2025 | SW | X | | | | |
| MC_OMP01 | 17/11/2025 | SW | | | X | | |
| MC_OMP02 | 20/11/2025 | SW | X | | | | |
| MC_OMP03 | 20/11/2025 | SW | X | | | | |
| MC_OMP04A | 20/11/2025 | SW | X | | | | |
| MC_OMP05A | 20/11/2025 | SW | X | | | | |
| MC_OMP06 | 20/11/2025 | SW | X | | | | |
| MC_OMP07A | 20/11/2025 | SW | X | | | | |
| MC_OMP09 | 17/11/2025 | SW | X | | | | |
| MC_OMP10 | 19/11/2025 | SW | X | | | | |
| MC_OMP11 | 17/11/2025 | SW | X | | | | |
| PWS_BUMBORA_TOILET | 19/11/2025 | SW | | | X | | |
| PWS_HEAD_DAM | 19/11/2025 | SW | X | | | | |

7/11/25

Andrew

28/11/25

Environmental Division
 Sydney
 Work Order Reference
ES2537238



Telephone : - 61-2-8784 8555

| | | | | | | | |
|------------------|------------|-------|---|---|---|---|----------------|
| PWS_HEAD_TOILET | 19/11/2025 | SW | | | X | | |
| PWS_HOSP_TANK1 | 18/11/2025 | SW | | | X | | |
| PWS_HOSP_TAP1 | 18/11/2025 | SW | | X | | | |
| PWS_HOSP_TAP3 | 18/11/2025 | SW | | X | | | |
| PWS_HOSP_TAP4 | 18/11/2025 | SW | | X | | | |
| PWS_HOSP_TAP6 | 18/11/2025 | SW | | X | | | |
| PWS_HOSP_TAP8 | 18/11/2025 | SW | | X | | | |
| WASTE_TAP1 | 20/11/2025 | SW | | X | | | |
| WC_OMP01 | 17/11/2025 | SW | X | | | | |
| WC_OMP02 | 17/11/2025 | SW | X | | | | |
| WC_OMP03 | 18/11/2025 | SW | X | | | | |
| WC_OMP04_DUCKDAM | 19/11/2025 | SED | X | | | | |
| WC_OMP04_DUCKDAM | 18/11/2025 | SW | X | | | | |
| WC_OMP05 | 18/11/2025 | SW | X | | | | |
| WC01 | 18/11/2025 | SW | | X | | | |
| WC02 | 18/11/2025 | SW | | X | | | |
| WW11_DAM | 20/11/2025 | SW | X | | | | Elevated PFAS |
| WW11_DAM | 17/11/2025 | SW | | | X | | Elevated PFAS |
| WC_BIOTA1 | 19/11/2025 | BIOTA | X | | | | water hyacinth |
| WC_BIOTA2 | 19/11/2025 | BIOTA | | | X | | Knot weed |
| ID008_BIOTA1 | 20/11/2025 | BIOTA | X | | | | silverbeet |
| ID008_BIOTA2 | 20/11/2025 | BIOTA | | | X | | parsley |
| ID008_BIOTA3 | 20/11/2025 | BIOTA | X | | | | zucchini |
| ID008_BIOTA4 | 20/11/2025 | BIOTA | | | X | | tomato |
| QC100 | | SOIL | X | | | | |
| QC101 | | SW | X | | | | |
| QC102 | | SW | X | | | | |
| QC103 | | SW | X | | | | |
| QC104 | | SW | X | | | | |
| QC105 | | SW | X | | | | |
| QC200 | | SOIL | | | | X | |
| QC201 | | SW | | | | X | |
| QC202 | | SW | | | | X | |
| QC203 | | SW | | | | X | |
| QC204 | | SW | | | | X | |
| QC205 | | SW | | | | X | |
| QC400 | | Water | | X | | | |
| QC500 | | Water | | X | | | |

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QUALITY CONTROL REPORT

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|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583 | Page | : 1 of 19 |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 15-Dec-2025 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 68 | | |
| No. of samples analysed | : 53 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| <i>Signatories</i> | <i>Position</i> | <i>Accreditation Category</i> |
|--------------------|---------------------|------------------------------------|
| Alex Rossi | Organic Chemist | Sydney Inorganics, Smithfield, NSW |
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |
| Edwandy Fadjar | Organic Coordinator | Sydney Inorganics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key :
 Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot
 CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
 LOR = Limit of reporting
 RPD = Relative Percentage Difference
 # = Indicates failed QC
 * = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

| Sub-Matrix: SOIL | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|-------------|--|------------|-----------------------------------|-------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EA055: Moisture Content (Dried @ 105-110°C) (QC Lot: 7064172) | | | | | | | | | |
| ES2537583-052 | ID008_SS2 | EA055: Moisture Content | ---- | 0.1 | % | 27.8 | 28.6 | 2.8 | 0% - 20% |
| ES2538662-002 | Anonymous | EA055: Moisture Content | ---- | 0.1 | % | 88.1 | 88.0 | 0.1 | 0% - 20% |
| EA055: Moisture Content (Dried @ 105-110°C) (QC Lot: 7071955) | | | | | | | | | |
| EB2543165-002 | Anonymous | EA055: Moisture Content | ---- | 0.1 | % | 4.8 | 5.2 | 6.2 | 0% - 20% |
| ES2538978-002 | Anonymous | EA055: Moisture Content | ---- | 0.1 | % | 61.9 | 63.4 | 2.4 | 0% - 20% |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7047711) | | | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | 0.0052 | 0.0048 | 7.2 | 0% - 20% |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7068728) | | | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | 0.0008 | 0.0008 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | 0.0100 | 0.0087 | 14.6 | 0% - 20% |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7047711) | | | | | | | | | |



Sub-Matrix: SOIL

| | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|-------------|---|------------|-----------------------------------|--------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7047711) - continued | | | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | 0.0004 | 0.0003 | 0.0 | No Limit |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | 0.0004 | 0.0004 | 0.0 | No Limit |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | 0.0004 | 0.0003 | 0.0 | No Limit |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | 0.0005 | 0.0005 | 0.0 | No Limit |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | 0.0003 | 0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | <0.001 | 0.0 | No Limit | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7068728) | | | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | <0.001 | 0.0 | No Limit | | |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7047711) | | | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7068728) | | | | | | | | | |



| Sub-Matrix: SOIL | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|-------------|---|-------------|-----------------------------------|-------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7068728) - continued | | | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7047711) | | | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7068728) | | | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0 | No Limit |
| Sub-Matrix: WATER | | | | Laboratory Duplicate (DUP) Report | | | | | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7057925) | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 3.87 | 3.90 | 0.9 | 0% - 20% |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 13.5 | 14.5 | 6.8 | 0% - 20% |
| | | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.39 | 0.37 | 3.4 | 0% - 50% |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.47 | 0.48 | 0.0 | 0% - 20% |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | 0.27 | 0.26 | 0.0 | 0% - 50% |



| Sub-Matrix: WATER | | | | Laboratory Duplicate (DUP) Report | | | | | | | |
|--|------------|--|------------|---|-----------|-----------------|------------------|---------|--------------------|-----|----------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) | | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7057925) - continued | | | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| ES2537583-004 | MC_OMP03 | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.37 | 0.38 | 3.0 | 0% - 20% | | |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 0.33 | 0.34 | 4.0 | 0% - 20% | | |
| | | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.06 | 0.07 | 0.0 | No Limit | | |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.06 | 0.06 | 0.0 | No Limit | | |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 7057928) | | | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 (0.05)* | µg/L | 6.50 | 6.58 | 1.1 | 0% - 20% | | |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 (0.05)* | µg/L | 16.3 | 14.4 | 12.5 | 0% - 20% | | |
| | | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.84 | 0.79 | 5.8 | 0% - 20% | | |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.71 | 0.74 | 4.2 | 0% - 20% | | |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | 0.48 | 0.46 | 4.1 | 0% - 20% | | |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7057925) | | | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.30 | 0.29 | 4.1 | 0% - 20% | | |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.12 | 0.12 | 0.0 | No Limit | | |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.49 | 0.53 | 7.5 | 0% - 20% | | |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.11 | 0.11 | 0.0 | No Limit | | |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit | | |
| | | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | <0.1 | 0.0 | No Limit | | |
| | | ES2537583-004 | MC_OMP03 | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.02 | 0.02 | 0.0 | No Limit |
| | | | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.13 | 0.13 | 0.0 | No Limit |
| | | | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.09 | 0.10 | 0.0 | No Limit |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | | | 0.02 | µg/L | 0.03 | 0.03 | 0.0 | No Limit | | |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | | | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | | | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | | | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | | | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | | | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit | | |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | | | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | | | 0.1 | µg/L | <0.1 | <0.1 | 0.0 | No Limit | | |



Sub-Matrix: **WATER**

Laboratory Duplicate (DUP) Report

| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
|--|-----------|---|------------|------|------|-----------------|------------------|---------|--------------------|
| EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 7057928) | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.47 | 0.48 | 0.0 | 0% - 20% |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.23 | 0.24 | 0.0 | 0% - 50% |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.92 | 0.88 | 4.3 | 0% - 20% |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.18 | 0.18 | 0.0 | No Limit |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | 0.1 | 0.1 | 0.0 | No Limit | | |
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7057925) | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| ES2537583-004 | MC_OMP03 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | 0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |

EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7057928)



Sub-Matrix: **WATER**

Laboratory Duplicate (DUP) Report

| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
|--|-----------|---|-------------|------|------|-----------------|------------------|---------|--------------------|
| EP231C: Perfluoroalkyl Sulfonamides (QC Lot: 7057928) - continued | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | <0.02 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7057925) | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| ES2537583-004 | MC_OMP03 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 7057928) | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | 0.0 | No Limit |
| EP231P: PFAS Sums (QC Lot: 7057925) | | | | | | | | | |

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 Work Order : ES2537583
 Client : SENVERSA PTY LTD
 Project : C17776 Norfolk DSI



| Sub-Matrix: WATER | | | | Laboratory Duplicate (DUP) Report | | | | | |
|--|-----------|---------------------|------------|-----------------------------------|------|-----------------|------------------|---------|--------------------|
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | LOR | Unit | Original Result | Duplicate Result | RPD (%) | Acceptable RPD (%) |
| EP231P: PFAS Sums (QC Lot: 7057925) - continued | | | | | | | | | |
| ES2537583-001 | WW11_DAM | EP231X: Sum of PFAS | ---- | 0.01 | µg/L | 19.7 | 20.8 | 5.2 | 0% - 20% |
| ES2537583-004 | MC_OMP03 | EP231X: Sum of PFAS | ---- | 0.01 | µg/L | 1.09 | 1.17 | 7.1 | 0% - 20% |
| EP231P: PFAS Sums (QC Lot: 7057928) | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Sum of PFAS | ---- | 0.01 (0.05)* | µg/L | 27.1 | 25.2 | 7.2 | 0% - 20% |



Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

Sub-Matrix: SOIL

| | | | | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | |
|---|------------|--------|-------|-----------------------------|---------------------------------------|------------------------|---------------------------|-----------------------|
| | | | | | Result | Spike Concentration | Spike Recovery (%) LCS | Acceptable Limits (%) |
| Method: Compound | CAS Number | LOR | Unit | Low | | | | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7047711) | | | | | | | | |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 104 | 72.0 | 128 |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 112 | 73.0 | 123 |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 67.0 | 130 |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 92.1 | 70.0 | 132 |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 104 | 68.0 | 136 |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 107 | 59.0 | 134 |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7068728) | | | | | | | | |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 91.4 | 72.0 | 128 |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 101 | 73.0 | 123 |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 94.6 | 67.0 | 130 |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 98.9 | 70.0 | 132 |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 94.2 | 68.0 | 136 |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 59.0 | 134 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7047711) | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | 0.00625 mg/kg | 110 | 71.0 | 135 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 102 | 69.0 | 132 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 111 | 70.0 | 132 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 113 | 71.0 | 131 |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 107 | 69.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 108 | 72.0 | 129 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 99.0 | 69.0 | 133 |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 97.4 | 64.0 | 136 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 69.0 | 135 |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 106 | 66.0 | 139 |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 95.5 | 69.0 | 133 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7068728) | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | 0.00625 mg/kg | 111 | 71.0 | 135 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 101 | 69.0 | 132 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 70.0 | 132 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 108 | 71.0 | 131 |



Sub-Matrix: SOIL

| | | | | Method Blank (MB) Report Result | Laboratory Control Spike (LCS) Report | | | |
|---|-------------|--------|-------|---------------------------------------|---------------------------------------|---------------------------|-----------------------------------|------|
| | | | | | Spike Concentration | Spike Recovery (%) LCS | Acceptable Limits (%) Low High | |
| Method: Compound | CAS Number | LOR | Unit | Result | Spike Concentration | Spike Recovery (%) LCS | Low | High |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7068728) - continued | | | | | | | | |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 69.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 105 | 72.0 | 129 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 94.6 | 69.0 | 133 |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 102 | 64.0 | 136 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 102 | 69.0 | 135 |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 108 | 66.0 | 139 |
| EP231X: Perfluorotetradecanoic acid (PFTEDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 113 | 69.0 | 133 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7047711) | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 117 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 123 | 71.6 | 129 |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 111 | 69.8 | 131 |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 112 | 68.7 | 130 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 119 | 65.1 | 134 |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 99.6 | 63.0 | 144 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 104 | 61.0 | 139 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7068728) | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 121 | 71.6 | 129 |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 104 | 69.8 | 131 |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 126 | 68.7 | 130 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | 0.00312 mg/kg | 112 | 65.1 | 134 |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 103 | 63.0 | 144 |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | 0.00125 mg/kg | 111 | 61.0 | 139 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7047711) | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 103 | 62.0 | 145 |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 116 | 64.0 | 140 |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 122 | 65.0 | 137 |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 112 | 69.2 | 143 |



Sub-Matrix: **SOIL**

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report Result | Laboratory Control Spike (LCS) Report | | | | |
|--|-------------|--------|-------|------------------------------------|---------------------------------------|--------------------|------|-----------------------|--|
| | | | | | Spike Concentration | Spike Recovery (%) | | Acceptable Limits (%) | |
| | | | | | | LCS | Low | High | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7068728) | | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 91.1 | 62.0 | 145 | |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 107 | 64.0 | 140 | |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 83.4 | 65.0 | 137 | |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | 0.00125 mg/kg | 108 | 69.2 | 143 | |

Sub-Matrix: **WATER**

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report Result | Laboratory Control Spike (LCS) Report | | | | |
|---|------------|-------|------|------------------------------------|---------------------------------------|--------------------|------|-----------------------|--|
| | | | | | Spike Concentration | Spike Recovery (%) | | Acceptable Limits (%) | |
| | | | | | | LCS | Low | High | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7055805) | | | | | | | | | |
| EP231X-LL: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 106 | 72.0 | 130 | |
| EP231X-LL: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 102 | 71.0 | 127 | |
| EP231X-LL: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 87.3 | 68.0 | 131 | |
| EP231X-LL: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 106 | 69.0 | 134 | |
| EP231X-LL: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 118 | 65.0 | 140 | |
| EP231X-LL: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 74.6 | 53.0 | 142 | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7057925) | | | | | | | | | |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 94.8 | 72.0 | 130 | |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 104 | 71.0 | 127 | |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 95.0 | 68.0 | 131 | |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 91.9 | 69.0 | 134 | |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 103 | 65.0 | 140 | |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 106 | 53.0 | 142 | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7057928) | | | | | | | | | |
| EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 108 | 72.0 | 130 | |
| EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 92.5 | 71.0 | 127 | |
| EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 101 | 68.0 | 131 | |
| EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 103 | 69.0 | 134 | |
| EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 93.9 | 65.0 | 140 | |
| EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 96.1 | 53.0 | 142 | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7058571) | | | | | | | | | |
| EP231X-LL: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 110 | 72.0 | 130 | |
| EP231X-LL: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 117 | 71.0 | 127 | |
| EP231X-LL: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 112 | 68.0 | 131 | |
| EP231X-LL: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 115 | 69.0 | 134 | |
| EP231X-LL: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 106 | 65.0 | 140 | |



Sub-Matrix: WATER

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | |
|---|------------|-------|------|--------------------------|---------------------------------------|--------------------|-----------------------|-----|
| | | | | Result | Spike Concentration | Spike Recovery (%) | Acceptable Limits (%) | |
| | | | | | LCS | Low | High | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7058571) - continued | | | | | | | | |
| EP231X-LL: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 94.8 | 53.0 | 142 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7055805) | | | | | | | | |
| EP231X-LL: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | 0.125 µg/L | 104 | 73.0 | 129 |
| EP231X-LL: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 98.8 | 72.0 | 129 |
| EP231X-LL: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 94.6 | 72.0 | 129 |
| EP231X-LL: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 97.8 | 72.0 | 130 |
| EP231X-LL: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 97.8 | 71.0 | 133 |
| EP231X-LL: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 104 | 69.0 | 130 |
| EP231X-LL: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 94.8 | 71.0 | 129 |
| EP231X-LL: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 95.5 | 69.0 | 133 |
| EP231X-LL: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 90.9 | 72.0 | 134 |
| EP231X-LL: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 81.4 | 65.0 | 144 |
| EP231X-LL: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 105 | 71.0 | 132 |
| EP231X-LL: Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 100 | 65.6 | 133 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7057925) | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | 1.25 µg/L | 111 | 73.0 | 129 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 95.7 | 72.0 | 129 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 112 | 72.0 | 129 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 108 | 72.0 | 130 |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 109 | 71.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 98.8 | 69.0 | 130 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 106 | 71.0 | 129 |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 99.7 | 69.0 | 133 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 101 | 72.0 | 134 |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 90.0 | 65.0 | 144 |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 107 | 71.0 | 132 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7057928) | | | | | | | | |
| EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | 1.25 µg/L | 104 | 73.0 | 129 |
| EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 106 | 72.0 | 129 |
| EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 116 | 72.0 | 129 |
| EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 104 | 72.0 | 130 |
| EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | 0.25 µg/L | 102 | 71.0 | 133 |
| EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 115 | 69.0 | 130 |
| EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 111 | 71.0 | 129 |



Sub-Matrix: WATER

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | |
|---|------------|-------|------|-----------------------------|---------------------------------------|--------------------|-----------------------|------|
| | | | | Result | Spike | Spike Recovery (%) | Acceptable Limits (%) | |
| | | | | | Concentration | LCS | Low | High |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7057928) - continued | | | | | | | | |
| EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 107 | 69.0 | 133 |
| EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 105 | 72.0 | 134 |
| EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 107 | 65.0 | 144 |
| EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 107 | 71.0 | 132 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7058571) | | | | | | | | |
| EP231X-LL: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | 0.125 µg/L | 106 | 73.0 | 129 |
| EP231X-LL: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 114 | 72.0 | 129 |
| EP231X-LL: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 114 | 72.0 | 129 |
| EP231X-LL: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 124 | 72.0 | 130 |
| EP231X-LL: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 114 | 71.0 | 133 |
| EP231X-LL: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 121 | 69.0 | 130 |
| EP231X-LL: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 110 | 71.0 | 129 |
| EP231X-LL: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 109 | 69.0 | 133 |
| EP231X-LL: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 118 | 72.0 | 134 |
| EP231X-LL: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 88.8 | 65.0 | 144 |
| EP231X-LL: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 120 | 71.0 | 132 |
| EP231X-LL: Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 117 | 65.6 | 133 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7055805) | | | | | | | | |
| EP231X-LL: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 108 | 67.0 | 137 |
| EP231X-LL: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 106 | 68.0 | 141 |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 98.0 | 61.1 | 139 |
| EP231X-LL: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 111 | 72.3 | 128 |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 107 | 63.2 | 134 |
| EP231X-LL: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 94.2 | 65.0 | 136 |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 106 | 61.0 | 135 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057925) | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 103 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 128 | 68.0 | 141 |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 113 | 62.6 | 147 |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 121 | 66.0 | 145 |



Sub-Matrix: WATER

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report | Laboratory Control Spike (LCS) Report | | | | |
|---|-------------|-------|------|-----------------------------|---------------------------------------|--------------------|------|-----------------------|--|
| | | | | Result | Spike Concentration | Spike Recovery (%) | | Acceptable Limits (%) | |
| | | | | | | LCS | Low | High | |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057925) - continued | | | | | | | | | |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 117 | 57.6 | 145 | |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 118 | 65.0 | 136 | |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 99.7 | 61.0 | 135 | |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057928) | | | | | | | | | |
| EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 110 | 67.0 | 137 | |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 103 | 68.0 | 141 | |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 95.8 | 62.6 | 147 | |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 113 | 66.0 | 145 | |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | 0.625 µg/L | 117 | 57.6 | 145 | |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 110 | 65.0 | 136 | |
| EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | 0.25 µg/L | 98.5 | 61.0 | 135 | |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7058571) | | | | | | | | | |
| EP231X-LL: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 107 | 67.0 | 137 | |
| EP231X-LL: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 127 | 68.0 | 141 | |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 121 | 61.1 | 139 | |
| EP231X-LL: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 123 | 72.3 | 128 | |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | 0.0625 µg/L | 128 | 63.2 | 134 | |
| EP231X-LL: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 121 | 65.0 | 136 | |
| EP231X-LL: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | 0.025 µg/L | 123 | 61.0 | 135 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7055805) | | | | | | | | | |
| EP231X-LL: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 86.4 | 63.0 | 143 | |
| EP231X-LL: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 86.7 | 64.0 | 140 | |
| EP231X-LL: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 112 | 67.0 | 138 | |
| EP231X-LL: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 95.6 | 75.2 | 137 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7057925) | | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 104 | 63.0 | 143 | |



Sub-Matrix: **WATER**

| Method: Compound | CAS Number | LOR | Unit | Method Blank (MB) Report Result | Laboratory Control Spike (LCS) Report | | | |
|--|-------------|-------|------|------------------------------------|---------------------------------------|--------------------|-----------------------|------|
| | | | | | Spike Concentration | Spike Recovery (%) | Acceptable Limits (%) | |
| | | | | | | LCS | Low | High |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7057925) - continued | | | | | | | | |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 90.2 | 64.0 | 140 |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 102 | 67.0 | 138 |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 101 | 71.4 | 144 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7057928) | | | | | | | | |
| EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 114 | 63.0 | 143 |
| EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 106 | 64.0 | 140 |
| EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 112 | 67.0 | 138 |
| EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | 0.25 µg/L | 95.4 | 71.4 | 144 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7058571) | | | | | | | | |
| EP231X-LL: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 128 | 63.0 | 143 |
| EP231X-LL: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 113 | 64.0 | 140 |
| EP231X-LL: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 109 | 67.0 | 138 |
| EP231X-LL: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | 0.025 µg/L | 98.0 | 75.2 | 137 |

Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

Sub-Matrix: **SOIL**

| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Matrix Spike (MS) Report | | | |
|---|-------------|--|------------|--------------------------|------------------|-----------------------|------|
| | | | | Spike Concentration | SpikeRecovery(%) | Acceptable Limits (%) | |
| | | | | | MS | Low | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7047711) | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.00125 mg/kg | 93.5 | 72.0 | 128 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.00125 mg/kg | 121 | 73.0 | 123 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.00125 mg/kg | 102 | 67.0 | 130 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.00125 mg/kg | 85.2 | 70.0 | 132 |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.00125 mg/kg | 104 | 68.0 | 136 |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.00125 mg/kg | 107 | 59.0 | 134 |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7068728) | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.00125 mg/kg | 93.6 | 72.0 | 128 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.00125 mg/kg | 101 | 73.0 | 123 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.00125 mg/kg | 91.6 | 67.0 | 130 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.00125 mg/kg | 105 | 70.0 | 132 |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.00125 mg/kg | 79.3 | 68.0 | 136 |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.00125 mg/kg | 102 | 59.0 | 134 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7047711) | | | | | | | |



Sub-Matrix: SOIL

| | | | | Matrix Spike (MS) Report | | | |
|---|-------------|---|------------|--------------------------|-------------------|-----------------------|------|
| | | | | Spike | Spike Recovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7047711) - continued | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.00625 mg/kg | 108 | 71.0 | 135 |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.00125 mg/kg | 95.7 | 69.0 | 132 |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.00125 mg/kg | 112 | 70.0 | 132 |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.00125 mg/kg | 117 | 71.0 | 131 |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.00125 mg/kg | 106 | 69.0 | 133 |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.00125 mg/kg | 105 | 72.0 | 129 |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.00125 mg/kg | 98.5 | 69.0 | 133 |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.00125 mg/kg | 101 | 64.0 | 136 |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.00125 mg/kg | 95.2 | 69.0 | 135 |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.00125 mg/kg | 94.5 | 66.0 | 139 |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.00312 mg/kg | 89.9 | 69.0 | 133 |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7068728) | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.00625 mg/kg | 112 | 71.0 | 135 |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.00125 mg/kg | 114 | 69.0 | 132 |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.00125 mg/kg | 104 | 70.0 | 132 |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.00125 mg/kg | 109 | 71.0 | 131 |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.00125 mg/kg | 106 | 69.0 | 133 |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.00125 mg/kg | 113 | 72.0 | 129 |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.00125 mg/kg | 102 | 69.0 | 133 |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.00125 mg/kg | 94.0 | 64.0 | 136 |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.00125 mg/kg | 108 | 69.0 | 135 |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.00125 mg/kg | 114 | 66.0 | 139 |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.00312 mg/kg | 121 | 69.0 | 133 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7047711) | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.00125 mg/kg | 116 | 67.0 | 137 |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.00312 mg/kg | 122 | 71.6 | 129 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.00312 mg/kg | 124 | 69.8 | 131 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.00312 mg/kg | 116 | 68.7 | 130 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.00312 mg/kg | 118 | 65.1 | 134 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.00125 mg/kg | 97.2 | 63.0 | 144 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.00125 mg/kg | 110 | 61.0 | 139 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7068728) | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.00125 mg/kg | 110 | 67.0 | 137 |



Sub-Matrix: **SOIL**

| | | | | Matrix Spike (MS) Report | | | |
|---|-------------|---|-------------|--------------------------|------------------|-----------------------|------|
| | | | | Spike | SpikeRecovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7068728) - continued | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.00312 mg/kg | 112 | 71.6 | 129 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.00312 mg/kg | 99.9 | 69.8 | 131 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.00312 mg/kg | 118 | 68.7 | 130 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.00312 mg/kg | 106 | 65.1 | 134 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.00125 mg/kg | 103 | 63.0 | 144 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.00125 mg/kg | 99.9 | 61.0 | 139 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7047711) | | | | | | | |
| ES2537583-052 | ID008_SS2 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.00125 mg/kg | 103 | 62.0 | 145 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.00125 mg/kg | 96.2 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.00125 mg/kg | 118 | 65.0 | 137 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.00125 mg/kg | 97.0 | 69.2 | 143 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7068728) | | | | | | | |
| ES2537583-046 | WC_OMP04_SD | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.00125 mg/kg | 97.4 | 62.0 | 145 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.00125 mg/kg | 91.7 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.00125 mg/kg | 81.2 | 65.0 | 137 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.00125 mg/kg | 98.0 | 69.2 | 143 |

Sub-Matrix: **WATER**

| | | | | Matrix Spike (MS) Report | | | |
|---|-----------|--|------------|--------------------------|------------------|-----------------------|------|
| | | | | Spike | SpikeRecovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7057925) | | | | | | | |
| ES2537583-003 | MC_OMP02 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.25 µg/L | 92.2 | 72.0 | 130 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.25 µg/L | 100 | 71.0 | 127 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.25 µg/L | 89.4 | 68.0 | 131 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.25 µg/L | 89.3 | 69.0 | 134 |
| | | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.25 µg/L | 95.5 | 65.0 | 140 |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.25 µg/L | 106 | 53.0 | 142 |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7057928) | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.25 µg/L | 106 | 72.0 | 130 |
| | | EP231X: Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.25 µg/L | 74.8 | 71.0 | 127 |
| | | EP231X: Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.25 µg/L | # Not Determined | 68.0 | 131 |
| | | EP231X: Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.25 µg/L | 84.9 | 69.0 | 134 |



Sub-Matrix: WATER

| | | | | Matrix Spike (MS) Report | | | | | |
|---|------------|---|------------|--|------------------|-----------------------|------|------|-----|
| | | | | Spike | SpikeRecovery(%) | Acceptable Limits (%) | | | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High | | |
| EP231A: Perfluoroalkyl Sulfonic Acids (QCLot: 7057928) - continued | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.25 µg/L | # Not Determined | 65.0 | 140 | | |
| | | EP231X: Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.25 µg/L | 94.4 | 53.0 | 142 | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7057925) | | | | | | | | | |
| ES2537583-003 | MC_OMP02 | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 1.25 µg/L | 109 | 73.0 | 129 | | |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.25 µg/L | 94.6 | 72.0 | 129 | | |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.25 µg/L | 106 | 72.0 | 129 | | |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.25 µg/L | 96.8 | 72.0 | 130 | | |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.25 µg/L | 98.6 | 71.0 | 133 | | |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.25 µg/L | 98.1 | 69.0 | 130 | | |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.25 µg/L | 104 | 71.0 | 129 | | |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.25 µg/L | 96.0 | 69.0 | 133 | | |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.25 µg/L | 102 | 72.0 | 134 | | |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.25 µg/L | 87.4 | 65.0 | 144 | | |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.625 µg/L | 110 | 71.0 | 132 | | |
| EP231B: Perfluoroalkyl Carboxylic Acids (QCLot: 7057928) | | | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorobutanoic acid (PFBA) | 375-22-4 | 1.25 µg/L | 105 | 73.0 | 129 | | |
| | | EP231X: Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.25 µg/L | 111 | 72.0 | 129 | | |
| | | EP231X: Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.25 µg/L | 95.9 | 72.0 | 129 | | |
| | | EP231X: Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.25 µg/L | 104 | 72.0 | 130 | | |
| | | EP231X: Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.25 µg/L | 105 | 71.0 | 133 | | |
| | | EP231X: Perfluorononanoic acid (PFNA) | 375-95-1 | 0.25 µg/L | 120 | 69.0 | 130 | | |
| | | EP231X: Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.25 µg/L | 114 | 71.0 | 129 | | |
| | | EP231X: Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.25 µg/L | 111 | 69.0 | 133 | | |
| | | EP231X: Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.25 µg/L | 103 | 72.0 | 134 | | |
| | | EP231X: Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.25 µg/L | 108 | 65.0 | 144 | | |
| | | EP231X: Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.625 µg/L | 109 | 71.0 | 132 | | |
| | | EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057925) | | | | | | | |
| | | ES2537583-003 | MC_OMP02 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.25 µg/L | 104 | 67.0 | 137 |
| EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | | | 0.625 µg/L | 116 | 68.0 | 141 | | |
| EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | | | 0.625 µg/L | 98.6 | 62.6 | 147 | | |
| EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | | | 0.625 µg/L | 123 | 66.0 | 145 | | |
| EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | | | 0.625 µg/L | 114 | 57.6 | 145 | | |
| EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | | | 0.25 µg/L | 107 | 65.0 | 136 | | |



Sub-Matrix: WATER

| | | | | Matrix Spike (MS) Report | | | |
|---|-----------|---|-------------|--------------------------|-------------------|-----------------------|------|
| | | | | Spike | Spike Recovery(%) | Acceptable Limits (%) | |
| Laboratory sample ID | Sample ID | Method: Compound | CAS Number | Concentration | MS | Low | High |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057925) - continued | | | | | | | |
| ES2537583-003 | MC_OMP02 | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.25 µg/L | 98.8 | 61.0 | 135 |
| EP231C: Perfluoroalkyl Sulfonamides (QCLot: 7057928) | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.25 µg/L | 109 | 67.0 | 137 |
| | | EP231X: N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.625 µg/L | 98.8 | 68.0 | 141 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.625 µg/L | 99.9 | 62.6 | 147 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.625 µg/L | 114 | 66.0 | 145 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.625 µg/L | 118 | 57.6 | 145 |
| | | EP231X: N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.25 µg/L | 122 | 65.0 | 136 |
| | | EP231X: N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.25 µg/L | 97.8 | 61.0 | 135 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7057925) | | | | | | | |
| ES2537583-003 | MC_OMP02 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.25 µg/L | 96.5 | 63.0 | 143 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.25 µg/L | 88.4 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.25 µg/L | 99.6 | 67.0 | 138 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.25 µg/L | 90.5 | 71.4 | 144 |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids (QCLot: 7057928) | | | | | | | |
| ES2537583-080 | MC_OMP01 | EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.25 µg/L | 113 | 63.0 | 143 |
| | | EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.25 µg/L | 121 | 64.0 | 140 |
| | | EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.25 µg/L | 120 | 67.0 | 138 |
| | | EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.25 µg/L | 79.8 | 71.4 | 144 |



QA/QC Compliance Assessment to assist with Quality Review

| | | | |
|--------------|---------------------------------|-------------------------|---------------------------------|
| Work Order | : ES2537583 | Page | : 1 of 10 |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 |
| Site | : ---- | Issue Date | : 15-Dec-2025 |
| Sampler | : Michelle Agnew Hollie Dunsten | No. of samples received | : 68 |
| Order number | : ---- | No. of samples analysed | : 53 |

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

Summary of Outliers

Outliers : Quality Control Samples

This report highlights outliers flagged in the Quality Control (QC) Report.

- **NO** Method Blank value outliers occur.
- **NO** Duplicate outliers occur.
- **NO** Laboratory Control outliers occur.
- Matrix Spike outliers exist - please see following pages for full details.
- For all regular sample matrices, where applicable to the methodology, **NO** surrogate recovery outliers occur.

Outliers : Analysis Holding Time Compliance

- Analysis Holding Time Outliers exist - please see following pages for full details.

Outliers : Frequency of Quality Control Samples

- Quality Control Sample Frequency Outliers exist - please see following pages for full details.



Outliers : Quality Control Samples

Duplicates, Method Blanks, Laboratory Control Samples and Matrix Spikes

Matrix: **WATER**

| Compound Group Name | Laboratory Sample ID | Client Sample ID | Analyte | CAS Number | Data | Limits | Comment |
|---------------------------------------|----------------------|------------------|---------------------------------------|------------|----------------|--------|---|
| Matrix Spike (MS) Recoveries | | | | | | | |
| EP231A: Perfluoroalkyl Sulfonic Acids | ES2537583--080 | MC_OMP01 | Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | Not Determined | ---- | MS recovery not determined, background level greater than or equal to 4x spike level. |
| EP231A: Perfluoroalkyl Sulfonic Acids | ES2537583--080 | MC_OMP01 | Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | Not Determined | ---- | MS recovery not determined, background level greater than or equal to 4x spike level. |

Outliers : Analysis Holding Time Compliance

Matrix: **SOIL**

| Method | Extraction / Preparation | | | Analysis | | | |
|--|---------------------------------|----------------|--------------------|--------------|---------------|------------------|--------------|
| | Container / Client Sample ID(s) | Date extracted | Due for extraction | Days overdue | Date analysed | Due for analysis | Days overdue |
| EA055: Moisture Content (Dried @ 105-110°C) | | | | | | | |
| HDPE Soil Jar QC100 | | ---- | ---- | ---- | 09-Dec-2025 | 03-Dec-2025 | 6 |
| HDPE Soil Jar WC_OMP04_SD | | ---- | ---- | ---- | 11-Dec-2025 | 03-Dec-2025 | 8 |
| HDPE Soil Jar ID008_SS2, A_TRUCKFILL_SS1 | | ---- | ---- | ---- | 09-Dec-2025 | 04-Dec-2025 | 5 |

Outliers : Frequency of Quality Control Samples

Matrix: **WATER**

| Quality Control Sample Type | Method | Count | | Rate (%) | | Quality Control Specification |
|--|-----------|-------|---------|----------|----------|--------------------------------|
| | | QC | Regular | Actual | Expected | |
| Laboratory Duplicates (DUP) | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS by LCMSMS) | EP231X-LL | 0 | 38 | 0.00 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 3 | 31 | 9.68 | 10.00 | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS by LCMSMS) | EP231X-LL | 0 | 38 | 0.00 | 5.00 | NEPM 2013 B3 & ALS QC Standard |

Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for VOC in soils vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: **SOIL**

Evaluation: * = Holding time breach ; ✓ = Within holding time.



Matrix: **SOIL**

Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EA055: Moisture Content (Dried @ 105-110°C) | | | | | | | |
| HDPE Soil Jar (EA055) QC100 | 19-Nov-2025 | ---- | ---- | ---- | 09-Dec-2025 | 03-Dec-2025 | * |
| HDPE Soil Jar (EA055) WC_OMP04_SD | 19-Nov-2025 | ---- | ---- | ---- | 11-Dec-2025 | 03-Dec-2025 | * |
| HDPE Soil Jar (EA055) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | ---- | ---- | ---- | 09-Dec-2025 | 04-Dec-2025 | * |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | |
| HDPE Soil Jar (EP231X) QC100 | 19-Nov-2025 | 03-Dec-2025 | 18-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_OMP04_SD | 19-Nov-2025 | 11-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 20-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | 03-Dec-2025 | 19-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | |
| HDPE Soil Jar (EP231X) QC100 | 19-Nov-2025 | 03-Dec-2025 | 18-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_OMP04_SD | 19-Nov-2025 | 11-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 20-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | 03-Dec-2025 | 19-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | |
| HDPE Soil Jar (EP231X) QC100 | 19-Nov-2025 | 03-Dec-2025 | 18-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_OMP04_SD | 19-Nov-2025 | 11-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 20-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | 03-Dec-2025 | 19-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | |
| HDPE Soil Jar (EP231X) QC100 | 19-Nov-2025 | 03-Dec-2025 | 18-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_OMP04_SD | 19-Nov-2025 | 11-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 20-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | 03-Dec-2025 | 19-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| EP231P: PFAS Sums | | | | | | | |
| HDPE Soil Jar (EP231X) QC100 | 19-Nov-2025 | 03-Dec-2025 | 18-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) WC_OMP04_SD | 19-Nov-2025 | 11-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 20-Jan-2026 | ✓ |
| HDPE Soil Jar (EP231X) ID008_SS2, A_TRUCKFILL_SS1 | 20-Nov-2025 | 03-Dec-2025 | 19-May-2026 | ✓ | 08-Dec-2025 | 12-Jan-2026 | ✓ |

Matrix: **WATER**

Evaluation: * = Holding time breach ; ✓ = Within holding time.



Matrix: **WATER** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | |
| HDPE (no PTFE) (EP231X-LL) DEPOT_TAP1, A_TAP1 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 10-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) MC_OMP09, MC_OMP11, WC_OMP01, WC_OMP02, DEPOT_TANK1, DEPOT_TANK2, AIRPORT_BORE, ID013_BORE, ID014_BORE, ID016_BORE, A_TAP15 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 12-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP8, PWS_HOSP_TAP6 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 10-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) WC_OMP03, WC_OMP04_DUCKDAM, WC_OMP05, COCKPIT_SW01, WC01, WC02 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 12-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) FRE_TAP1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) MC_OMP10, FRE_TAP2, PWS_HEAD_DAM, A_TAP4, A_TAP5, FRE_TAP6, ELEC_TAP1, MC_OMP01 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) WW11_DAM, MC_OMP02, MC_OMP03, MC_OMP04a, MC_OMP05a, MC_OMP06, MC_OMP07a, ID019_BORE, WASTE_TAP1, COUNCIL_TAP1, QC101, QC102, QC103, QC104, QC105 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 12-Dec-2025 | 19-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) QC500 | 21-Nov-2025 | 05-Dec-2025 | 20-May-2026 | ✓ | 12-Dec-2025 | 20-May-2026 | ✓ |



Matrix: **WATER** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|--|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | |
| HDPE (no PTFE) (EP231X-LL) DEPOT_TAP1, A_TAP1 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 10-Dec-2025 | 16-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) MC_OMP09, MC_OMP11, WC_OMP01, WC_OMP02, DEPOT_TANK1, DEPOT_TANK2, AIRPORT_BORE, ID013_BORE, ID014_BORE, ID016_BORE, A_TAP15 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 12-Dec-2025 | 16-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP8, PWS_HOSP_TAP6 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 10-Dec-2025 | 17-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) WC_OMP03, WC_OMP04_DUCKDAM, WC_OMP05, COCKPIT_SW01, WC01, WC02 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 12-Dec-2025 | 17-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) FRE_TAP1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 18-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X) MC_OMP10, FRE_TAP2, PWS_HEAD_DAM, A_TAP4, A_TAP5, FRE_TAP6, ELEC_TAP1, MC_OMP01 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 18-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X) WW11_DAM, MC_OMP02, MC_OMP03, MC_OMP04a, MC_OMP05a, MC_OMP06, MC_OMP07a, ID019_BORE, WASTE_TAP1, COUNCIL_TAP1, QC101, QC102, QC103, QC104, QC105 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 12-Dec-2025 | 19-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) QC500 | 21-Nov-2025 | 05-Dec-2025 | 20-May-2026 | ✓ | 12-Dec-2025 | 20-May-2026 | ✓ | |



Matrix: **WATER** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|--|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | |
| HDPE (no PTFE) (EP231X-LL) DEPOT_TAP1, A_TAP1 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 10-Dec-2025 | 16-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) MC_OMP09, MC_OMP11, WC_OMP01, WC_OMP02, DEPOT_TANK1, DEPOT_TANK2, AIRPORT_BORE, ID013_BORE, ID014_BORE, ID016_BORE, A_TAP15 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 12-Dec-2025 | 16-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP8, PWS_HOSP_TAP6 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 10-Dec-2025 | 17-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) WC_OMP03, WC_OMP04_DUCKDAM, WC_OMP05, COCKPIT_SW01, WC01, WC02 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 12-Dec-2025 | 17-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) FRE_TAP1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 18-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X) MC_OMP10, FRE_TAP2, PWS_HEAD_DAM, A_TAP4, A_TAP5, FRE_TAP6, ELEC_TAP1, MC_OMP01 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 18-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X) WW11_DAM, MC_OMP02, MC_OMP03, MC_OMP04a, MC_OMP05a, MC_OMP06, MC_OMP07a, ID019_BORE, WASTE_TAP1, COUNCIL_TAP1, QC101, QC102, QC103, QC104, QC105 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 12-Dec-2025 | 19-May-2026 | ✓ | |
| HDPE (no PTFE) (EP231X-LL) QC500 | 21-Nov-2025 | 05-Dec-2025 | 20-May-2026 | ✓ | 12-Dec-2025 | 20-May-2026 | ✓ | |



Matrix: **WATER** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | |
| HDPE (no PTFE) (EP231X-LL) DEPOT_TAP1, A_TAP1 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 10-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) MC_OMP09, MC_OMP11, WC_OMP01, WC_OMP02, DEPOT_TANK1, DEPOT_TANK2, AIRPORT_BORE, ID013_BORE, ID014_BORE, ID016_BORE, A_TAP15 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 12-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP8, PWS_HOSP_TAP6 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 10-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) WC_OMP03, WC_OMP04_DUCKDAM, WC_OMP05, COCKPIT_SW01, WC01, WC02 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 12-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) FRE_TAP1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) MC_OMP10, FRE_TAP2, PWS_HEAD_DAM, A_TAP4, A_TAP5, FRE_TAP6, ELEC_TAP1, MC_OMP01 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) WW11_DAM, MC_OMP02, MC_OMP03, MC_OMP04a, MC_OMP05a, MC_OMP06, MC_OMP07a, ID019_BORE, WASTE_TAP1, COUNCIL_TAP1, QC101, QC102, QC103, QC104, QC105 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 12-Dec-2025 | 19-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) QC500 | 21-Nov-2025 | 05-Dec-2025 | 20-May-2026 | ✓ | 12-Dec-2025 | 20-May-2026 | ✓ |



Matrix: **WATER** Evaluation: * = Holding time breach ; ✓ = Within holding time.

| Method Container / Client Sample ID(s) | Sample Date | Extraction / Preparation | | | Analysis | | |
|--|-------------|--------------------------|--------------------|------------|---------------|------------------|------------|
| | | Date extracted | Due for extraction | Evaluation | Date analysed | Due for analysis | Evaluation |
| EP231P: PFAS Sums | | | | | | | |
| HDPE (no PTFE) (EP231X-LL) DEPOT_TAP1, A_TAP1 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 10-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) MC_OMP09, MC_OMP11, WC_OMP01, WC_OMP02, DEPOT_TANK1, DEPOT_TANK2, AIRPORT_BORE, ID013_BORE, ID014_BORE, ID016_BORE, A_TAP15 | 17-Nov-2025 | 05-Dec-2025 | 16-May-2026 | ✓ | 12-Dec-2025 | 16-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) PWS_HOSP_TAP1, PWS_HOSP_TAP3, PWS_HOSP_TAP4, PWS_HOSP_TAP8, PWS_HOSP_TAP6 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 10-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) WC_OMP03, WC_OMP04_DUCKDAM, WC_OMP05, COCKPIT_SW01, WC01, WC02 | 18-Nov-2025 | 05-Dec-2025 | 17-May-2026 | ✓ | 12-Dec-2025 | 17-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) FRE_TAP1 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 10-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) MC_OMP10, FRE_TAP2, PWS_HEAD_DAM, A_TAP4, A_TAP5, FRE_TAP6, ELEC_TAP1, MC_OMP01 | 19-Nov-2025 | 05-Dec-2025 | 18-May-2026 | ✓ | 12-Dec-2025 | 18-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X) WW11_DAM, MC_OMP02, MC_OMP03, MC_OMP04a, MC_OMP05a, MC_OMP06, MC_OMP07a, ID019_BORE, WASTE_TAP1, COUNCIL_TAP1, QC101, QC102, QC103, QC104, QC105 | 20-Nov-2025 | 05-Dec-2025 | 19-May-2026 | ✓ | 12-Dec-2025 | 19-May-2026 | ✓ |
| HDPE (no PTFE) (EP231X-LL) QC500 | 21-Nov-2025 | 05-Dec-2025 | 20-May-2026 | ✓ | 12-Dec-2025 | 20-May-2026 | ✓ |



Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: **SOIL**

Evaluation: ✖ = Quality Control frequency not within specification ; ✔ = Quality Control frequency within specification.

| Quality Control Sample Type | Method | Count | | Rate (%) | | | Quality Control Specification |
|--|--------|-------|---------|----------|----------|------------|--------------------------------|
| | | QC | Regular | Actual | Expected | Evaluation | |
| Analytical Methods | | | | | | | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Moisture Content | EA055 | 4 | 33 | 12.12 | 10.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 4 | 50.00 | 10.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 4 | 50.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 4 | 50.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 4 | 50.00 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |

Matrix: **WATER**

Evaluation: ✖ = Quality Control frequency not within specification ; ✔ = Quality Control frequency within specification.

| Quality Control Sample Type | Method | Count | | Rate (%) | | | Quality Control Specification |
|--|-----------|-------|---------|----------|----------|------------|--------------------------------|
| | | QC | Regular | Actual | Expected | Evaluation | |
| Analytical Methods | | | | | | | |
| Laboratory Duplicates (DUP) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X-LL | 0 | 38 | 0.00 | 10.00 | ✖ | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 3 | 31 | 9.68 | 10.00 | ✖ | NEPM 2013 B3 & ALS QC Standard |
| Laboratory Control Samples (LCS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X-LL | 2 | 38 | 5.26 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 31 | 6.45 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Method Blanks (MB) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X-LL | 2 | 38 | 5.26 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 31 | 6.45 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |
| Matrix Spikes (MS) | | | | | | | |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X-LL | 0 | 38 | 0.00 | 5.00 | ✖ | NEPM 2013 B3 & ALS QC Standard |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | 2 | 31 | 6.45 | 5.00 | ✔ | NEPM 2013 B3 & ALS QC Standard |



Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

| Analytical Methods | Method | Matrix | Method Descriptions |
|--|-----------|--------|--|
| Moisture Content | EA055 | SOIL | In house: A gravimetric procedure based on weight loss over a 12 hour drying period at 105-110 degrees C. This method is compliant with NEPM Schedule B(3). |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | SOIL | In-house: Analysis of soils by solvent extraction followed by negative mode LC-ESI-MS/MS using MRM and isotope dilution or internal standard quantitation. A portion of homogenised sample is extracted along with isotope dilution standards (where commercially available) in a solution of ammonium acetate in acetonitrile/methanol. Where relevant, interferences from co-extracted organics are removed using dispersive clean-up media (dSPE). A portion of extract is combined with an equal volume of reagent water and filtered for instrumental analysis. |
| Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS | EP231X | WATER | In-house: Analysis of fresh and saline waters by solid phase extraction (SPE) followed by negative mode LC-ESI-MS/MS using MRM and isotope dilution or internal standard quantitation. Isotope dilution standards (where commercially available) and surrogates are added to the sample container. The entire contents are transferred to a weak anion exchange (WAX) solid phase extraction (SPE) cartridge. The sample container is successively rinsed with aliquots of the elution solvent. The eluted extract is combined with an equal volume of reagent water and a portion is filtered for analysis. Data quality objectives for internal standard recoveries are based on US EPA method 1633. |
| Per- and Polyfluoroalkyl Substances (PFAS by LCMSMS) | EP231X-LL | WATER | In-house: Analysis of fresh and saline waters by solid phase extraction (SPE) followed by negative mode LC-ESI-MS/MS using MRM and isotope dilution or internal standard quantitation. Isotope dilution standards (where commercially available) and surrogates are added to the sample container. The entire contents are transferred to a weak anion exchange (WAX) solid phase extraction (SPE) cartridge. The sample container is successively rinsed with aliquots of the elution solvent. The eluted extract is combined with an equal volume of reagent water and a portion is filtered for analysis. Data quality objectives for internal standard recoveries are based on US EPA method 1633. |
| Preparation Methods | Method | Matrix | Method Descriptions |
| QuEChERS Extraction of Solids | ORG71 | SOIL | In house: Sequential extractions with Acetonitrile/Methanol by shaking. Extraction efficiency aided by the addition of salts under acidic conditions. Where relevant, interferences from co-extracted organics are removed with dispersive clean-up media (dSPE). The extract is either diluted or concentrated and exchanged into the analytical solvent. |
| Solid Phase Extraction (SPE) for PFAS in water | ORG72 | WATER | In-house: Isotopically labelled analogues of target analytes used as internal standards and surrogates are added to the sample container. The entire contents are transferred to a solid phase extraction (SPE) cartridge. The sample container is successively rinsed with aliquots of the elution solvent. The eluted extract is combined with an equal volume of reagent water and a portion is filtered for analysis. Method procedures conform to US DoD QSM 5.3, table B-15 requirements. |



SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : **ES2537583**

| | | | |
|--------------|---|--------------|--|
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| E-mail | : michelle.agnew@senversa.com.au | E-mail | : andrew.wotherspoon@alsglobal.com |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Facsimile | : ---- | Facsimile | : +61-2-8784 8500 |
| Project | : C17776 Norfolk DSI | Page | : 1 of 4 |
| Order number | : ---- | Quote number | : EB2023SENVVER0001 (EN/000) |
| C-O-C number | : ---- | QC Level | : NEPM 2013 B3 & ALS QC Standard |
| Site | : ---- | | |
| Sampler | : Michelle Agnew Hollie Dunsten | | |

Dates

| | | | |
|---------------------------|---------------------|--------------------------|----------------------|
| Date Samples Received | : 24-Nov-2025 08:45 | Issue Date | : 10-Dec-2025 |
| Client Requested Due Date | : 12-Dec-2025 | Scheduled Reporting Date | : 12-Dec-2025 |

Delivery Details

| | | | |
|----------------------|-------------|------------------------------------|---|
| Mode of Delivery | : Undefined | Security Seal | : Intact. |
| No. of coolers/boxes | : 2 | Temperature | : 21.6°C, 22.3°C & 21.4°C - Ice Bricks present |
| Receipt Detail | : | No. of samples received / analysed | : 68 / 53 |

General Comments

- This report contains the following information:
 - Sample Container(s)/Preservation Non-Compliances
 - Summary of Sample(s) and Requested Analysis
 - Proactive Holding Time Report
 - Requested Deliverables
- Sample 80 received as extra.
- Sample ID ID008_551 was not received.
- **Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.**
- 10/12/2025: This is an updated SRN which indicates the addition of PFAS analysis to sample 46 as requested by Michelle.
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Unless otherwise stated, analytical work for this work order will be conducted at ALS Sydney, NATA accreditation no. 825, site no. 10911.
- Sample Disposal - Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.



Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

- No sample container / preservation non-compliance exists.

Summary of Sample(s) and Requested Analysis

Some items described below may be part of a laboratory process necessary for the execution of client requested tasks. Packages may contain additional analyses, such as the determination of moisture content and preparation tasks, that are included in the package.

If no sampling time is provided, the sampling time will default 00:00 on the date of sampling. If no sampling date is provided, the sampling date will be assumed by the laboratory and displayed in brackets without a time component

Matrix: **SOIL**

| Laboratory sample ID | Sampling date / time | Sample ID | (On Hold) SOIL No analysis requested | SOIL - EA055-103 Moisture Content | SOIL - EP231X (solids) PFAS - Full Suite (30 analytes) |
|----------------------|----------------------|-----------------|---|--------------------------------------|---|
| ES2537583-046 | 19-Nov-2025 00:00 | WC_OMP04_SD | | ✓ | ✓ |
| ES2537583-051 | 20-Nov-2025 00:00 | ID008_SS1 | ✓ | | |
| ES2537583-052 | 20-Nov-2025 00:00 | ID008_SS2 | | ✓ | ✓ |
| ES2537583-054 | 20-Nov-2025 00:00 | A_TRUCKFILL_SS2 | ✓ | | |
| ES2537583-055 | 20-Nov-2025 00:00 | A_TRUCKFILL_SS1 | | ✓ | ✓ |
| ES2537583-056 | 21-Nov-2025 00:00 | BALL_SS3 | ✓ | | |
| ES2537583-057 | 21-Nov-2025 00:00 | BALL_SS4 | ✓ | | |
| ES2537583-058 | 21-Nov-2025 00:00 | BALL_SS5 | ✓ | | |
| ES2537583-060 | 19-Nov-2025 00:00 | QC100 | | ✓ | ✓ |

Matrix: **WATER**

| Laboratory sample ID | Sampling date / time | Sample ID | (On Hold) WATER No analysis requested | WATER - EP231X PFAS - Full Suite (31 analytes) | WATER - EP231X-LL PFAS - Full Suite (31 analytes) Low Level |
|----------------------|----------------------|-----------|--|---|--|
| ES2537583-001 | 20-Nov-2025 00:00 | WW11_DAM | | ✓ | |
| ES2537583-002 | 17-Nov-2025 00:00 | MC_OMP01 | ✓ | | |
| ES2537583-003 | 20-Nov-2025 00:00 | MC_OMP02 | | ✓ | |
| ES2537583-004 | 20-Nov-2025 00:00 | MC_OMP03 | | ✓ | |
| ES2537583-005 | 20-Nov-2025 00:00 | MC_OMP04a | | ✓ | |
| ES2537583-006 | 20-Nov-2025 00:00 | MC_OMP05a | | ✓ | |
| ES2537583-007 | 20-Nov-2025 00:00 | MC_OMP06 | | ✓ | |
| ES2537583-008 | 20-Nov-2025 00:00 | MC_OMP07a | | ✓ | |
| ES2537583-009 | 17-Nov-2025 00:00 | MC_OMP09 | | ✓ | |
| ES2537583-010 | 19-Nov-2025 00:00 | MC_OMP10 | | ✓ | |
| ES2537583-011 | 17-Nov-2025 00:00 | MC_OMP11 | | ✓ | |
| ES2537583-012 | 17-Nov-2025 00:00 | WC_OMP01 | | ✓ | |
| ES2537583-013 | 17-Nov-2025 00:00 | WC_OMP02 | | ✓ | |
| ES2537583-014 | 18-Nov-2025 00:00 | WC_OMP03 | | ✓ | |



| | | | (On Hold) WATER No analysis requested | WATER - EP231X PFAS - Full Suite (31 analytes) | WATER - EP231X-LL PFAS - Full Suite (31 analytes) Low Level |
|---------------|-------------------|--------------------|--|---|--|
| ES2537583-015 | 18-Nov-2025 00:00 | WC_OMP04_DUCKDAM | | ✓ | |
| ES2537583-016 | 18-Nov-2025 00:00 | WC_OMP05 | | ✓ | |
| ES2537583-017 | 18-Nov-2025 00:00 | COCKPIT_SW01 | | ✓ | |
| ES2537583-018 | 19-Nov-2025 00:00 | FRE_TAP1 | | | ✓ |
| ES2537583-019 | 19-Nov-2025 00:00 | FRE_TAP2 | | ✓ | |
| ES2537583-020 | 17-Nov-2025 00:00 | DEPOT_TAP1 | | | ✓ |
| ES2537583-021 | 17-Nov-2025 00:00 | DEPOT_TANK1 | | ✓ | |
| ES2537583-022 | 17-Nov-2025 00:00 | DEPOT_TANK2 | | ✓ | |
| ES2537583-023 | 17-Nov-2025 00:00 | DEPOT_TANK3 | ✓ | | |
| ES2537583-024 | 17-Nov-2025 00:00 | A_TAP1 | | | ✓ |
| ES2537583-025 | 17-Nov-2025 00:00 | AIRPORT_BORE | | ✓ | |
| ES2537583-026 | 19-Nov-2025 00:00 | PWS_HEAD_DAM | | ✓ | |
| ES2537583-027 | 19-Nov-2025 00:00 | PWS_HEAD_TOILET | ✓ | | |
| ES2537583-028 | 19-Nov-2025 00:00 | PWS_BUMBORA_TOILET | ✓ | | |
| ES2537583-029 | 17-Nov-2025 00:00 | ID013_BORE | | ✓ | |
| ES2537583-030 | 17-Nov-2025 00:00 | ID014_BORE | | ✓ | |
| ES2537583-031 | 20-Nov-2025 00:00 | ID019_BORE | | ✓ | |
| ES2537583-032 | 17-Nov-2025 00:00 | ID016_BORE | | ✓ | |
| ES2537583-033 | 18-Nov-2025 00:00 | PWS_HOSP_TAP1 | | | ✓ |
| ES2537583-034 | 18-Nov-2025 00:00 | PWS_HOSP_TAP3 | | | ✓ |
| ES2537583-035 | 18-Nov-2025 00:00 | PWS_HOSP_TAP4 | | | ✓ |
| ES2537583-036 | 18-Nov-2025 00:00 | PWS_HOSP_TAP8 | | | ✓ |
| ES2537583-037 | 18-Nov-2025 00:00 | PWS_HOSP_TAP6 | | | ✓ |
| ES2537583-038 | 18-Nov-2025 00:00 | PWS_HOSP_TANK1 | ✓ | | |
| ES2537583-039 | 17-Nov-2025 00:00 | A_TAP12 | ✓ | | |
| ES2537583-040 | 17-Nov-2025 00:00 | A_TAP16A | ✓ | | |
| ES2537583-041 | 17-Nov-2025 00:00 | A_TAP15 | | | ✓ |
| ES2537583-042 | 19-Nov-2025 00:00 | A_TAP4 | | | ✓ |
| ES2537583-043 | 19-Nov-2025 00:00 | A_TAP5 | | | ✓ |
| ES2537583-044 | 18-Nov-2025 00:00 | WC01 | | | ✓ |
| ES2537583-045 | 18-Nov-2025 00:00 | WC02 | | | ✓ |
| ES2537583-047 | 19-Nov-2025 00:00 | FRE_TAP6 | | | ✓ |
| ES2537583-048 | 19-Nov-2025 00:00 | ELEC_TAP1 | | | ✓ |
| ES2537583-049 | 20-Nov-2025 00:00 | WASTE_TAP1 | | | ✓ |
| ES2537583-050 | 20-Nov-2025 00:00 | COUNCIL_TAP1 | | | ✓ |
| ES2537583-061 | 20-Nov-2025 00:00 | QC101 | | ✓ | |
| ES2537583-062 | 20-Nov-2025 00:00 | QC102 | | ✓ | |
| ES2537583-063 | 20-Nov-2025 00:00 | QC103 | | ✓ | |
| ES2537583-064 | 20-Nov-2025 00:00 | QC104 | | ✓ | |
| ES2537583-065 | 20-Nov-2025 00:00 | QC105 | | ✓ | |
| ES2537583-066 | 21-Nov-2025 00:00 | QC500 | | | ✓ |



| | | | (On Hold) WATER No analysis requested | WATER - EP231X PFAS - Full Suite (31 analytes) | WATER - EP231X-LL PFAS - Full Suite (31 analytes) Low Level |
|---------------|-------------------|----------|--|---|--|
| ES2537583-067 | 20-Nov-2025 00:00 | QC400 | ✓ | | |
| ES2537583-068 | 21-Nov-2025 00:00 | BALL_SW1 | ✓ | | |
| ES2537583-080 | 19-Nov-2025 00:00 | MC_OMP01 | | ✓ | |

Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

Requested Deliverables

MICHELLE AGNEW

- *AU Certificate of Analysis - NATA (COA) Email michelle.agnew@senversa.com.au
- *AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) Email michelle.agnew@senversa.com.au
- *AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) Email michelle.agnew@senversa.com.au
- A4 - AU Sample Receipt Notification - Environmental HT (SRN) Email michelle.agnew@senversa.com.au
- Chain of Custody (CoC) (COC) Email michelle.agnew@senversa.com.au
- EDI Format - ESDAT (ESDAT) Email michelle.agnew@senversa.com.au

SUPPLIER ACCOUNTS

- A4 - AU Tax Invoice (INV) Email supplieraccounts@senversa.com.au



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AA | Page | : 1 of 33 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:12 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 50 | | |
| No. of samples analysed | : 49 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|------------------------------------|
| Alex Rossi | Organic Chemist | Sydney Inorganics, Smithfield, NSW |
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20mL or 125mL bottles have been tested in accordance with the QSM5.4 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20ml or 125ml bottles have been tested in accordance with the QSM5.3 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spill reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.
- EP231: Stable isotope enriched internal standards are added to samples prior to extraction. Target compounds have a direct analogous internal standard with the exception of PFPeS, PFHpA, PFDS, PFTrDA and 10:2 FTS. These compounds use an internal standard that is chemically related and has a retention time close to that of the target compound. The DQO for internal standard response is 50-150% of that established at initial calibration or as per USEPA 1633 limits where listed. PFOS is quantified using a certified, traceable standard consisting of linear and branched PFOS isomers. These practices are in line with recommendations in the National Environmental Management Plan for PFAS and also conform to QSM 5.4 (US DoD) requirements.



Analytical Results

| Sub-Matrix: SEDIMENT (Matrix: SOIL) | | | | Sample ID | WC_OMP04_SD | QC100 | ---- | ---- | ---- |
|--|------------|--------|-------|-------------------|-------------------|-------|-------|-------|------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-046 | ES2537583-060 | ----- | ----- | ----- | |
| | | | | Result | Result | ---- | ---- | ---- | |
| EA055: Moisture Content (Dried @ 105-110°C) | | | | | | | | | |
| Moisture Content | ---- | 0.1 | % | 51.6 | 51.8 | ---- | ---- | ---- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | 0.0008 | 0.0009 | ---- | ---- | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | 0.0100 | 0.0083 | ---- | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | <0.001 | ---- | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: SEDIMENT (Matrix: SOIL) | | | | Sample ID | WC_OMP04_SD | QC100 | ---- | ---- | ---- |
|---|--------------------|--------|-------|-------------------|-------------------|-------|-------|-------|------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-046 | ES2537583-060 | ----- | ----- | ----- | |
| | | | | Result | Result | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | <0.0005 | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.0002 | mg/kg | 0.0108 | 0.0092 | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.0002 | mg/kg | 0.0108 | 0.0092 | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.0002 | mg/kg | 0.0108 | 0.0092 | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.0002 | % | 91.6 | 89.1 | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.0002 | % | 92.8 | 85.2 | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: SOIL (Matrix: SOIL) | | | | Sample ID | A_TRUCKFILL_SS1 | BALL_SS3 | BALL_SS4 | BALL_SS5 | ---- |
|--|------------|--------|-------|-------------------|-------------------|-------------------|-------------------|-------------------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-055 | ES2537583-056 | ES2537583-057 | ES2537583-058 | ----- | |
| | | | | Result | Result | Result | Result | ---- | |
| EA055: Moisture Content (Dried @ 105-110°C) | | | | | | | | | |
| Moisture Content | ---- | 0.1 | % | 30.8 | 27.3 | 13.9 | 12.2 | ---- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | <0.0002 | <0.0002 | <0.0002 | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | <0.0002 | <0.0002 | <0.0002 | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | 0.0009 | 0.0033 | 0.0028 | 0.0005 | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | 0.0004 | <0.0002 | <0.0002 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | 0.0841 | 0.0437 | 0.0197 | 0.0084 | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | 0.0002 | 0.0159 | 0.0105 | 0.0006 | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | <0.001 | 0.043 | 0.011 | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | 0.0002 | 0.0002 | 0.167 | 0.0541 | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | 0.0005 | 0.0004 | 0.0918 | 0.0569 | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | 0.0004 | 0.0462 | 0.0503 | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | 0.0012 | 0.0005 | 0.0601 | 0.0680 | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | 0.0008 | 0.0361 | 0.0260 | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0182 | 0.0706 | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | <0.0002 | 0.0003 | 0.0049 | 0.0934 | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | <0.0002 | 0.0004 | 0.0025 | 0.0713 | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0005 | 0.0317 | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0010 | 0.0348 | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | 0.0015 | <0.0002 | ---- | |



Analytical Results

| Sub-Matrix: SOIL (Matrix: SOIL) | | | | Sample ID | A_TRUCKFILL_SS1 | BALL_SS3 | BALL_SS4 | BALL_SS5 | ---- |
|---|--------------------|--------|-------|-------------------|-------------------|-------------------|-------------------|-------------------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | 21-Nov-2025 00:00 | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-055 | ES2537583-056 | ES2537583-057 | ES2537583-058 | ----- | |
| | | | | Result | Result | Result | Result | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | <0.0005 | <0.0005 | <0.0005 | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | <0.0005 | <0.0005 | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | <0.0005 | <0.0005 | <0.0005 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | <0.0005 | <0.0005 | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | <0.0002 | <0.0002 | <0.0002 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | <0.0002 | <0.0002 | <0.0002 | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.0032 | <0.0005 | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.829 | 1.77 | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 2.28 | 4.07 | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | <0.0005 | 0.129 | 0.549 | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.0002 | mg/kg | 0.0877 | 0.0762 | 3.75 | 6.97 | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.0002 | mg/kg | 0.0850 | 0.0470 | 0.0225 | 0.0089 | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.0002 | mg/kg | 0.0869 | 0.0485 | 3.54 | 6.09 | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.0002 | % | 85.2 | 98.6 | 93.8 | 105 | ---- | |
| 13C8-PFOA | ---- | 0.0002 | % | 88.8 | 84.4 | 89.4 | 96.8 | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WW11_DAM | MC_OMP02 | MC_OMP03 | MC_OMP04a | MC_OMP05a |
|--|------------|------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 |
| Compound | CAS Number | LOR | Unit | ES2537583-001 | ES2537583-003 | ES2537583-004 | ES2537583-005 | ES2537583-006 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.39 | 0.04 | 0.06 | 0.38 | 0.39 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.47 | 0.04 | 0.06 | 0.42 | 0.43 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 3.87 | 0.24 | 0.37 | 3.18 | 3.19 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | 0.27 | <0.02 | <0.02 | 0.19 | 0.17 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 13.5 | 0.05 | 0.33 | 10.7 | 5.77 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | <0.1 | <0.1 | 0.1 | 0.1 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.12 | 0.13 | 0.13 | 0.17 | 0.17 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.49 | 0.07 | 0.09 | 0.53 | 0.56 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.11 | 0.03 | 0.03 | 0.10 | 0.10 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.30 | 0.02 | 0.02 | 0.22 | 0.23 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WW11_DAM | MC_OMP02 | MC_OMP03 | MC_OMP04a | MC_OMP05a |
|---|--------------------|------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-----------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-001 | ES2537583-003 | ES2537583-004 | ES2537583-005 | ES2537583-006 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 19.7 | 0.62 | 1.09 | 16.2 | 11.3 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 17.4 | 0.29 | 0.70 | 13.9 | 8.96 | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 18.8 | 0.58 | 1.03 | 15.4 | 10.5 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 100 | 90.5 | 109 | 102 | 95.9 | |
| 13C8-PFOA | ---- | 0.02 | % | 107 | 98.1 | 100.0 | 93.8 | 99.1 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | MC_OMP06 | MC_OMP07a | MC_OMP09 | MC_OMP10 | MC_OMP11 |
|--|------------|------|------|-------------------|-------------------|-------------------|-------------------|-------------------|----------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-007 | ES2537583-008 | ES2537583-009 | ES2537583-010 | ES2537583-011 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.39 | 0.39 | 0.31 | 0.24 | 0.26 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.41 | 0.43 | 0.33 | 0.26 | 0.30 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 3.37 | 2.96 | 2.43 | 2.04 | 2.31 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | 0.18 | 0.17 | 0.14 | 0.12 | 0.12 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 5.09 | 5.38 | 3.99 | 3.49 | 3.79 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | 0.1 | 0.1 | <0.1 | <0.1 | <0.1 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.16 | 0.18 | 0.10 | 0.10 | 0.10 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.55 | 0.52 | 0.35 | 0.31 | 0.31 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.10 | 0.08 | 0.06 | 0.06 | 0.06 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.22 | 0.22 | 0.14 | 0.14 | 0.13 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | MC_OMP06 | MC_OMP07a | MC_OMP09 | MC_OMP10 | MC_OMP11 |
|---|--------------------|------|------|-------------------|-------------------|-------------------|-------------------|-------------------|----------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-007 | ES2537583-008 | ES2537583-009 | ES2537583-010 | ES2537583-011 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 10.7 | 10.6 | 7.98 | 6.86 | 7.49 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 8.46 | 8.34 | 6.42 | 5.53 | 6.10 | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 9.98 | 9.83 | 7.38 | 6.38 | 6.96 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 94.6 | 98.5 | 107 | 101 | 102 | |
| 13C8-PFOA | ---- | 0.02 | % | 97.6 | 98.2 | 98.8 | 99.5 | 99.5 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCKDA M | WC_OMP05 |
|--|------------|------|------|-------------------|-------------------|-------------------|-------------------|----------------------|----------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-012 | ES2537583-013 | ES2537583-014 | ES2537583-015 | ES2537583-016 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.11 | 0.06 | <0.02 | <0.02 | <0.02 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.10 | 0.07 | <0.02 | <0.02 | <0.02 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.73 | 0.43 | 0.12 | 0.10 | 0.03 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 0.28 | 0.12 | 0.05 | 0.04 | 0.01 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.04 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.12 | 0.06 | <0.02 | <0.02 | <0.02 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.02 | <0.01 | <0.01 | <0.01 | <0.01 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC_OMP01 | WC_OMP02 | WC_OMP03 | WC_OMP04_DUCKDA M | WC_OMP05 |
|---|--------------------|------|------|-------------------|-------------------|-------------------|-------------------|----------------------|----------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-012 | ES2537583-013 | ES2537583-014 | ES2537583-015 | ES2537583-016 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 1.44 | 0.76 | 0.17 | 0.14 | 0.04 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 1.01 | 0.55 | 0.17 | 0.14 | 0.04 | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 1.30 | 0.67 | 0.17 | 0.14 | 0.04 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 96.4 | 103 | 103 | 104 | 100 | |
| 13C8-PFOA | ---- | 0.02 | % | 99.0 | 100.0 | 103 | 102 | 103 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COCKPIT_SW01 | FRE_TAP1 | FRE_TAP2 | DEPOT_TAP1 | DEPOT_TANK1 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-017 | ES2537583-018 | ES2537583-019 | ES2537583-020 | ES2537583-021 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.01 | ---- | 0.02 | ---- | <0.01 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 0.01 | ---- | 0.03 | ---- | <0.01 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | ---- | <0.01 | ---- | <0.01 | ---- | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | <0.1 | ---- | <0.1 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COCKPIT_SW01 | FRE_TAP1 | FRE_TAP2 | DEPOT_TAP1 | DEPOT_TANK1 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-017 | ES2537583-018 | ES2537583-019 | ES2537583-020 | ES2537583-021 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | ---- | <0.01 | ---- | <0.01 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COCKPIT_SW01 | FRE_TAP1 | FRE_TAP2 | DEPOT_TAP1 | DEPOT_TANK1 |
|---|-------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-017 | ES2537583-018 | ES2537583-019 | ES2537583-020 | ES2537583-021 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | <0.02 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COCKPIT_SW01 | FRE_TAP1 | FRE_TAP2 | DEPOT_TAP1 | DEPOT_TANK1 |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-017 | ES2537583-018 | ES2537583-019 | ES2537583-020 | ES2537583-021 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids - Continued | | | | | | | | | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | ---- | <0.005 | ---- | <0.005 | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | <0.05 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Sum of PFAS | ---- | 0.01 | µg/L | 0.02 | ---- | 0.05 | ---- | <0.01 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 0.02 | ---- | 0.05 | ---- | <0.01 | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | <0.002 | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 0.02 | ---- | 0.05 | ---- | <0.01 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | ---- | 94.1 | ---- | 89.5 | ---- | |
| 13C4-PFOS | ---- | 0.02 | % | 113 | ---- | 102 | ---- | 105 | |
| 13C8-PFOA | ---- | 0.002 | % | ---- | 92.5 | ---- | 90.5 | ---- | |
| 13C8-PFOA | ---- | 0.02 | % | 100 | ---- | 98.3 | ---- | 101 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | DEPOT_TANK2 | A_TAP1 | AIRPORT_BORE | PWS_HEAD_DAM | PWS_HOSP_TAP1 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-022 | ES2537583-024 | ES2537583-025 | ES2537583-026 | ES2537583-033 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | ---- | 0.61 | <0.02 | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | ---- | 0.54 | <0.02 | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | <0.01 | ---- | 6.05 | <0.01 | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | 0.34 | <0.02 | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | <0.01 | ---- | 12.5 | <0.01 | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | ---- | <0.01 | ---- | ---- | <0.01 | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | 0.1 | <0.1 | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | ---- | 0.16 | <0.02 | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | ---- | 0.66 | <0.02 | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | ---- | 0.14 | <0.02 | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | DEPOT_TANK2 | A_TAP1 | AIRPORT_BORE | PWS_HEAD_DAM | PWS_HOSP_TAP1 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-022 | ES2537583-024 | ES2537583-025 | ES2537583-026 | ES2537583-033 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | ---- | 0.35 | <0.01 | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | DEPOT_TANK2 | A_TAP1 | AIRPORT_BORE | PWS_HEAD_DAM | PWS_HOSP_TAP1 |
|---|-------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-022 | ES2537583-024 | ES2537583-025 | ES2537583-026 | ES2537583-033 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | <0.02 | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | DEPOT_TANK2 | A_TAP1 | AIRPORT_BORE | PWS_HEAD_DAM | PWS_HOSP_TAP1 |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 17-Nov-2025 00:00 | 19-Nov-2025 00:00 | 18-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-022 | ES2537583-024 | ES2537583-025 | ES2537583-026 | ES2537583-033 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids - Continued | | | | | | | | | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | <0.005 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | <0.05 | <0.05 | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Sum of PFAS | ---- | 0.01 | µg/L | <0.01 | ---- | 21.7 | <0.01 | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | <0.01 | ---- | 18.6 | <0.01 | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | <0.002 | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | <0.01 | ---- | 20.6 | <0.01 | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | ---- | 92.2 | ---- | ---- | 89.0 | |
| 13C4-PFOS | ---- | 0.02 | % | 92.5 | ---- | 102 | 100 | ---- | |
| 13C8-PFOA | ---- | 0.002 | % | ---- | 89.3 | ---- | ---- | 88.9 | |
| 13C8-PFOA | ---- | 0.02 | % | 100 | ---- | 94.4 | 91.4 | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | PWS_HOSP_TAP3 | PWS_HOSP_TAP4 | PWS_HOSP_TAP8 | PWS_HOSP_TAP6 | A_TAP15 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-034 | ES2537583-035 | ES2537583-036 | ES2537583-037 | ES2537583-041 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | PWS_HOSP_TAP3 | PWS_HOSP_TAP4 | PWS_HOSP_TAP8 | PWS_HOSP_TAP6 | A_TAP15 |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|---------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 18-Nov-2025 00:00 | 17-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-034 | ES2537583-035 | ES2537583-036 | ES2537583-037 | ES2537583-041 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | 95.5 | 98.3 | 92.6 | 94.4 | 103 | |
| 13C8-PFOA | ---- | 0.002 | % | 91.6 | 92.7 | 90.5 | 93.7 | 95.5 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | A_TAP4 | A_TAP5 | FRE_TAP6 | ELEC_TAP1 | WASTE_TAP1 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|------------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-042 | ES2537583-043 | ES2537583-047 | ES2537583-048 | ES2537583-049 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | 0.007 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | 0.008 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | 0.060 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | 0.005 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | 0.148 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | 0.004 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | 0.014 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | 0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | 0.005 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | A_TAP4 | A_TAP5 | FRE_TAP6 | ELEC_TAP1 | WASTE_TAP1 |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|------------|
| Sampling date / time | | | | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 19-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-042 | ES2537583-043 | ES2537583-047 | ES2537583-048 | ES2537583-049 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | 0.253 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | 0.208 | <0.002 | <0.002 | <0.002 | <0.002 | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | 0.240 | <0.002 | <0.002 | <0.002 | <0.002 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | 97.9 | 104 | 104 | 101 | 100 | |
| 13C8-PFOA | ---- | 0.002 | % | 96.1 | 94.3 | 93.9 | 92.8 | 93.6 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COUNCIL_TAP1 | QC101 | QC102 | QC103 | QC104 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-050 | ES2537583-061 | ES2537583-062 | ES2537583-063 | ES2537583-064 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | ---- | 0.45 | 0.89 | 0.49 | 0.09 | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | ---- | 0.40 | 0.74 | 0.42 | 0.07 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | ---- | 4.20 | 7.04 | 4.04 | 0.55 | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | ---- | 0.28 | 0.49 | 0.22 | 0.02 | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | ---- | 12.8 | 12.4 | 4.51 | 0.47 | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | ---- | 0.1 | 0.1 | <0.1 | <0.1 | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | ---- | 0.14 | 0.23 | 0.17 | 0.14 | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | ---- | 0.53 | 0.92 | 0.59 | 0.11 | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | ---- | 0.10 | 0.17 | 0.10 | 0.03 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COUNCIL_TAP1 | QC101 | QC102 | QC103 | QC104 |
|--|------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-050 | ES2537583-061 | ES2537583-062 | ES2537583-063 | ES2537583-064 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | ---- | 0.29 | 0.48 | 0.21 | 0.03 | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COUNCIL_TAP1 | QC101 | QC102 | QC103 | QC104 |
|---|-------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-050 | ES2537583-061 | ES2537583-062 | ES2537583-063 | ES2537583-064 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | ---- | <0.02 | <0.02 | <0.02 | <0.02 | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | COUNCIL_TAP1 | QC101 | QC102 | QC103 | QC104 |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|-------------------|-------------------|-------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | 20-Nov-2025 00:00 | |
| Compound | CAS Number | LOR | Unit | ES2537583-050 | ES2537583-061 | ES2537583-062 | ES2537583-063 | ES2537583-064 | |
| | | | | Result | Result | Result | Result | Result | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids - Continued | | | | | | | | | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | ---- |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | ---- | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Sum of PFAS | ---- | 0.01 | µg/L | ---- | 19.5 | 23.8 | 10.9 | 1.54 | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | ---- | 17.0 | 19.4 | 8.55 | 1.02 | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | ---- | 18.6 | 22.2 | 10.1 | 1.42 | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | 102 | ---- | ---- | ---- | ---- | ---- |
| 13C4-PFOS | ---- | 0.02 | % | ---- | 97.6 | 94.8 | 96.8 | 97.4 | |
| 13C8-PFOA | ---- | 0.002 | % | 101 | ---- | ---- | ---- | ---- | ---- |
| 13C8-PFOA | ---- | 0.02 | % | ---- | 93.2 | 92.1 | 91.1 | 87.3 | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | QC105 | QC500 | MC_OMP01 | ---- | ---- |
|--|------------|-------|------|-------------------|-------------------|-------------------|----------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-065 | ES2537583-066 | ES2537583-080 | ----- | ----- | |
| | | | | Result | Result | Result | ---- | ---- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.05 | ---- | 0.84 | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.03 | ---- | 0.71 | ---- | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.28 | ---- | 6.50 | ---- | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | 0.48 | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 0.05 | ---- | 16.3 | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | ---- | <0.01 | ---- | ---- | ---- | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | 0.1 | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.15 | ---- | 0.23 | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.09 | ---- | 0.92 | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.03 | ---- | 0.18 | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | QC105 | QC500 | MC_OMP01 | ---- | ---- |
|--|------------|-------|------|-------------------|-------------------|-------------------|----------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-065 | ES2537583-066 | ES2537583-080 | ----- | ----- | |
| | | | | Result | Result | Result | ---- | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids - Continued | | | | | | | | | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.02 | ---- | 0.47 | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | QC105 | QC500 | MC_OMP01 | ---- | ---- |
|---|-------------|-------|------|-------------------|-------------------|-------------------|----------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-065 | ES2537583-066 | ES2537583-080 | ----- | ----- | |
| | | | | Result | Result | Result | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | <0.02 | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | QC105 | QC500 | MC_OMP01 | ---- | ---- |
|---|--------------------|-------|------|-------------------|-------------------|-------------------|----------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | 21-Nov-2025 00:00 | 19-Nov-2025 00:00 | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-065 | ES2537583-066 | ES2537583-080 | ----- | ----- | |
| | | | | Result | Result | Result | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids - Continued | | | | | | | | | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | ---- | <0.005 | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | <0.05 | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Sum of PFAS | ---- | 0.01 | µg/L | 0.70 | ---- | 27.1 | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 0.33 | ---- | 22.8 | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | ---- | <0.002 | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 0.67 | ---- | 25.5 | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | ---- | 108 | ---- | ---- | ---- | |
| 13C4-PFOS | ---- | 0.02 | % | 95.7 | ---- | 99.3 | ---- | ---- | |
| 13C8-PFOA | ---- | 0.002 | % | ---- | 100 | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.02 | % | 89.4 | ---- | 88.4 | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: SEDIMENT | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |

| Sub-Matrix: SOIL | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AB | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| <i>Signatories</i> | <i>Position</i> | <i>Accreditation Category</i> |
|--------------------|---------------------|------------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |
| Edwandy Fadjar | Organic Coordinator | Sydney Inorganics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

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Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
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^ = This result is computed from individual analyte detections at or above the level of reporting
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~ = Indicates an estimated value.

- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spill reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.
- EP231: Stable isotope enriched internal standards are added to samples prior to extraction. Target compounds have a direct analogous internal standard with the exception of PFPeS, PFHpA, PFDS, PFTrDA and 10:2 FTS. These compounds use an internal standard that is chemically related and has a retention time close to that of the target compound. The DQO for internal standard response is 50-150% of that established at initial calibration or as per USEPA 1633 limits where listed. PFOS is quantified using a certified, traceable standard consisting of linear and branched PFOS isomers. These practices are in line with recommendations in the National Environmental Management Plan for PFAS and also conform to QSM 5.4 (US DoD) requirements.



Analytical Results

| Sub-Matrix: SOIL (Matrix: SOIL) | | | Sample ID | ID008_SS2 | ---- | ---- | ---- | ---- |
|--|------------|--------|-------------------|---------------|-------|-------|-------|-------|
| Sampling date / time | | | 20-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-052 | ----- | ----- | ----- | ----- |
| | | | | Result | ---- | ---- | ---- | ---- |
| EA055: Moisture Content (Dried @ 105-110°C) | | | | | | | | |
| Moisture Content | ---- | 0.1 | % | 27.8 | ---- | ---- | ---- | ---- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.0002 | mg/kg | 0.0052 | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.001 | mg/kg | <0.001 | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.0002 | mg/kg | 0.0004 | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.0002 | mg/kg | 0.0004 | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.0002 | mg/kg | 0.0004 | ---- | ---- | ---- | ---- |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.0002 | mg/kg | 0.0005 | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.0002 | mg/kg | 0.0003 | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: SOIL (Matrix: SOIL) | | | | Sample ID | ID008_SS2 | ---- | ---- | ---- | ---- |
|---|--------------------|--------|-------|-------------------|-----------|-------|-------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-052 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.0002 | mg/kg | <0.0002 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.0005 | mg/kg | <0.0005 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.0002 | mg/kg | 0.0072 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.0002 | mg/kg | 0.0052 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.0002 | mg/kg | 0.0056 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.0002 | % | 91.0 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.0002 | % | 91.4 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: SOIL | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AC | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



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- General Comments
- Analytical Results
- Surrogate Control Limits

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Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

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LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20mL or 125mL bottles have been tested in accordance with the QSM5.4 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spilt reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID013_BORE | ---- | ---- | ---- | ---- |
|--|------------|------|------|-------------------|------------|-------|-------|-------|-------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-029 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- | --- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID013_BORE | ---- | ---- | ---- | ---- |
|---|--------------------|------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-029 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 96.5 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.02 | % | 89.9 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AD | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
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| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



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| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



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- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID014_BORE | ---- | ---- | ---- | ---- |
|--|------------|------|------|-------------------|------------|-------|-------|-------|-------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-030 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- | --- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.31 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.26 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 2.20 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | 0.13 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 2.90 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | 0.11 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.35 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | 0.06 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | 0.13 | ---- | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID014_BORE | ---- | ---- | ---- | ---- |
|---|--------------------|------|------|-------------------|------------|-------|-------|-------|-------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-030 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- | --- |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 6.57 | ---- | ---- | ---- | ---- | ---- |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 5.10 | ---- | ---- | ---- | ---- | ---- |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 6.06 | ---- | ---- | ---- | ---- | ---- |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 95.7 | ---- | ---- | ---- | ---- | ---- |
| 13C8-PFOA | ---- | 0.02 | % | 90.3 | ---- | ---- | ---- | ---- | ---- |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AE | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

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- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spilt reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID019_BORE | ---- | ---- | ---- | ---- |
|--|------------|------|------|----------------------|-------------------|-------|-------|-------|-------|
| | | | | Sampling date / time | 20-Nov-2025 00:00 | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-031 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | ---- | ---- | ---- | ---- | ---- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | 0.03 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | 0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.12 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | 0.07 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | 0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | ---- |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID019_BORE | ---- | ---- | ---- | ---- |
|---|--------------------|------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 20-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-031 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 0.26 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 0.19 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 0.24 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 98.6 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.02 | % | 92.4 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AF | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 03-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



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- Analytical Results
- Surrogate Control Limits

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Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

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- Amendment (12/01/2026): This report has been amended following the request to organise spilt reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID016_BORE | ---- | ---- | ---- | ---- |
|--|------------|------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-032 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.01 | µg/L | 0.09 | ---- | ---- | ---- | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.1 | µg/L | <0.1 | ---- | ---- | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | ID016_BORE | ---- | ---- | ---- | ---- |
|---|--------------------|------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 17-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-032 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.02 | µg/L | <0.02 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.05 | µg/L | <0.05 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.01 | µg/L | 0.09 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.01 | µg/L | 0.09 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.01 | µg/L | 0.09 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.02 | % | 94.9 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.02 | % | 91.0 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AG | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 05-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



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| <i>Signatories</i> | <i>Position</i> | <i>Accreditation Category</i> |
|--------------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



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LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20ml or 125ml bottles have been tested in accordance with the QSM5.3 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spilt reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC-03_BORE | ---- | ---- | ---- | ---- |
|--|------------|-------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-044 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | 0.009 | ---- | ---- | ---- | ---- | |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | 0.008 | ---- | ---- | ---- | ---- | |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | 0.016 | ---- | ---- | ---- | ---- | |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | 0.004 | ---- | ---- | ---- | ---- | |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC-03_BORE | ---- | ---- | ---- | ---- |
|---|--------------------|-------|------|-------------------|------------|-------|-------|-------|------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-044 | ----- | ----- | ----- | ----- | |
| | | | | Result | ---- | ---- | ---- | ---- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | 0.037 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | 0.020 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | 0.029 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | 102 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.002 | % | 92.4 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |



CERTIFICATE OF ANALYSIS

| | | | |
|-------------------------|---|-------------------------|---|
| Work Order | : ES2537583-AH | Page | : 1 of 5 |
| Amendment | : 2 | | |
| Client | : SENVERSA PTY LTD | Laboratory | : Environmental Division Sydney |
| Contact | : MICHELLE AGNEW | Contact | : Andrew Wotherspoon |
| Address | : Level 24, 1 Market St, SYDNEY NSW 2000 | Address | : 277-289 Woodpark Road Smithfield NSW Australia 2164 |
| Telephone | : ---- | Telephone | : +61-2-8784 8555 |
| Project | : C17776 Norfolk DSI | Date Samples Received | : 24-Nov-2025 08:45 |
| Order number | : ---- | Date Analysis Commenced | : 05-Dec-2025 |
| C-O-C number | : ---- | Issue Date | : 12-Feb-2026 12:13 |
| Sampler | : Michelle Agnew Hollie Dunsten | | |
| Site | : ---- | | |
| Quote number | : EN/000 | | |
| No. of samples received | : 1 | | |
| No. of samples analysed | : 1 | | |



This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

| Signatories | Position | Accreditation Category |
|-------------|-----------------|----------------------------------|
| Alex Rossi | Organic Chemist | Sydney Organics, Smithfield, NSW |



General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.
LOR = Limit of reporting
^ = This result is computed from individual analyte detections at or above the level of reporting
ø = ALS is not NATA accredited for these tests.
~ = Indicates an estimated value.

- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20ml or 125ml bottles have been tested in accordance with the QSM5.3 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- Amendment (16/12/2025): This report has been amended following the request to add additional analysis EP231X Full Suite PFAS to samples 056, 057, and 058.
- Amendment (12/01/2026): This report has been amended following the request to organise spilt reports as requested by Michelle Agnew.
- Amendment (12/02/2026): This report has been amended following the request to update sample ID #44 to WC-03_BORE and #45 to WC-04 as requested by Michelle Agnew.



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC-04 | ---- | ---- | ---- | ---- |
|--|------------|-------|------|----------------------|-------------------|-------|-------|-------|-------|
| | | | | Sampling date / time | 18-Nov-2025 00:00 | ---- | ---- | ---- | ---- |
| Compound | CAS Number | LOR | Unit | ES2537583-045 | ----- | ----- | ----- | ----- | ----- |
| | | | | Result | --- | --- | --- | --- | --- |
| EP231A: Perfluoroalkyl Sulfonic Acids | | | | | | | | | |
| Perfluorobutane sulfonic acid (PFBS) | 375-73-5 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentane sulfonic acid (PFPeS) | 2706-91-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexane sulfonic acid (PFHxS) | 355-46-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptane sulfonic acid (PFHpS) | 375-92-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctane sulfonic acid (PFOS) | 1763-23-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecane sulfonic acid (PFDS) | 335-77-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| EP231B: Perfluoroalkyl Carboxylic Acids | | | | | | | | | |
| Perfluorobutanoic acid (PFBA) | 375-22-4 | 0.01 | µg/L | <0.01 | ---- | ---- | ---- | ---- | ---- |
| Perfluoropentanoic acid (PFPeA) | 2706-90-3 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexanoic acid (PFHxA) | 307-24-4 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroheptanoic acid (PFHpA) | 375-85-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorooctanoic acid (PFOA) | 335-67-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorononanoic acid (PFNA) | 375-95-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorodecanoic acid (PFDA) | 335-76-2 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluoroundecanoic acid (PFUnDA) | 2058-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorododecanoic acid (PFDoDA) | 307-55-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotridecanoic acid (PFTrDA) | 72629-94-8 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |
| Perfluorotetradecanoic acid (PFTeDA) | 376-06-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | ---- |
| Perfluorohexadecanoic acid (PFHxDA) | 67905-19-5 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | ---- |
| EP231C: Perfluoroalkyl Sulfonamides | | | | | | | | | |
| Perfluorooctane sulfonamide (FOSA) | 754-91-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | ---- |



Analytical Results

| Sub-Matrix: WATER (Matrix: WATER) | | | | Sample ID | WC-04 | ---- | ---- | ---- | ---- |
|---|--------------------|-------|------|-------------------|-------|-------|-------|-------|------|
| Sampling date / time | | | | 18-Nov-2025 00:00 | ---- | ---- | ---- | ---- | |
| Compound | CAS Number | LOR | Unit | ES2537583-045 | ----- | ----- | ----- | ----- | |
| | | | | Result | --- | --- | --- | --- | |
| EP231C: Perfluoroalkyl Sulfonamides - Continued | | | | | | | | | |
| N-Methyl perfluorooctane sulfonamide (MeFOSA) | 31506-32-8 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamide (EtFOSA) | 4151-50-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoethanol (MeFOSE) | 24448-09-7 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoethanol (EtFOSE) | 1691-99-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| N-Methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA) | 2355-31-9 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| N-Ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA) | 2991-50-6 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| EP231D: (n:2) Fluorotelomer Sulfonic Acids | | | | | | | | | |
| 4:2 Fluorotelomer sulfonic acid (4:2 FTS) | 757124-72-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 6:2 Fluorotelomer sulfonic acid (6:2 FTS) | 27619-97-2 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 8:2 Fluorotelomer sulfonic acid (8:2 FTS) | 39108-34-4 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| 10:2 Fluorotelomer sulfonic acid (10:2 FTS) | 120226-60-0 | 0.005 | µg/L | <0.005 | ---- | ---- | ---- | ---- | |
| EP231P: PFAS Sums | | | | | | | | | |
| Sum of PFAS | ---- | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Sum of PFHxS and PFOS | 355-46-4/1763-23-1 | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| Sum of PFAS (WA DER List) | ---- | 0.002 | µg/L | <0.002 | ---- | ---- | ---- | ---- | |
| EP231S: PFAS Surrogate | | | | | | | | | |
| 13C4-PFOS | ---- | 0.002 | % | 101 | ---- | ---- | ---- | ---- | |
| 13C8-PFOA | ---- | 0.002 | % | 93.0 | ---- | ---- | ---- | ---- | |



Surrogate Control Limits

| Sub-Matrix: WATER | | Recovery Limits (%) | |
|-------------------------------|------------|---------------------|------|
| Compound | CAS Number | Low | High |
| EP231S: PFAS Surrogate | | | |
| 13C4-PFOS | ---- | 60 | 120 |
| 13C8-PFOA | ---- | 60 | 120 |

Chain of Custody Documentation

Laboratory: ALS NSW
Address: 277-289 Woodpark Road, Smithfield
Contact: Sample Receipt
Phone:

| | | | |
|------------------|--------------------------------|-------------------|--------|
| Job Number: | C17776 | Purchase Order: | |
| Project Name: | Norfolk DSI | Quote No.: | |
| Sampled By: | | Turn Around Time: | |
| Project Manager: | | Page: | 2 of 5 |
| Email Report To: | michelle.agnew@senversa.com.au | Phone/Mobile: | |

| Sample Information | | | | Container Information | | | | Analysis Required | | | | | | | | | | | | | | | | |
|--------------------|--------------------|----------|----------|-----------------------|-------------|---------------|----|-------------------|--|---|---|--|--|--|--|--|--|--|--|--|--|--|--|--|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | | | | | | | | | | | | | | | | | | |
| ✓ 20 | DEPOT-TAPI | WATER | 17/11/25 | | | | | | | | | | | | | | | | | | | | | |
| ✓ 21 | DEPOT-TANK1 | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 22 | DEPOT-TANK2 | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 23 | DEPOT-TANK3 | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 24 | A-TAPI | | 17/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 25 | AIRPORT BORE | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 26 | PWS-HEAD-DAM | | 19/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 27 | PWS-HEAD-TOILET | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 28 | PWS-BUMBORA-TOILET | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 29 | ID013-BORE | | 17/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 30 | ID014-BORE | | " | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 31 | ID015-BORE | | 20/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 32 | ID016-BORE | | 17/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| ✓ 33 | PWS-HOSP-TAPI | | 18/11/25 | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 34 | PWS-HOSP-TAP3 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 35 | PWS-HOSP-TAP4 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 36 | PWS-HOSP-TAP5 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 37 | PWS-HOSP-TAP6 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 38 | PWS-HOSP-TAP7 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 39 | PWS-HOSP-TAP8 | | | | | | | | | X | X | | | | | | | | | | | | | |
| ✓ 39 | A-TAP2 | | 17/11/25 | | | | X | X | | | | | | | | | | | | | | | | |
| Total | | | | | | | 20 | | | | | | | | | | | | | | | | | |

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: Sampler Name: michelle agnew Signature: [Signature] Date: 21/11/25

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|-----------------|-------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic;
 V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic;
 F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____

Chain of Custody Documentation

Laboratory: ALS NSW
Address: 277-289 Woodpark Road, Smithfield
Contact: Sample Receipt
Phone:

Job Number: C17776 Purchase Order:
Project Name: Norfolk DSI Quote No:
Sampled By: Turn Around Time:
Project Manager: Page: 3 of 5
Email Report To: michelle.agnew@senversa.com.au Phone/Mobile:

| Sample Information | | Container Information | | Analysis Required | | | | | | | | | | Comments: e.g. Highly contaminate sample; hazardous materials prese LORs etc. | | | | | | | |
|--------------------|-----------------------|-----------------------|----------|-------------------|-------------|---------------|--|--|--|--|--|--|--|---|--|--|--|--|--|--|--|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | | | | | | | | | | | | | | | |
| ✓ 40 | A-TAP316A | Water | 17/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 41 | A-TAP5 | | 19/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 42 | A-TAP4 | | 19/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 43 | A-TAP5 | | " | | | | | | | | | | | | | | | | | | |
| ✓ 44 | WC01 | | 18/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 45 | WC02 | | 18/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 46 | WC-OMPOT_SD | SED. | 19/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 47 | FRE-TAP6 | Water | " | | | | | | | | | | | | | | | | | | |
| ✓ 48 | WASTE-TAP1 | | " | | | | | | | | | | | | | | | | | | |
| ✓ 49 | WASTE-TAP1 | | 20/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 50 | COUNCIL-TAP1 | | | | | | | | | | | | | | | | | | | | |
| ✓ 51 | ID008-SS1 | SOIL | | | | | | | | | | | | | | | | | | | |
| ✓ 52 | ID008-SS2 | | | | | | | | | | | | | | | | | | | | |
| ✓ 53 | WASTE-DIITAL | | | | | | | | | | | | | | | | | | | | |
| ✓ 54 | WASTE-A TRUCKFILL-SS2 | | | | | | | | | | | | | | | | | | | | |
| ✓ 55 | A TRUCKFILL-SS1 | | | | | | | | | | | | | | | | | | | | |
| ✓ 56 | BALL-SS3 | 2 SOI | 21/11/25 | | | | | | | | | | | | | | | | | | |
| ✓ 57 | BALL-SS4 | | " | | | | | | | | | | | | | | | | | | |
| ✓ 58 | BALL-SS5 | | " | | | | | | | | | | | | | | | | | | |
| ✓ 59 | BALL-SD1 | sed | " | | | | | | | | | | | | | | | | | | |
| Total | | 15+ | | | | | | | | | | | | | | | | | | | |

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: Sampler Name: michelle Agnew Signature: [Signature] Date: 21/11/25

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|-----------------|-------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic; V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's Iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by: _____
Checked by: _____

Chain of Custody Documentation

Laboratory:
Address:
Contact:
Phone:

Analysis Required

Comments: e.g. Highly contaminated sample; hazardous materials present; LORs etc.

| Job Number: C17776 | | Purchase Order: | | | | | |
|---|------------------|-------------------|----------|-----------------------|-------------|---------------|---|
| Project Name: Norfolk DSI | | Quote No: | | | | | |
| Sampled By: | | Turn Around Time: | | | | | |
| Project Manager: | | Page: 4 of 45 | | | | | |
| Email Report To: michelle.agnew@senversa.com.au | | Phone/Mobile: | | | | | |
| Sample Information | | | | Container Information | | | |
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles | |
| 60 | QC100 | SED | 19/11/25 | | | | |
| 61 | QC101 | WATER | 20/11/25 | | | | X |
| 62 | QC102 | | | | | | |
| 63 | QC103 | | | | | | |
| 64 | QC104 | | | | | | |
| 65 | QC105 | | | | | | |
| | QC106 | | | | | | |
| | QC200 | SED | 19/11/25 | | | | |
| | QC201 | WATER | 20/11/25 | | | | |
| | QC202 | | | | | | |
| | QC203 | | | | | | |
| | QC204 | | | | | | |
| | QC205 | | | | | | |
| | QC206 | | | | | | |
| 66 | QC500 | WATER | 20/11/25 | | | | |
| 67 | QC300 | " | 20/11/25 | | | | |
| | WATER | | | | | | |
| 68 | BALL SWI | Water | 21/11/25 | | | | |
| Total 14 | | | | | | | |

| | | | | | | | | | | | | | | | | | | | |
|--|---|--|--|--|--|--|--|--|--|--|--|--|--|--|--|--|--|--|--|
| PFAS Standard PFAS Email Analysis Request send to EnviroLab HOLD | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |
| | X | | | | | | | | | | | | | | | | | | |

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples: Sampler Name: michelle Agnew Signature: mAgnew Date: 21/11/25

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|--|-----------------|-------|
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |
| Name/Signature: | Date: | Carrier / Reference #: | | Name/Signature: | Date: |
| Of: | Time: | Date/Time: | | Of: | Time: |

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Completed by: _____
Checked by: _____

Chain of Custody Documentation

Laboratory: **ALS NSW**
Address:
Contact:
Phone:

Analysis Required

Comments: e.g. Highly contaminate sample; hazardous materials preser LORs etc.

HOLD

| | | | |
|------------------|--------------------------------|-------------------|-------------------|
| Job Number: | C17776 | Purchase Order: | |
| Project Name: | Norfolk DSI | Quote No: | senversa National |
| Sampled By: | MA, HD | Turn Around Time: | std |
| Project Manager: | Michelle Agnew | Page: | 5 of 5 |
| Email Report To: | michelle.agnew@senversa.com.au | Phone/Mobile: | 0448910424 |

| Sample Information | | | | Container Information | | |
|--------------------|-----------------|----------|----------|-----------------------|-------------|---------------|
| Lab ID | Sample ID | Matrix * | Date | Time | Type / Code | Total Bottles |
| 69 | BWC-OMP04-SD | SOIL | 19/11/25 | | | |
| 70 | 1D008-SS1 | | 20/11/25 | | | |
| 71 | 1D008-SS1 | | 20/11/25 | | | |
| 72 | QC100 | | 19/11/25 | | | |
| 73 | QC200 | | " | | | |
| 74 | A-TRUCKFILL-SS1 | | 20/11/25 | | | |
| 75 | A-TRUCKFILL-SS2 | | " | | | |
| 76 | BALL-SS4 | | 21/11/25 | | | |
| 77 | BALL-SS5 | | " | | | |
| 78 | BALL-SS3 | | " | | | |
| 79 | BALL-SD1 | | " | | | |
| Total | | | | | | |

Sampler: I attest that proper field sampling procedures in accordance with Senversa standard procedures and/or project specifications were used during the collection of these samples:
 Sampler Name: **Michelle Agnew** Signature: **MAgnew** Date: **21/11/25**

| Relinquished By: | | Method of Shipment (if applicable): | | Received by: | |
|------------------|-------|-------------------------------------|---------------------------|-----------------------|--|
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: MA | Date: 21/11/25 | |
| Of: | Time: | Date/Time: | Of: MA | Time: 08:45 | |
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: | Date: | |
| Of: | Time: | Date/Time: | Of: | Time: | |
| Name/Signature: | Date: | Carrier / Reference #: | Name/Signature: | Date: | |
| Of: | Time: | Date/Time: | Of: | Time: | |

Water Container Codes: P = Unpreserved Plastic; N = Nitric Acid (HNO₃) Preserved Plastic; ORC = Nitric Preserved ORC; SH = Sodium Hydroxide (NaOH)/Cadmium (Cd) Preserved; S = Sodium Hydroxide Preserved Plastic; STH = Sodium thiosulfate preserved plastic; V = VOA Vial Hydrochloric Acid (HCl) Preserved; VS = VOA Vial Sulphuric Preserved; VSA = Sulphuric Preserved Amber Glass; H = HCl Preserved Plastic; HS = HCl Preserved Speciation Bottle; SP = Sulphuric Preserved Plastic; F = Formaldehyde Preserved Glass; Z = Zinc Acetate Preserved Bottle; E = EDTA Preserved Bottles; ST = Sterile Bottle; UA = Unpreserved Amber Glass; L=Lugol's iodine preserved white plastic bottle; SW= sulfuric acid preserved wide mouth glass jar

Completed by:
Checked by:



Appendix E: Mann Kendall Analysis

| Location Code | Monitoring Zone | Perfluorooctanoic acid (PFOA) µg/L | Perfluorooctanesulfonic acid (PFOS) µg/L | Sum of PFHxS and PFOS µg/L |
|-------------------|-----------------|---------------------------------------|---|-------------------------------|
| AIRPORT_BORE | Mission Creek | TDn, S=-10, N=6, P=0.0426 | TDn, S=-13, N=6, P=0.0121 | TDn, S=-13, N=6, P=0.0121 |
| WWII_DAM | Mission Creek | TDn, S=-11, N=6, P=0.0301 | TDn, S=-11, N=6, P=0.0301 | TDn, S=-11, N=6, P=0.0301 |
| Cockpit_SW01 | Cascade Creek | NT, S=0, N=6, P=0.5000 | NT, S=-5, N=6, P=0.2178 | NT, S=-7, N=6, P=0.1212 |
| ID013_BORE | Mission Creek | NT, S=0, N=4, P=0.5000 | NT, S=0, N=4, P=0.5000 | NT, S=0, N=4, P=0.5000 |
| ID014_BORE | Mission Creek | NT, LH, S=8, N=6, P=0.0903 | NT, LH, S=7, N=6, P=0.1298 | NT, S=5, N=6, P=0.2262 |
| ID015_BORE | Mission Creek | NT, S=-2, N=5, P=0.3970 | NT, S=-2, N=5, P=0.4032 | NT, S=-4, N=5, P=0.2312 |
| ID016_BORE | Mission Creek | NT, S=0, N=5, P=0.5000 | NT, S=0, N=5, P=0.5000 | NT, S=0, N=5, P=0.5000 |
| MC_OMP01 | Mission Creek | NT, S=-6, N=5, P=0.1103 | NT, S=-6, N=5, P=0.1103 | TDn, S=-8, N=5, P=0.0432 |
| MC_OMP02 | Mission Creek | NT, S=6, N=5, P=0.0958 | NT, S=3, N=5, P=0.3067 | NT, S=4, N=5, P=0.2312 |
| MC_OMP03 | Mission Creek | NT, S=-2, N=5, P=0.4032 | NT, S=-2, N=5, P=0.4032 | NT, S=-2, N=5, P=0.4032 |
| MC_OMP04 | Mission Creek | NT, S=-4, N=4, P=0.1541 | NT, S=-2, N=4, P=0.3670 | NT, S=-2, N=4, P=0.3670 |
| MC_OMP05 | Mission Creek | NT, S=-5, N=4, P=0.0743 | NT, S=0, N=4, P=0.5000 | NT, S=-2, N=4, P=0.3670 |
| MC_OMP06 | Mission Creek | NT, S=-5, N=5, P=0.1561 | NT, S=-2, N=5, P=0.4032 | NT, S=-4, N=5, P=0.2312 |
| MC_OMP07 | Mission Creek | TDn, S=-6, N=4, P=0.0447 | NT, S=2, N=4, P=0.3670 | NT, S=2, N=4, P=0.3670 |
| MC_OMP08 | Mission Creek | NT, S=0, N=4, P=0.5000 | NT, S=0, N=4, P=0.5000 | NT, S=0, N=4, P=0.5000 |
| MC_OMP09 | Mission Creek | NT, S=5, N=5, P=0.1561 | NT, S=4, N=5, P=0.2312 | NT, S=4, N=5, P=0.2312 |
| MC_OMP10 | Mission Creek | NT, S=-5, N=4, P=0.0743 | NT, S=-4, N=4, P=0.1541 | NT, S=-4, N=4, P=0.1541 |
| MC_OMP11 | Mission Creek | NT, S=0, N=4, P=0.5000 | NT, S=-2, N=4, P=0.3670 | NT, S=-2, N=4, P=0.3670 |
| PWS_HEAD_DAM | Headstone Creek | NT, S=0, N=6, P=0.5000 | NT, S=-5, N=6, P=0.1208 | NT, S=-5, N=6, P=0.1208 |
| WC_OMP01 | Watermill Creek | NT, S=2, N=5, P=0.3864 | NT, S=-1, N=5, P=0.5000 | NT, S=0, N=5, P=0.5000 |
| WC_OMP02 | Watermill Creek | NT, S=-4, N=5, P=0.1444 | NT, S=-1, N=5, P=0.5000 | NT, S=-3, N=5, P=0.3067 |
| WC_OMP03 | Watermill Creek | NT, S=0, N=6, P=0.5000 | NT, S=-3, N=6, P=0.3484 | NT, S=-3, N=6, P=0.3536 |
| WC_OMP04_DUCK DAM | Watermill Creek | NT, S=0, N=6, P=0.5000 | NT, S=6, N=6, P=0.1570 | NT, LH, S=4, N=6, P=0.2830 |
| WC_OMP05 | Watermill Creek | NT, S=0, N=5, P=0.5000 | NT, S=1, N=5, P=0.5000 | NT, S=1, N=5, P=0.5000 |

Legend

S: Mann-Kendall score
N: number of samples
P: p-value

| | |
|-----|--------------------------------|
| NT | No trend |
| TDn | Downward trend |
| LH | Latest result highest recorded |

Analysed in Esdat version 5.1.26042.0, significance level 0.05 (95% confidence)

Senversa Pty Ltd

ABN 89 132 231 380

www.senversa.com.au

enquiries@senversa.com.au

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Certified



Corporation

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